Original research

Mendelian randomisation of eosinophils and other cell types in relation to lung function and disease

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ABSTRACT

Rationale Eosinophils are associated with airway inflammation in respiratory disease. Eosinophil production and survival is controlled partly by interleukin-5: anti-interleukin-5 agents reduce asthma and response correlates with baseline eosinophil counts. However, whether raised eosinophils are causally related to chronic obstructive pulmonary disease (COPD) and other respiratory phenotypes is not well understood.

Objectives We investigated causality between eosinophils and: lung function, acute exacerbations of COPD, asthma-COPD overlap (ACO), moderate-to-severe asthma and respiratory infections.

Methods We performed Mendelian randomisation (MR) using 151 variants from genome-wide association studies of blood eosinophils in UK Biobank/INTERVAL, and respiratory traits in UK Biobank/SpiroMeta, using methods relying on different assumptions for validity. We performed multivariable analyses using eight cell types where there was possible evidence of causation by eosinophils.

Measurements and main results Causal estimates derived from individual variants were highly heterogeneous, which may arise from pleiotropy. The average effect of raising eosinophils was to increase risk of ACO (weighted median OR per SD eosinophils, 1.44 (95% CI 1.19 to 1.74)), and moderate-severe asthma (weighted median OR 1.50 (95% CI 1.23 to 1.83)), and to reduce forced expiratory volume in 1 s (FEV1) forced vital capacity (FVC) and FEV1/FVC (weighted median estimator, SD FEV1/FVC: −0.054 (95% CI −0.078 to −0.029), effect only prominent in individuals with asthma).

Conclusions Broad consistency across MR methods may suggest causation by eosinophils (although of uncertain magnitude), yet heterogeneity necessitates caution: other important mechanisms may be responsible for the impairment of respiratory health by these eosinophil-raising variants. These results could suggest that anti-IL5 agents (designed to lower eosinophils) may be valuable in treating other respiratory conditions, including people with overlapping features of asthma and COPD.

INTRODUCTION

Eosinophils are proinflammatory granulocytes associated with symptom severity and exacerbation frequency in asthma and chronic obstructive pulmonary disease (COPD).1–3 The degree of eosinophilia (raised eosinophils) in these obstructive lung diseases varies: while eosinophil inflammation due to allergic sensitisation has been considered characteristic of asthma, not all patients with asthma have eosinophilia.1–4 Moreover, while airway inflammation in COPD is typically mediated by neutrophils, some individuals with COPD have raised eosinophils.1–5

The production and survival of eosinophils is partly regulated by interleukin-5 (IL-5), and anti-IL5 therapies (eg, mepolizumab, reslizumab, and the anti-IL5Rα agent, benralizumab) are now used in asthma and COPD overlap (ACO) to treat asthma. However, the causal nature of the relationship between eosinophils and a broad range of respiratory traits related to asthma and chronic obstructive pulmonary disease (COPD) is not fully understood.

What this study adds?

⇒ In this Mendelian randomisation study, while the average effect of raising eosinophils was to increase risk of asthma-COPD overlap and asthma, and worsen forced expiratory volume in 1 s (FEV1) and FEV1/FVC forced vital capacity in individuals with asthma, heterogeneity of individual causal estimates means caution is needed when interpreting these results causally, as these results could also be consistent with eosinophil-raising genetic variants impairing respiratory health via other causal pathways.

How this study might affect research, practice or policy?

⇒ These results could suggest that anti-IL5 agents (designed to lower eosinophils) may be valuable in treating other respiratory conditions, including people overlapping features of both asthma and COPD. Future work should seek to explore other potential mechanisms besides eosinophils by which anti-IL5 agents may improve respiratory health, to inform whether the clinical indications for anti-IL5 agents or biomarkers for stratifying their use could be extended.

Key messages

What is already known on this topic?

⇒ Blood eosinophil counts are predictive of response to anti-interleukin-5 (IL5) drugs used to treat asthma. However, the causal nature of the relationship between eosinophils and a broad range of respiratory traits related to asthma and chronic obstructive pulmonary disease (COPD) is not fully understood.

What this study adds?

⇒ In this Mendelian randomisation study, while the average effect of raising eosinophils was to increase risk of asthma-COPD overlap and asthma, and worsen forced expiratory volume in 1 s (FEV1) and FEV1/FVC forced vital capacity in individuals with asthma, heterogeneity of individual causal estimates means caution is needed when interpreting these results causally, as these results could also be consistent with eosinophil-raising genetic variants impairing respiratory health via other causal pathways.

How this study might affect research, practice or policy?

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licensed in many countries for the treatment of severe eosinophilic asthma.6–12 The decision to treat asthma with these drugs is currently based on blood eosinophil count, among other factors,1 since post-hoc analyses of clinical trials stratified by eosinophil levels have shown increased efficacy of mepolizumab for treating severe asthma in those with higher baseline eosinophils.5 Results from Mendelian randomisation (MR) analyses have also provided evidence for a role of eosinophils in asthma (estimated OR 1.70 (95% CI 1.53 to 1.91)).13 MR analyses use genetic variants as instrumental variables (IVs) to investigate causality between exposure and outcome, and under certain assumptions may obviate problems with traditional observational epidemiology (eg, reverse causation, confounding), permitting causal inference.

In addition to asthma, blood eosinophils are associated with quantitative lung function in general populations (ie, including individuals without asthma).14 However, causality has yet to be established: an inverse relationship between eosinophils and lung health has been suggested, yet a previous MR of lung function (plus another including asthma and COPD) were of small sample size, with imprecise estimates precluding confident inference.15 16 Moreover, causality of eosinophils on other respiratory phenotypes, for example, asthma-COPD overlap (ACO), and respiratory infections are yet to be investigated. COPD is diagnosed by spirometry if the ratio of the forced expiratory volume in 1 s (FEV1) to forced vital capacity (FVC), FEV1/FVC, is <0.7, with airflow obstruction graded by predicted FEV1. Therefore, studying eosinophils as determinants of quantitative lung function is a powerful way of understanding their role in the development of fixed airflow obstruction such as in COPD.17 18 Investigating causality between eosinophils and fixed airflow obstruction is pertinent given interest in the potential use of mepolizumab in COPD9–12; evidence for causality of eosinophils in a wider range of respiratory phenotypes could suggest that anti-IL5 agents (designed to lower eosinophils) might be helpful in conditions beyond asthma.

