Add-on azithromycin reduces sputum cytokines in non-eosinophilic asthma: an AMAZES substudy

Shakti D Shukla (1), ¹ Steven L Taylor (1), ^{2,3} Peter G Gibson, ^{1,4} Daniel Barker, ¹ John W Upham, ^{5,6} Ian A Yang (1), ^{5,7} Paul N Reynolds, ^{8,9} Sandra Hodge, ^{8,9} Alan L James, ^{10,11} Geraint B Rogers, ^{2,3} Jodie L Simpson (1), ^{1,4}

► Additional material is published online only. To view, please visit the journal online (http://dx.doi.org/10.1136/ thoraxjnl-2020-216331).

For numbered affiliations see end of article.

Correspondence to

Dr Jodie L Simpson, Respiratory and Sleep Medicine, The University of Newcastle Faculty of Health and Medicine, Callaghan, NSW 2305, Australia; jodie.simpson@newcastle. edu.au

Received 7 October 2020 Revised 16 November 2020 Accepted 7 December 2020



© Author(s) (or their employer(s)) 2021. No commercial re-use. See rights and permissions. Published by BMJ.

To cite: Shukla SD, Taylor SL, Gibson PG, et al. Thorax Epub ahead of print: [please include Day Month Year]. doi:10.1136/ thoraxjnl-2020-216331

BMJ

ABSTRACT

Add-on azithromycin (AZM) significantly reduces exacerbations in poorly controlled asthma irrespective of disease phenotype. In a predefined substudy of the original AMAZES protocol (500 mg, three times a week for 48 weeks), we report that AZM treatment reduces key sputum inflammatory proteins (interleukin (IL)-6, IL-1 β and extracellular DNA), which is more evident in non-eosinophilic asthma (NEA). Moreover, AZM reduced *Haemophilus influenzae* load only in NEA. Our data support the anti-inflammatory effects of AZM in poorly controlled asthma. Prospective studies are required to identify patients that derive greatest benefit from AZM add-on therapy.

Adults with poorly controlled asthma experience exacerbations, despite maintenance treatment with inhaled corticosteroids and long-acting bronchodilators. This population is heterogeneous and includes those with eosinophilic asthma (EA), characterised by type 2 airway inflammation, and those with non-eosinophilic asthma (NEA), who exhibit predominantly neutrophilic inflammation¹ and whose airways are commonly colonised by high levels of Gammaproteobacteria, including Haemophilus influenzae and Moraxella catarrhalis.² For patients with EA, biologic therapies (anti-IgE, anti-interleukin (IL)-5 and anti-IL-5Ra) have proven transformative in preventing exacerbations by acting on eosinophilic inflammation. However, effective treatment options for NEA are limited.

We have previously shown that add-on oral azithromycin (AZM) reduces exacerbations in poorly controlled asthma, including NEA.¹ We now report an a priori substudy of the original trial protocol to assess the effect of AZM on soluble proinflammatory mediators IL-6, IL-1 β and extracellular DNA (eDNA) in EA and NEA. Given the recent findings that AZM reduces *H. influenzae* abundance,³ and high baseline *H. influenzae* predicts clinical benefit from AZM treatment,⁴ we further relate *H. influenzae* to eosinophilic phenotype, and proinflammatory mediator levels.

Paired induced sputum samples from 212 participants before and after add-on AZM (n=109; 500 mg, three times a week for 48 weeks) or placebo (n=103) (table 1) were assessed. This subgroup showed a significant reduction in total exacerbations with AZM (p=0.001, table 1), consistent with the original study. Participants underwent a clinical assessment and gave written informed consent.¹ Sputum inflammatory cell counts were performed after dithiothreitol dispersion and characterised as eosinophilic or non-eosinophilic.¹ Supernatant concentrations of IL-6, IL-8, IL-1B and eDNA, and raw sputum H. influenzae abundance were measured as previously described⁴⁻⁶ and detailed in the online supplemental. IL-8 and H. influenzae were measured in smaller subset of samples (IL-8: n=87 baseline sample, H. influenzae n=112 baseline and 61 endpoint samples) due to sample availability. The relationship between baseline measures were assessed by Spearman's rank correlation with p value adjusted for multiple comparisons using the Bonferroni test. Associations between end of treatment levels of inflammation and H. influenzae with treatment allocation were analysed using linear mixed models, with a fixed effect used to adjust for baseline levels and a random effect for study site, as defined a priori and performed in the initial Asthma and Macrolides: the Azithromycin Efficacy and Safety Study (AMAZES) trial.¹