We undertook two-sample MR analyses using summary-level genome-wide association study (GWAS) data to assess causality between eosinophils and conditions encompassing fixed and reversible airflow obstruction, using genetic variants associated with blood eosinophils as IVs.13 We investigated causality of eosinophils on three quantitative lung function spirometry traits, and four clinical phenotypes (moderate-to-severe asthma, acute exacerbations of COPD (AECOPD), ACO and respiratory infections). We used MR approaches relying on different assumptions for validity, and followed up traits showing evidence of possible causality to assess evidence that the IVs affected lung function via eosinophil counts and not via other blood cell types. Overall, our aim was to provide a comprehensive assessment of the causal role of blood eosinophil counts in relation to respiratory health and disease.

**METHODS**

We assessed causality between eosinophils and other blood cell counts in relation to respiratory outcomes using MR.19 20 MR involves using genetic variants (here single-nucleotide polymorphisms, SNPs), as IVs for an exposure of interest, in this case eosinophil counts, by comparing the magnitude of the effect of the SNPs on the outcome to the effect of the SNPs on the exposure.19 20 All analyses reported are two-sample MR analyses, since SNP-exposure and SNP-outcome associations were extracted from different (yet overlapping) samples. Core MR assumptions for inferring causality between are that: (1) the genetic variants are associated with the exposure of interest; (2) there are no unmeasured confounders of the associations between genetic variants and outcome; and (3) the genetic variants affect the outcome only via the exposure of interest (figure 1).19 Additional assumptions for accurate point estimation of effect sizes are discussed in online supplemental file 1, and elsewhere.22

All GWAS datasets analysed included UK Biobank, a prospective cohort study including spirometry, biological assays, questionnaire data, and linked healthcare records, and 450 000 participants with genotype data.23 Other studies were incorporated where available, and all GWAS data were from individuals of European ancestry. Datasets are summarised below, and descriptions of covariate adjustments, and exposure-outcome GWAS overlap are given in the extended methods (online supplemental file 1).

**Exposure GWAS data sets (blood cell parameters)**

We used summary-level data from eight published GWASs of blood cell counts13 in the initial release of UK Biobank genetic data (N up to 132,959, that is, around 30% of participants with genotype data), plus the INTERVAL study (N up to 40 521).13 GWASs were of blood eosinophils, basophils, neutrophils,
monocytes, lymphocytes, platelets, red blood cells and reticulocytes, with adjustments for technical and seasonal covariates, plus age, menopausal status, height, weight, smoking and alcohol (online supplemental file 1).

Outcome GWAS data sets (respiratory outcomes)
See also online supplemental file 1.

Quantitative lung function GWAs
We used published summary-level data from three GWAS of FEV1, FVC and FEV1/FVC, in UK Biobank (n=321 047) and the SpiroMeta consortium (n=79 055).19 Prior to GWAS, traits were preadjusted for age, age^2, sex, height, smoking status and other covariates as appropriate, for example, ancestry principal components. Residuals were inverse-normal rank transformed.

Clinical outcome GWAS
Moderate-to-severe asthma
We used a published GWAS of moderate-to-severe asthma within the Genetics of Asthma Severity and Phenotypes initiative, the U-BIOPRED asthma cohort, and UK Biobank.20 Cases (n=5135) were taking asthma medication, and met criteria for moderate-to-severe asthma (British Thoracic Society 2014 guidelines). Controls (n=25 675) excluded those with a doctor diagnosis of asthma, rhinitis, eczema, allergy, emphysema, or chronic bronchitis, or missing medication data. Analyses were adjusted for 10 ancestry principal components.

Acute exacerbations of COPD
We defined AECOPD in UK Biobank; the eligible sample was restricted to individuals with FEV1/FVC<0.7. Exacerbation cases (n=2771) had an ICD-10 code for AECOPD or a lower respiratory tract infection in Hospital Episode Statistics data (online supplemental table 1). Controls (n=42 052) had FEV1/FVC>0.7, without an AECOPD code. Associations were adjusted for age (at recruitment), age^2, sex, smoking status (ever/never), genotyping array and 10 principal components.

Asthma-COPD overlap
We defined ACO in UK Biobank (N=8068) as individuals self-reporting a doctor diagnosis of asthma, with FEV1/FVC<0.7 and FEV1<80% predicted at any study visit. Controls (N=40 360) were selected in approximately a 5:1 ratio, from participants reporting no asthma or COPD, (FEV1>80% predicted, FEV1/FVC>0.7). Associations were adjusted for age (at recruitment), sex, smoking status and 10 principal components.21

Respiratory infections
We defined respiratory tract infections requiring hospital admission in UK Biobank, using the ICD-10 codes in online supplemental table 2. Cases had ≥1 admission for respiratory infections (N=19 459). Controls had no admissions for respiratory infections and were selected in approximately a 5:1 ratio (N=101 438). Associations were adjusted for age (at recruitment), age^2, sex, smoking status, genotyping array, and 10 principal components.22

Statistical methods
Univariable MR of eosinophils and respiratory traits and diseases
We performed separate MR analyses of eosinophils on three quantitative lung function traits (FEV1, FVC, FEV1/FVC); and four clinical phenotypes (asthma, AECOPD, ACO, respiratory infections) using genetic IVs from the work of Astle and colleagues.13 Selection of 151 eosinophil IVs and harmonisation of SNP-exposure and SNP-outcome datasets is detailed in the online supplemental file 1. The primary MR analysis used the inverse-variance weighted (IVW) method and a random-effects model, which will return a valid causal estimate provided that the average pleiotropic effect is zero. We investigated the ‘no pleiotropy’ assumption using MR-Egger regression,27 the weighted median estimator28 and MR-PRESSO29 (see online supplemental file 1 for details on assumptions relied on for validity by each method). Further sensitivity analyses: (1) investigated robustness of findings to heterogeneity using MR-PRESSO (for traits with some evidence of causation by eosinophils), (2) restricted to non-UKB FEV1/FVC GWAS data, to assess sensitivity to sample overlap and (3) restricted to FEV1/FVC GWAS data in UKB, stratifying by asthma status.

Multivariable MR analyses of multiple blood cell types and respiratory outcomes
Since SNPs affecting eosinophils also affect other blood cell types,13 we used multivariable MR to estimate the influence of multiple cell types on respiratory outcomes, after conditioning on the effects of the SNPs on other cell types. Multivariable MR analyses were performed for respiratory outcomes with evidence of eosinophil causation in the IVW MR analyses above, and with broadly consistent effect estimates in the weighted median and MR-Egger analyses. We also performed an analysis of FEV1/FVC (stratifying by asthma status).

There were 1166 SNPs associated with at least one of eight blood traits reported by Astle and colleagues13 at a genome-wide threshold. These SNPs were LD clumped, and effect sizes extracted from each blood cell GWAS, and each outcome GWAS. Effects for 318 clumped SNPs were harmonised, that is, so effect sizes for SNP-exposure and SNP-outcome effects corresponded to the same allele (online supplemental table 3, online supplemental file 1). Conditional F-statistics were estimated using the strength_mvmr() function of the ‘MVMR’ R package.30

For IVW multivariable MR analyses, we used the mv_multiple() function of the ‘TwoSampleMR’ R package.31,32 This analysis aimed to further investigate the possibility of horizontal pleiotropy affecting the results of the univariable eosinophil MR; and to establish whether other blood cell types besides eosinophils could affect the respiratory outcomes studied.

Sensitivity MVMR methods (online supplemental file 1) included: (1) use of an MVMR method more robust to pleiotropy in the presence of weak instruments (using the qhet_mvmr() function of the ‘MVMR’ R package)—standard errors calculated by a jackknife approach and (2) recalculation of IVW MVMR estimates after removal of SNPs contributing most to heterogeneity (SNPs identified using the pleiotropy_mvmr() function).

RESULTS
Univariable MR analyses of eosinophils and respiratory outcomes
There were 151 SNPs available for the univariable MR analyses of three quantitative traits (FEV1, FVC and FEV1/FVC), and four respiratory disease phenotypes (moderate-to-severe asthma, AECOPD, ACO and respiratory infections). Details of SNP selection are described in figure 2.

Results are presented in figure 2. Among the quantitative traits, there was evidence for an effect of eosinophils on FEV1/FVC (SD change in FEV1/FVC per SD eosinophils, IVW estimate=−0.049
209 SNPs are reported as eosinophil signals in Astle et al. (2016)

167 SNPs available in lung function GWAS (Shrine et al. 2019)

Proxies at $r^2>0.8$ retrieved for 24/42 remaining SNPs

191 SNPs available for use as instrumental variables

151* SNPs after harmonising SNP-exposure and SNP-outcome effects, and removing correlated variants

Outcomes studied:
- Quantitative traits: FEV$_1$, FVC, FEV$_1$/FVC
- Respiratory disease phenotypes: asthma, AECOPD, ACO, respiratory infections

Figure 2  Selection of SNPs for univariable MR analyses of eosinophils and respiratory outcomes flow chart describing the analysis workflow for initial MR analyses of eosinophils. Of 209 SNPs associated with eosinophil count, 167 were available in lung function GWASs (missingness is due to some SpiroMeta studies not being imputed to the HRC panel). LD proxies at $r^2>0.8$ were retrieved for 24/42 missing variants. Of the resulting 191 SNPs, 188 were successfully harmonised between the SNP-eosinophil and SNP-lung function data sets, and 151* remained after LD clumping at an $R^2$ threshold of 0.01. These 151 SNPs were used in analyses. *One SNP, rs9974367, was missing in the moderate-severe asthma GWAS, AECOPD, acute exacerbation of COPD; ACO, asthma COPD overlap; COPD, chronic obstructive pulmonary disease; FEV$_1$, forced expiratory volume in 1 s, FVC, forced vital capacity; GWAS, genome-wide association study; MR, Mendelian randomisation; SNPs, single-nucleotide polymorphisms.

Sensitivity analysis to assess the effects of sample overlap for quantitative lung function traits

UK Biobank featured in all GWAS datasets used, although the blood cell count GWAS and asthma GWAS included only approximately one third of the UK Biobank genotype data. We conducted sensitivity analyses to assess for the effect of sample overlap, since we had access to quantitative lung function GWAS data without UK Biobank participants (see online supplemental file 1). Results were generally consistent (SD change in FEV$_1$/FVC per SD eosinophil count, IVW estimate = $-0.041$ (95% CI $-0.072$ to $-0.009$); SD change FEV$_1$, per SD eosinophil count = $-0.043$ (95% CI $-0.077$ to $-0.010$)) (online supplemental table 6).

Sensitivity analysis to assess the effect on FEV$_1$/FVC in individuals with and without asthma

The causal effect of eosinophils on FEV$_1$/FVC was recalculated using data from UK Biobank, stratifying by asthma status (37868 cases, 283179 controls). The effect size was larger in individuals with asthma (IVW $-0.083$ (95% CI $-0.139$ to $-0.028$)) than in those without asthma, in whom there was no effect (IVW $-0.013$ (95% CI $-0.044$ to $0.014$)). However, confidence intervals for both subgroups overlapped one another (see online supplemental table 7).

Multivariable MR analyses of blood cell counts and respiratory outcomes

To further explore causality between blood cell parameters and FEV$_1$/FVC, moderate-to-severe asthma and ACO, and to see if other exposures could have accounted for the heterogeneity observed in the previous analyses, we carried out multivariable MR analyses, using eight cell type exposures (eosinophils, basophils, neutrophils, monocytes, lymphocytes, platelets, red blood cells and reticulocytes).

Selection of 318 SNP IVs for multivariable MR is described in online supplemental file 1, online supplemental table 3. SNPs used in the univariable and multivariable MR are listed in online supplemental tables 8 and 9. Briefly, 1166 unique SNPs were associated with at least one of the eight cell types at a genome-wide level in
Respiratory epidemiology