Markers of neutrophilic inflammation and *H. influenzae* were highly co-correlated at baseline (figure 1A, online supplemental figure 1). After stratification by inflammatory phenotype, *H. influenzae* was only significantly correlated with IL-1 β in those with NEA (R_s=0.546, p=0.004, n=41, figure 1B, online supplemental figures 2 and 3). Eosinophil count correlated with neutrophil count, however not with any other markers of inflammation or *H. influenzae* (figure 1A–C).

Compared with placebo, add-on AZM resulted in a significant reduction in sputum *H. influenzae*, IL-6, IL-1 β and eDNA (figure 1D, online supplemental table 1). A significant reduction in *H. influenzae*, IL-6, IL-1 β and eDNA, remained in those with NEA (figure 1E, online supplemental table 1), while only IL-6 was significantly reduced following AZM in those with EA (estimate (95% CI)=-0.292 (-0.570 to -0.015); p=0.039, figure 1F, online supplemental table 1).

We further investigated whether changes in inflammatory markers related to changes in *H. influenzae* levels following AZM. In the 24 participants in whom *H. influenzae* and IL-1 β were measured at baseline and following AZM, reduction in *H. influenzae* was strongly correlated with the reduction in IL-1 β (R_s=0.949, p<0.001). This relationship remained strong for those with NEA (R_s=0.958, p<0.001, n=12), while weaker, although significant, for EA (R_s=0.790, p=0.046, n=12). Changes in IL-6, eDNA and neutrophils did not correlate with changes in *H. influenzae* following AZM.

Collectively, our findings show a relationship



	Placebo N=103	Azithromycin N=109 60.3 (12.4)	
Age (years)*	57.2 (16.5)		
Gender (M/F)	44/59	50/59	
Ex-smoker†	39 (38%)	39 (36%)	
Pack years‡	7.5 (1.4–25.0)	9.3 (1.5–26.3)	
Atopy	83 (81%)	79/107 (74%)	
ACQ6 Score*	1.83 (0.78)	1.79 (0.84)	
Eosinophilict	53 (51%)	46 (42%)	
Asthma history past year‡			
Emergency room visit or hospital admission	0 (0–0)	0 (0–0)	
Unscheduled doctor visits	0 (0–2)	1 (0–2)	
Oral corticosteroid courses	1 (0–2)	1 (0–2)	
Medications†			
ICS daily dose as beclomethasone equivalent µg/day‡	1000 (200–4500)	1480 (200–4000)	
ICS/LABA	102 (99%)	106 (97%)	
Oral corticosteroid	2 (1.9%)	3 (2.8%)	
Spirometry, pre β2-agonist*			
FEV ₁ % predicted	74.75 (15.84)	74.53 (19.68)	
FVC% predicted	84.16 (14.13)	84.94 (14.24)	
FEV ₁ /FVC%	68.77 (10.39)	67.31 (12.05)	
Sputum cell counts‡			
Viability	72 (9–100)	67 (7–100)	
Total cell count (10 ⁶ /mL)	4.41 (0.31–69.93)	4.05 (0.45–36.00)	
Neutrophils%	34.38 (0.25–95.75)	35.25 (0.68–95.25)	
Neutrophils×10 ⁴ /mL	116.8 (0.67–3486)	126.2 (3.04–3296)	
Eosinophils%	2.13 (0.00–52.50)	1.50 (0.00-83.00)	
Eosinophils×10 ⁴ /mL	8.34 (0.00–547.2)	6.23 (0.00–888.9)	
Macrophages%	50.38 (2.25–93.75)	43.88 (2.00–93.00)	
Lymphocytes%	0.50 (0.00–14.50)	0.50 (0.00–10.75)	
Columnar epithelial%	3.25 (0.00–59.50)	2.13 (0.00–79.00)	
Exacerbations/person-year during trial			
Total	1.58	0.95	
Severe	0.70	0.51	
Moderate	0.89	0.44	

*Mean(SD).

tn/N(%); FEV₁. ‡Median(Q1,Q3).