Table 1: Results of MR analyses for ACO and asthma

<table>
<thead>
<tr>
<th>Trait</th>
<th>N</th>
<th>Method</th>
<th>OR_95CI</th>
<th>P</th>
<th>Q</th>
<th>Qdf</th>
<th>Intercept_P</th>
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<tbody>
<tr>
<td>Asthma</td>
<td>5135/25675</td>
<td>IVW</td>
<td>2.45 (1.98, 3.00)</td>
<td>5.6e-16</td>
<td>662</td>
<td>149</td>
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<tr>
<td></td>
<td></td>
<td>MR Egger</td>
<td>1.57 (0.93, 2.65)</td>
<td>0.092</td>
<td>648</td>
<td>148</td>
<td>0.067</td>
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<td>Weighted median</td>
<td>1.5 (1.23, 1.83)</td>
<td>7.38e-05</td>
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<td>AECOPD</td>
<td>2771/42052</td>
<td>IVW</td>
<td>1.03 (0.9, 1.18)</td>
<td>0.637</td>
<td>143</td>
<td>150</td>
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<td></td>
<td></td>
<td>MR Egger</td>
<td>0.86 (0.63, 1.18)</td>
<td>0.355</td>
<td>141</td>
<td>149</td>
<td>0.219</td>
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<td>Weighted median</td>
<td>1.11 (0.89, 1.37)</td>
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<td>ACO</td>
<td>8068/40360</td>
<td>IVW</td>
<td>1.69 (1.52, 2.27)</td>
<td>5.97e-10</td>
<td>777</td>
<td>150</td>
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<tr>
<td></td>
<td></td>
<td>MR Egger</td>
<td>1.37 (0.85, 2.21)</td>
<td>0.195</td>
<td>768</td>
<td>149</td>
<td>0.174</td>
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<tr>
<td>Weighted median</td>
<td>1.44 (1.19, 1.74)</td>
<td>0.000175</td>
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<tr>
<td>Resp. IX</td>
<td>19459/101430</td>
<td>IVW</td>
<td>1.02 (0.96, 1.08)</td>
<td>0.529</td>
<td>176</td>
<td>150</td>
<td></td>
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<tr>
<td></td>
<td></td>
<td>MR Egger</td>
<td>0.9 (0.79, 1.03)</td>
<td>0.14</td>
<td>172</td>
<td>149</td>
<td>0.057</td>
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<tr>
<td>Weighted median</td>
<td>0.97 (0.89, 1.06)</td>
<td>0.463</td>
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Figure 3: MR analyses of eosinophils (exposure) on three quantitative lung function traits (top) and four respiratory disease phenotypes (bottom), using 151 eosinophil-associated SNPs. Top: Results of MR analyses of eosinophil counts (exposure) on three quantitative lung function traits (outcome), FEV1, FVC and FEV1/FVC. A forest plot of three estimates for each trait is shown (IVW, MR Egger, weighted median), along with the maximum sample size in the outcome GWAS (N), the effect size in SD change in outcome trait per SD increase eosinophil count, and 95% CI, values for Cochran’s Q statistic (Q) and the associated df (Q, df), and the p value for the MR Egger intercept (Intercept_P). Boxes of the forest plot represent effect sizes, whiskers are 95% CIs. Bottom: Results of MR analyses of eosinophil counts (exposure) on four respiratory disease phenotypes (outcome), moderate-to-severe asthma, acute exacerbations of COPD (AECOPD), asthma-COPD overlap (ACO), and respiratory infection (Resp. IX). A forest plot of three estimates for each trait is shown (IVW, MR Egger, weighted median), along with sample size in the outcome GWAS for cases and controls, respectively (N), the effect size as OR per SD eosinophil count, and 95% CI, values for Cochran’s Q statistic (Q) and the associated df (Q, df), and the p value for the MR Egger intercept (Intercept_P). Boxes of the forest plot represent ORs, whiskers are 95% CIs. NB only 150/151 of the eosinophil SNPs were available in the moderate-to-severe asthma GWAS. COPD, chronic obstructive pulmonary disease; FEV1, forced expiratory volume in 1 s; FVC, forced vital capacity; GWAS, genome-wide association study; IVW, inverse-variance weighted; MR, Mendelian randomisation; SNPs, single-nucleotide polymorphisms.

Sensitivity multivariable MR analyses

Sensitivity MVMR analyses (1) used an estimation technique more robust to balanced pleiotropy and (2) repeated IVW MVMR, omitting SNP IVs with the most evidence of heterogeneity. Effect directions of sensitivity analyses and the main on lung function being of greater magnitude in people with a history of asthma (online supplemental figure 3).

Results of the multivariable MR analysis for ACO and asthma are presented in figure 5. There was an association of eosinophil count with both ACO (OR 1.95 (95% CI 1.57 to 2.42)) and asthma (OR 2.90 (95% CI 2.31 to 3.65)), after adjusting for the effects of the SNPs on other cell types. Confidence intervals for other cell type estimates were consistent with the null, with the exception of neutrophils for ACO. None of the additional seven cell types showed strong evidence of causality.

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Results of the multivariable MR analysis for ACO and asthma are presented in figure 5. There was an association of eosinophil count with both ACO (OR 1.95 (95% CI 1.57 to 2.42)) and asthma (OR 2.90 (95% CI 2.31 to 3.65)), after adjusting for the effects of the SNPs on other cell types. Confidence intervals for other cell type estimates were consistent with the null, with the exception of neutrophils for ACO. None of the additional seven cell types showed strong evidence of causality.

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and asthma. However, CIs for FEV1 and FEV1/FVC were broad, and asthma respectively: this could be because our SNP-95% CIs. ACO, asthma-
of other cell types). Points of the forest plot represent ORs; whiskers are—comparison. ORs (95% CI) are per SD cell count (adjusted for the effects
sizes are shown next to one another for comparison. Effect sizes (beta,
run for each of FEV1 and the ratio of FEV1 to FVC separately, but effect
conditioning on the effects of the SNPs on other cell types. While
inclusion of genetic correlation with eosinophils), but also, around half of
s (FEV1) and FEV1/forced vital capacity (FVC) forest plot showing multivariable MR estimating the causal effect of multiple cell types on quantitative lung function outcomes, after conditioning on the effects of the SNPs on other cell types. Models were run for each of FEV1, and the ratio of FEV1 to FVC separately, but effect sizes are shown next to one another for comparison. Effect sizes (beta, 95% CI) are in SD change in lung function outcome per SD cell count (adjusted for the effects of other cell types). Points of the forest plot represent effect size estimate; whiskers are 95% CIs. MR, Mendelian randomisation.

MVMR analyses were concordant for FEV1, FEV1/FVC, ACO, and asthma. However, CIs for FEV1, and FEV1/FVC were broad, and overlapped the null. For ACO and asthma estimates, there was still evidence of an effect, although attenuated in both analyses (estimates from analysis more robust to pleiotropy; ACO OR 1.57 (95% CI 1.07 to 2.30); asthma OR 2.66 (95% CI 1.65 to 4.33); estimates after omitting the most heterogeneous SNPs: ACO OR 1.51 (95% CI 1.23 to 1.85); asthma OR 2.29 (95% CI 1.84 to 2.86)).

DISCUSSION

In MR analyses, we found that the average effect of raising eosinophils was to decrease FEV1/FVC and FEV1, and to increase ACO and asthma risk, and there was broad consistency across MR methods. However, causal estimates of individual variants were highly heterogeneous, suggesting that caution is needed in concluding causal inference: some IVs may have violated MR assumptions, and other important genetically correlated mechanisms could be responsible for the effect on lung health and disease by the eosinophil-raising variants studied.

To our knowledge, this is the largest MR of eosinophils and lung function, and the first to investigate eosinophils and AECOPD, ACO and respiratory infections. Terminology of ACO has changed over time, yet recognition that asthma and COPD coexist in some patients has not changed and this is what our analysis aimed to capture.