ACQ6, Asthma Control Questionnaire 6; FEV., forced expiratory volume in one second; FVC, forced vital capacity; ICS, inhaled corticosteroid; LABA, long-acting beta agonist.

between inflammatory mediators associated with neutrophilic inflammation and *H. influenzae*, and that add-on AZM therapy reduces these inflammatory mediators. These relationships were more pronounced in NEA compared with EA. Our previous substudy, which examined RNA gene copy number, identified that AZM did not affect inflammatory gene expression.⁷ The impact of AZM on inflammatory protein expression reported here may be due to a downstream mechanism, such as AZMinduced inhibition of protein translation.

It remains unclear whether the benefit provided by AZM occurs via different pathways in those with EA and NEA, or whether there is a common mechanism that is more pronounced in NEA. It is also yet to be determined whether relationships between reduction in *H. influenzae* and reduction in

inflammatory marker levels represent causal interactions, and what the direction of any such relationship might be.

In adults with NEA, the reduction in IL-1 β , eDNA, IL-6 and *H. influenzae* following AZM is a potential mechanism of clinical benefit. As reviewed in detail elsewhere,⁸ the reduction in inflammatory markers by macrolides such as AZM can reduce an overt neutrophil response that is associated with corticosteroid-resistant asthma exacerbations, as well as improve monocyte/macrophage activity leading to clearance of proinflammatory material, including *H. influenzae*.

Our study focused on identifying the effects of AZM in those with NEA, given the current lack of effective therapies for this patient group. However, AZM was found to reduce IL-6 in both EA and NEA. IL-6 may be involved in a conserved mechanism

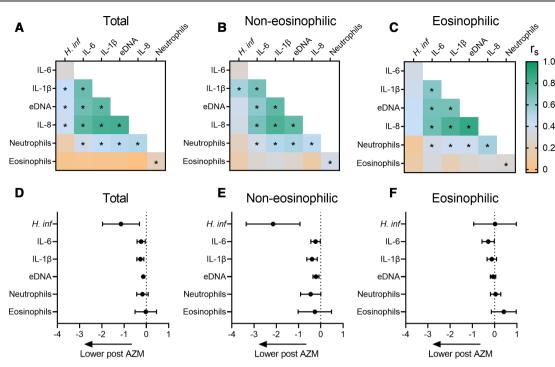


Figure 1 Relationship among azithromycin (AZM), markers of airway inflammation and airway *Haemophilus influenzae* levels in adults with persistent uncontrolled asthma, stratified by eosinophilic phenotype. (A–C) Heatmap showing Spearman's correlations of baseline measures for (A) all participants, (B) non-eosinophilic asthma and (C) eosinophilic asthma. Each cell represents a correlation between the measure on the left of the heat map and the top measure, coloured by the strength of the correlation coefficient (r_s). The asterisk (*) indicates p<0.05 (Bonferroni corrected). (D,E) Linear mixed model regression analyses assessing the effect of AZM on endpoint measures adjusting for baseline levels and study site for (D) all participants, (E) non-eosinophilic asthma and (F) eosinophilic asthma. Negative values indicate lower post treatment levels in the AZM group. eDNA, extracellular DNA; *H. inf, Haemophilus influenzae*; IL, interleukin.

by which AZM acts across asthma. For example, in EA, an AZMdependent reduction in IL-6 could reduce its role in promoting T helper (Th)2 differentiation over Th1,⁹ while in NEA, an AZM-dependent reduction in IL-6 could impair Th17 differentiation, limiting neutrophilic activity.⁹ Alternatively, IL-6 could contribute to asthma inflammation via trans-signalling of the soluble IL-6 receptor, which is crucial in expression of genes involved in regulation of airway remodelling and innate immune activation.¹⁰

In conclusion, our findings indicate that long-term administration of AZM attenuates key sputum inflammatory markers (IL-6, IL-1 β and eDNA) as well as levels of

H. influenzae, especially in patients with NEA. Sputum IL-6 levels were significantly reduced in both EA and NEA. These antiinflammatory effects of AZM may contribute to the reduction in asthma exacerbations observed in the main study.¹ Achieving a better understanding of the mechanistic basis of AZM benefit should now be prioritised as a means to enable identification of those patients most likely to benefit.