A previous two-sample MR of eosinophils and asthma was undertaken by the authors of the GWAS that discovered the eosinophil IVs used; this MR analysis used asthma GWAS data from the GABRIEL study. We are aware of one other small MR of eosinophils and asthma, COPD, FEV1 and FEV1/FVC, conducted in the LifeLines cohort (N=13,301, 5 SNPs IVs). In that study, CIs for causal estimates of eosinophils overlapped the null, although point estimates were consistent with a harmful effect for FEV1/FVC, asthma and COPD. We used a larger eosinophil GWAS (N=172,275) to derive IVs, and found that the average effect of eosinophil-raising IVs was to reduce FEV1/FVC, the trait used in COPD diagnosis and FEV1, used to grade COPD airflow limitation. However, sensitivity analyses highlighted a larger causal estimate of eosinophils on FEV1/FVC among those with asthma, with effect estimates attenuating when excluding this group. These findings may highlight the importance of eosinophils as a marker of impaired lung function and airflow obstruction in people with a history of asthma.

We highlight a need for caution in inferring simple causation between eosinophils and these phenotypes, since high degrees of heterogeneity in our results may arise from pleiotropy. To investigate, we compared MR methods relying on differing assumptions for validity (Methods section). Attenuation of some results when using the MR-Egger, weighted median, and MR-PRESSO approaches suggests that some SNP IVs are associated with asthma and ACO via pathways other than eosinophils, which is a known challenge in MR studies (see also Methods section).

Since many of the eosinophil SNP IVs are also associated with other cell counts, we performed multivariable MR to estimate the influence of multiple cell types simultaneously, after conditioning on the effects of the SNPs on other cell types. While we did not find substantial evidence for a harmful effect of neutrophils on asthma, nor a protective effect of monocytes and lymphocytes, as reported previously, effect directions in our IVW multivariable MR were consistent with the previous study for neutrophils, monocytes and lymphocytes. We observed a larger effect of eosinophils on asthma than reported previously: this could be because our SNP-outcome dataset was of moderate-to-severe asthma (which has a higher point estimate of genetic correlation with eosinophils), but also, around half of the cases and the majority of controls were also included in the
exposure GWAS, which may make this analysis closer to a one-sample MR, and inflate causal effect estimates. Notably, effect sizes partly attenuated in sensitivity analyses which may be more robust to heterogeneity. The MR estimates from multivariable analyses, and the MR-Egger regression and weighted median univariable analyses were consistent with the previous estimate reported for asthma in multivariable analysis by Astle et al. Nevertheless, these limitations may preclude precise estimation of effect sizes, and our results may be more useful in terms of assessing whether there is causality between eosinophils and the phenotypes studied, as opposed to providing estimates of the magnitude of any causal effect between phenotypes.

While we did not find strong evidence for causality of eosinophils on AECOPD and respiratory infections, point estimates were consistent with a harmful effect on AECOPD, and may have been limited by power. The effects of anti-IL5 drugs that have been attributed to the reduction of eosinophils have been noted to be smaller in AECOPD compared with asthma. Key strengths are that we used MR methods with differing sensitivities to underlying assumptions. We a large GWAS of eosinophil counts, to provide a comprehensive assessment of the role of blood eosinophils in relation to multiple respiratory health and disease outcomes. Another strength is that we undertook multivariable MR to investigate causality between multiple cell types and the outcomes studied, while controlling for the effects of IVs that may have had pleiotropic effects via other cell types.

We acknowledge several limitations. We did not have postbronchodilator measures of spirometry. We used GOLD Stage 2–4 COPD (prebronchodilator FEV₁ <80% predicted) when defining ACO; using the same prebronchodilator spirometry definition of COPD, a positive predictive value of 98% for diagnosis of postbronchodilation-defined COPD has been shown. Sample overlap between the SNP-eosinophil and SNP-outcome datasets (all included participants from UK Biobank) could bias estimates towards the observational eosinophil-outcome association; we repeated the univariable MR analysis of eosinophils using SNP-lung function results excluding UK Biobank participants, and observed a consistent IVW estimate. Nevertheless, our other analyses (particularly the asthma analysis) could be vulnerable to some non-conservative bias. GWAS analyses of cell counts have, since analysis, been extended to a larger sample across UKB, and future work deriving IVs from this study would be valuable. UK BiEVE participants (a subset of UK Biobank selected for extremes of respiratory traits), were overrepresented in Astle et al, which used the interim release of UKB data. While correlation between effect sizes from the two GWAS for the 151 IVs used in these analyses had high, the possibility of selection effects remains. Our MR analyses also use genome-wide results adjusted for covariates, and therefore may be susceptible to collider bias. There is also potential bias in the causal estimates for binary outcomes due to non-collapsibility of the OR, and we did not consider the possibility of non-linear effects. The multivariable analyses may still be vulnerable to pleiotropy via pathways other than the eight cell types studied, so while we cannot strongly assert causality of eosinophils on lung function, neither do we rule it out, as our results are consistent with a causal effect.

At present, treatment with anti-IL5/anti-IL5Rα agents in asthma is initiated according to eosinophil counts and other factors, yet it is possible that a more proximal factor may be an even better predictor of drug response. Future work could seek therefore to identify whether particular pathways upstream of eosinophil counts might help design better methods for deciding on treatment initiation. In addition, use of suitable IVs for IL5 levels would permit two-step MR analyses, assessing for a mediating effect of eosinophils on the action of anti-IL5 agents in reducing respiratory morbidity.

To conclude, using MR, we found that the average effect of raising eosinophils was to increase risk of ACO and asthma, and to reduce FEV₁/FVC (the latter association was only prominent in individuals with asthma). Broad consistency across MR methods is suggestive of a causal effect of eosinophils on asthma overall, and in individuals with features of both asthma and fixed airflow obstruction, although of uncertain magnitude. However, given heterogeneity in results derived from individual IVs, which may indicate violation of MR assumptions, we highlight a need for caution, since alternative mechanisms may be responsible for the impairment of respiratory health by these eosinophil-raising variants. These results could suggest that anti-IL5 agents (designed to lower eosinophils) may be of value in a wider range of respiratory traits, including people with features of both asthma and COPD. Future work should seek to explore other potential mechanisms besides eosinophils by which anti-IL5 agents may improve respiratory health.

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Contributors AG, NuS, FD and MDT contributed to the conception and design of the study. AG undertook data analysis and produced the first draft of the manuscript, and revised it with CI, IH, LW, NuS, FD and MDT. AG, CJ, ATW, NIS, NRF, SpiroMeta consortium, IS, IH, LW and MDT contributed to data acquisition. AG, CJ, IS, IH, LW, NuS, FD and MDT contributed to data interpretation. All authors critically reviewed the manuscript before submission. AG and MDT act as guarantors for the content of the manuscript.

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Respiratory epidemiology

Summary-level lung function GWAS and blood cell GWAS data are available from https://www.ebi.ac.uk/gwas/.

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Extended Methods

Further details on the published GWAS datasets included in this analysis are described below (see also original publication references, which are given for each study).

See also Supplementary References listed at the end of this document.

Exposure GWAS data sets (blood cell parameters) (Astle et al. 2016)¹

Summary-level data from a GWAS of blood cell count parameters¹ undertaken in the interim release of UK Biobank (UKB) and INTERVAL studies (N=172,275) were downloaded from the EBI GWAS Catalog (https://www.ebi.ac.uk/gwas/). We first used the results for eosinophil counts in MR analyses of lung function and respiratory disease. We then extended our analysis to simultaneously model the causal effects of additional cell types (e.g. counts of basophils, neutrophils, monocytes, lymphocytes, platelets, red blood cells and reticulocytes) in multivariable MR analyses (see ‘Multivariable MR analyses’).

In UKB, blood samples were collected at the assessment centre visit, and in INTERVAL, blood samples were taken during routine blood donation. Prior to GWAS, the study’s authors adjusted the counts for biological and technical laboratory covariates, and GWAS results were provided as SD change in transformed cell count, per risk allele. Adjustments were made for technical and seasonal covariates, as well as age, menopausal status, height, weight, smoking and alcohol. Full details of covariates and transformations are given in Astle et al.¹

Published outcome GWAS data sets (respiratory outcomes)

Quantitative lung function GWASs (Shrine et al., 2019a)²

We used published summary-level data from three GWAS of FEV₁, FVC and FEV₁/FVC, undertaken in UK Biobank (N=321,047) and the SpiroMeta consortium (N=79,055).² Prior to GWAS, traits were pre-adjusted for age,² sex, height, smoking status and other covariates as appropriate, e.g. ancestry principal components. Residuals were inverse-normal rank transformed. UK Biobank and SpiroMeta results were combined by meta-analysis. GWAS results restricted to the SpiroMeta consortium only were used to assess the effect of sample overlap in sensitivity analyses.

Moderate-to-severe asthma GWAS (Shrine et al., 2019b)³

We used a GWAS of moderate-to-severe asthma within the Genetics of Asthma Severity and Phenotypes (GASP) initiative, with additional cases included from the U-BIOPRED asthma cohort, and UK Biobank.³ All cases (N=5135) were taking medication for asthma, and met the criteria for moderate-to-severe asthma according to the British Thoracic Society (BTS) 2014 guidelines. Controls (N=25,675) were from UK Biobank, and excluded those with a doctor-diagnosis of asthma, rhinitis, eczema, allergy, emphysema, or chronic bronchitis, or those with missing medication data. Analyses were adjusted for the first 10 principal components.

Overlap between the exposure and outcome datasets

Individuals from UK Biobank were included in the exposure blood cell GWAS and all outcome respiratory GWAS. There are varying degrees of sample overlap for each individual MR analysis.

UK Biobank initially released genetic data for up to ~150,000 participants (~50,000 genotyped on the UK BiLEVE array and selected according to extremes of lung function and smoking behaviour,⁴ and an additional ~100,000 genotyped on the closely related UK Biobank Axiom array).³ This is referred to as the ‘interim release’ of UK Biobank genetic data, and includes about 1/3 of UK Biobank

¹ Astle et al. 2016
² Shrine et al. 2019a
³ Shrine et al. 2019b
⁴ BiLEVE
⁵ UK Biobank Axiom array
participants. The ‘full release’ of UK biobank genetic data followed later, and included data on >450,000 participants.

The sampling strategies for each GWAS were as follows: the exposure blood cell GWAS included up to ~173,000 individuals in total (exact sample size varied according to cell type, \(N=172,275\) for eosinophils, see Supplementary Table 3 for sample sizes of all cell types). This included up to 132,959 individuals from the UK Biobank first release of genetic data, and up to 40,521 samples from the INTERVAL study. Samples were of European ancestry.

The most prominent overlap was for the asthma GWAS dataset, since individuals were also only sampled from the interim release of UK Biobank data (around 1/3 of participants). However, the asthma outcome GWAS was supplemented with cases from GASP and UBIOPRED.

The lung function, ACO and AECOPD GWAS data sets were sampled from 321,057 European ancestry individuals within the full release of UK Biobank genetic data who also had lung function measures passing QC.

Whilst it is not possible to calculate the exact degree of sample overlap, likely estimates are presented overleaf, where overlap is the proportion of participants in the outcome GWAS who are also likely to feature in the exposure GWAS. For studies including UK Biobank data only, this figure is likely to be around 30%.
### Estimation of % participants in outcome GWAS expected to feature in exposure GWAS

<table>
<thead>
<tr>
<th>Exposure data set</th>
<th>Outcome data sets</th>
<th>% participants in outcome GWAS expected to feature in exposure GWAS*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blood cell types</td>
<td></td>
<td></td>
</tr>
<tr>
<td>~132,959 participants sampled from interim release of UK Biobank genetic data</td>
<td>Lung function (four traits)&lt;sup&gt;2&lt;/sup&gt; 321,047 (full release) 79,055 (SpiroMeta)</td>
<td>23%</td>
</tr>
<tr>
<td></td>
<td>Moderate-severe asthma&lt;sup&gt;3&lt;/sup&gt; 2,996 cases (interim release) 25,600 controls (interim release)</td>
<td>55% (cases) 94% (controls)</td>
</tr>
<tr>
<td></td>
<td>Respiratory infections&lt;sup&gt;2&lt;/sup&gt; 19,459 cases (full release) 101,438 controls (full release)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>AECOPD 2,771 cases (full release) 42,052 controls (full release)</td>
<td></td>
</tr>
<tr>
<td>~40,521 INTERVAL participants&lt;sup&gt;1&lt;/sup&gt;</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>ACO&lt;sup&gt;6&lt;/sup&gt; 8,068 cases (full release) 40,360 controls (full release)</td>
<td>29% (cases) 29% (controls)</td>
</tr>
<tr>
<td></td>
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<td></td>
</tr>
</tbody>
</table>

*Core assumptions for calculations above:
- Assume phenotype availability is random with respect to genotype availability, for all GWAS
- 463,844 participants with genotype data and of European ancestry in full release<sup>3</sup>
- 152,725 genotyped participants in interim release<sup>8</sup>
- 141,751 of the above designated European ancestry in interim release<sup>9</sup>
- 132,959 (assumed as a subset of the above) in blood cell type GWAS (Astle et al. 2016)<sup>1</sup>
Statistical methods

Univariable MR analyses of eosinophils and all respiratory traits and diseases

As described in the main manuscript, we first performed MR analyses of eosinophils on all outcomes, including three quantitative lung function traits (FEV₁, FVC, and FEV₁/FVC); and four clinical disease phenotypes (asthma, AECOPD, ACO and respiratory infections), using genetic instrumental variables (IVs) selected from Astle et al. (2016)¹.

Astle et al. identified 209 conditionally distinct eosinophil count signals at p<8.31x10⁻¹⁰ (their threshold for genome-wide significance), and we extracted effect sizes and standard errors for these SNPs from the meta-analysis of UKB and SpiroMeta from the GWAS of the three lung function traits.² Where SNPs were unavailable, we sought linkage disequilibrium (LD) proxies at r²>0.8 in a European sample using the ‘Aggr’ tool (https://preventivemedicine.usc.edu/divisions/biostatistics/biostatistics-software/) that were also associated at p<8.31x10⁻⁹ in the eosinophil GWAS.

Using the R package ‘TwoSampleMR’ (https://mrclieu.github.io/TwoSampleMR/, version 0.4.23), we harmonised the SNP-eosinophil and SNP-lung function associations so that the effect sizes corresponded to the same allele. For A/T and C/G SNPs, minor allele frequency (MAF) was used to infer strand, and SNPs were dropped from the analysis if they had a MAF of >0.42, since this precluded reliable strand inference. We then performed LD clumping to retain a set of 151 SNPs that were independent at r²<0.01, using LD data from the European 1000 Genomes population. These SNPs (and proxies) were then additionally extracted from the other outcome GWAS (one SNP was missing from the asthma GWAS).

In the analyses of eosinophils and respiratory phenotypes, we report estimates from three MR methods, each of which are robust to different violations of the core assumptions shown in Box 1.

Inverse-variance weighted analyses

The primary analysis used the inverse-variance weighted (IVW) MR method, which combines Wald ratios (or for binary outcomes, Wald-type ratios²⁰) of SNP-outcome to SNP-exposure effects across all SNPs by meta-analysis (we used a multiplicative random-effects model that corrects for under-distribution in the model). The method requires that if SNPs are associated with the outcome via pathways other than the exposure (Box 1), the average effect through these pathways for these SNPs should be zero (e.g. any “horizontal pleitropy” is ‘balanced’). Moreover, horizontal pleitropic effects should be unrelated to SNP-exposure effects (the “InSIDE” assumption—Instrument Strength Independent of Direct Effect).¹¹ ¹² We assessed horizontal pleitropy by: i) computing Cochran’s Q statistic to assess evidence of over-dispersion of causal estimates, ii) plotting SNP-exposure effects against SNP-outcome effects.¹²

MR-Egger analyses

MR-Egger analysis performs a weighted regression of SNP-outcome associations on SNP-exposure associations, allowing a non-zero intercept, so that potentially all IVs used could be invalid (e.g. have a non-zero effect on the outcome even when the effect of the exposure on the outcome is zero).¹¹ However, MR-Egger is sensitive to violation of the InSIDE assumption, and has less statistical power than the IVW and weighted median methods.

Weighted median analyses

The weighted median estimate is robust to violation of the InSIDE assumption and the presence of horizontal pleitropy, provided that the IVs providing ≥50% of the total weight are valid, without having to specify which ones are invalid.¹³
**MR-PRESSO analyses**

We used MR-PRESSO to further assess for the possible impact of horizontal pleiotropy on our results. MR-PRESSO first conducts a ‘global test’, by comparing the observed residual sum of squares to the expected value, assuming no horizontal pleiotropy, for a group of variants. It then tries to identify specific variants which may be outliers due to horizontal pleiotropy, by comparing the observed and expected distributions of one variant only. A distortion test then quantifies the impact of removing the outliers on the causal estimate.\(^\text{14}\)

**Multivariable MR analyses of multiple blood cell types and respiratory outcomes**

**Instrument selection**

Since SNPs affecting eosinophils also affect other blood count types,\(^\text{1}\) we used multivariable MR in order to estimate the influence of multiple cell types on respiratory outcomes, after conditioning on the effects of the SNPs on other cell types (see below). Multivariable MR (MVMR) analyses were run for respiratory outcomes that showed evidence of eosinophil causation in the IVW MR analyses above, and that had broadly consistent effect estimates in the weighted median and MR-Egger analyses.

The aim of this analysis was therefore twofold: i) to further investigate the possibility of horizontal pleiotropy affecting the results of the eosinophil MR; and ii) to establish whether any other cell types besides eosinophils could affect the respiratory outcomes studied.

We extracted all of the genome-wide signals reported by Astle et al.\(^\text{1}\) for GWAS of counts of the following blood cell types: eosinophils (as described), basophils, neutrophils, monocytes, lymphocytes, platelets, red blood cells, and reticulocytes. Across all traits, a total of 1166 SNPs were also available in the outcome GWAS.

We performed LD clumping across all 1166 SNPs (**Supplementary Table 3**). This resulting set of 318 SNPs (including SNPs associated with multiple traits) was then extracted from the GWAS results of each cell type, and also from each of the outcome GWAS. Harmonisation of SNP-exposure and SNP-outcome effects was as described previously.

**Inverse-variance weighted MVMR**

To implement inverse-variance weighted multivariable MR (IVW MVMR), we used the \texttt{mv\_multiple()} function of the ‘TwoSampleMR’ R package.\(^\text{12}\) This implements an approach described as a modification\(^\text{15}\) to the MVMR method described by Burgess and Thompson.\(^\text{16}\) Briefly, the method is performed by regressing the SNP-outcome associations on the SNP-exposure associations for all cell types simultaneously. This is therefore a multivariable weighted regression model (without an intercept), that uses inverse-variance weights.

**Multivariable MR adjusting for weak instruments**

MVMR estimation using the IVW approach (as above) is robust to the presence of balanced horizontal pleiotropy if the instrumental variables used are strong. To estimate the strength of the IVs in predicting each of the eight exposures, we calculated the conditional F-statistic for each exposure (**Supplementary Table 3**).\(^\text{17}\) In the presence of weak instruments, false positive results for the detection of pleiotropy in MR analyses are more likely.

We calculated a modified form of Cochran’s Q statistic (\(Q_s\)), described by Sanderson et al., this exact test for detecting pleiotropy in MVMR is robust even in the presence of weak instruments.\(^\text{17}\) We implemented this using the pleiotropy\_mvmr() function of the ’MVMR’ R package.
We additionally used a method developed by the same authors to perform MVMR in the presence of moderately weak instruments. This approach estimates causal effects whilst accounting for excess heterogeneity (unrelated to variance in SNP-exposure or SNP-outcome associations) in the per-SNP effects, and is more robust to balanced pleiotropy. It was implemented using the qhet_mvmr() function in the ‘MVMR’ R package.

A jack-knife procedure was used to calculate standard errors ($SE_{jack}$) for \( \hat{\theta} \), the causal estimate, as adapted from \(^{18}\): briefly, each of \( i = 1, 2, \ldots, n \) SNP IVs was omitted in turn, and the causal estimate re-estimated for the \( i \)th jack-knife sample, giving \( n \) estimates in total, where \( \hat{\theta}_{(i)} \) is the \( i \)th jack-knife replication of \( \hat{\theta} \), e.g. the causal estimate from the dataset with the \( i \)th SNP IV removed. \( SE_{jack} \) for an exposure-outcome causal effect, \( \hat{\theta} \), are then given as:

\[
SE_{jack} = \left[ \frac{n - 1}{n} \sum_{i=1}^{n} (\hat{\theta}_{(i)} - \hat{\theta}_{(.)})^2 \right]^{1/2}
\]

where \( \hat{\theta}_{(.)} = \frac{\sum_{i=1}^{n} \hat{\theta}_{(i)}}{n} \)

**Multivariable MR, omitting variants contributing most to heterogeneity (quantified by Q statistic)**

Finally, we examined the individual contribution of each SNP IV to the MVMR estimates, by omitting each SNP in turn. The absolute percentage reduction in the \( Q_A \) statistic after omitting a given SNP, compared to the \( Q_A \) statistic when including all SNPs in the model was calculated. SNPs that led to a reduction in \( Q \) by at least 2.5% were noted (Supplementary Table 11), and IVW MVMR models were recalculated without this subset of SNPs.

**References**


These scatterplots show the relationship between the SNP-eosinophil and SNP-outcome effects for 151 SNPs used in univariable MR analyses, for three quantitative lung function traits: forced expiratory volume in 1 second (FEV₁), forced vital capacity, and the ratio of FEV₁/FVC. Each black dot represents a SNP, and whiskers represent precision. The three fitted lines correspond to the causal effect estimates using three separate MR methods. Units are in SD change (see Extended Methods).
These scatterplots show the relationship between the SNP-eosinophil and SNP-outcome effects for 151 SNPs used in univariable MR analyses, for four binary respiratory phenotypes: moderate-to-severe asthma, acute exacerbations of COPD, asthma-COPD overlap, and respiratory infections. Each black dot represents a SNP, and whiskers represent precision. The three fitted lines correspond to the causal effect estimates using three separate MR methods. Units are in SD change (see Extended Methods).
### Supplementary Figure 3

<table>
<thead>
<tr>
<th>Trait</th>
<th>Outcome</th>
<th>Beta_95CI</th>
<th>P</th>
</tr>
</thead>
<tbody>
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<td>Basophils</td>
<td>FEV1FVC</td>
<td>-0.002 (-0.06, 0.056)</td>
<td>0.939</td>
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<td>FEV1FVC (UKB)</td>
<td>-0.005 (-0.069, 0.058)</td>
<td>0.876</td>
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<tr>
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<td>FEV1FVC (UKB asthma)</td>
<td>-0.025 (-0.133, 0.083)</td>
<td>0.648</td>
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<tr>
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<td>FEV1FVC (UKB no asthma)</td>
<td>-0.006 (-0.068, 0.055)</td>
<td>0.839</td>
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<tr>
<td>Eosinophils</td>
<td>FEV1FVC</td>
<td>-0.065 (-0.104, -0.026)</td>
<td>0.001</td>
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<td>-0.071 (-0.113, -0.028)</td>
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<td>FEV1FVC (UKB no asthma)</td>
<td>-0.028 (-0.069, 0.013)</td>
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<td>Lymphocytes</td>
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<td>FEV1FVC (UKB no asthma)</td>
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<td>FEV1FVC (UKB no asthma)</td>
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<td>FEV1FVC</td>
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<td>-0.027 (-0.097, 0.043)</td>
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<td>0.009 (-0.031, 0.049)</td>
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<td>Platelets</td>
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<td></td>
<td>FEV1FVC (UKB asthma)</td>
<td>-0.009 (-0.053, 0.036)</td>
<td>0.706</td>
</tr>
<tr>
<td></td>
<td>FEV1FVC (UKB no asthma)</td>
<td>-0.001 (-0.027, 0.024)</td>
<td>0.926</td>
</tr>
<tr>
<td>RBCs</td>
<td>FEV1FVC</td>
<td>-0.019 (-0.049, 0.011)</td>
<td>0.22</td>
</tr>
<tr>
<td></td>
<td>FEV1FVC (UKB)</td>
<td>-0.019 (-0.052, 0.014)</td>
<td>0.255</td>
</tr>
<tr>
<td></td>
<td>FEV1FVC (UKB asthma)</td>
<td>-0.034 (-0.089, 0.021)</td>
<td>0.229</td>
</tr>
<tr>
<td></td>
<td>FEV1FVC (UKB no asthma)</td>
<td>-0.018 (-0.05, 0.013)</td>
<td>0.258</td>
</tr>
<tr>
<td>Retics</td>
<td>FEV1FVC</td>
<td>0.002 (-0.023, 0.028)</td>
<td>0.871</td>
</tr>
<tr>
<td></td>
<td>FEV1FVC (UKB)</td>
<td>0.001 (-0.027, 0.029)</td>
<td>0.959</td>
</tr>
<tr>
<td></td>
<td>FEV1FVC (UKB asthma)</td>
<td>-0.022 (-0.07, 0.025)</td>
<td>0.356</td>
</tr>
<tr>
<td></td>
<td>FEV1FVC (UKB no asthma)</td>
<td>0 (-0.027, 0.026)</td>
<td>0.973</td>
</tr>
</tbody>
</table>

**Legend**

Forest plot showing multivariable MR estimating the causal effect of multiple cell types on the ratio of forced expiratory volume in 1 second (FEV₁) to forced vital capacity (FVC), after conditioning on the effects of the SNPs on other cell types. Models were run using four sets of SNP-outcome results: GWAS results for FEV₁/FVC from UK Biobank and SpiroMeta [FEV1FVC, N=400,102]; results from UK Biobank only [FEV1FVC (UKB), N=321,047];
results from UK Biobank only, restricting to individuals with asthma [FEV1/FVC (UKB asthma), N=37,868]; and results from UK Biobank only, after excluding individuals with asthma [FEV1/FVC (UKB no asthma), N=283,179]. Effect sizes (beta, 95% confidence interval, 95CI) are in SD change in lung function outcome per SD cell count (adjusted for the effects of other cell types). Points of the forest plot represent effect size estimate; whiskers are 95% confidence intervals.