Author affiliations

¹Faculty of Health and Medicine, The University of Newcastle Priority Research Centre for Asthma and Respiratory Disease, Newcastle, New South Wales, Australia ²Microbiome and Host Health, South Australian Health and Medical Research Institute, Adelaide, South Australia, Australia

³SAHMRI Microbiome Research Laboratory, College of Medicine and Public Health, Flinders University, Adelaide, South Australia, Australia

⁴Hunter Medical Research Institute, Newcastle, NSW, Australia

⁵Faculty of Medicine, University of Queensland, Brisbane, QLD, Australia

⁶Translational Research Institute, Brisbane, QLD, Australia

⁷Department of Thoracic Medicine, The Prince Charles Hospital, Brisbane, QLD, Australia

⁸Department of Respiratory Medicine, Royal Adelaide Hospital, Adelaide, South Australia, Australia ⁹School of Medicine, University of Adelaide, Adelaide, SA, Australia
¹⁰Department of Pulmonary Physiology and Sleep Medicine, Sir Charles Gairdner Hospital, Nedlands, Western Australia, Australia

¹Medicine School, University of Western Australia, Crawley, WA, Australia

Twitter Shakti D Shukla @shaktishukla09 and Jodie L Simpson @jlgiffo

Acknowledgements We wish to acknowledge the AMAZES research group for undertaking the study, and Ms. Kellie Fakes and Dr. Kavita Pabreja (The University of Newcastle, Callaghan, Australia) who undertook the laboratory assessment of enzyme-linked immunosorbent assay and data analyses, as well as the AMAZES study participants.

Contributors Conception and design: JLS, PGG, JWU, IY, PNR, SH and AJ. Data acquisition: JLS, PGG, JWU, IY, PNR, SH, AJ, ST and GBR. Data analysis and interpretation: JLS, DB, SDS, ST and GBR. Drafting manuscript: SDS, JLS, DB, ST and GBR. Revision and approval of final manuscript: All authors.

Funding This study was supported by National Health and Medical Research Council (NHMRC) (grant 569246) and NHMRC Centre for Severe Asthma, University of Newcastle.

Competing interests JWU reports personal fees from AstraZeneca, personal fees from GSK, personal fees from Novartis, personal fees from Boehringer Ingelheim, personal fees from Sanofi, outside the submitted work. PGG reports personal fees from AstraZeneca, GlaxoSmithKline, Novartis, grants from AstraZeneca, GlaxoSmithKline, outside the submitted work.

Patient consent for publication Not required.

Ethics approval Ethical approval was granted by Hunter New England Human Research Ethics Committee (08/11/19/3.03)

ORCID iDs

Shakti D Shukla http://orcid.org/0000-0002-5796-0171 Steven L Taylor http://orcid.org/0000-0003-4357-8243 Ian A Yang http://orcid.org/0000-0001-8338-1993 Jodie L Simpson http://orcid.org/0000-0001-5299-4236

REFERENCES

- 1 Gibson PG, Yang IA, Upham JW, *et al.* Effect of azithromycin on asthma exacerbations and quality of life in adults with persistent uncontrolled asthma (AMAZES): a randomised, double-blind, placebo-controlled trial. *Lancet* 2017;390:659–68.
- 2 Green BJ, Wiriyachaiporn S, Grainge C, et al. Potentially pathogenic airway bacteria and neutrophilic inflammation in treatment resistant severe asthma. PLoS One 2014;9:e100645.
- 3 Taylor SL, Leong LEX, Mobegi FM, *et al*. Long-Term Azithromycin Reduces *Haemophilus influenzae* and Increases Antibiotic Resistance in Severe Asthma. *Am J Respir Crit Care Med* 2019;200:309–17.
- 4 Taylor SL, Ivey KL, Gibson PG, et al. Airway abundance of Haemophilus influenzae predicts response to azithromycin in adults with persistent uncontrolled asthma. Eur Respir J 2020;56. doi:10.1183/13993003.00194-2020. [Epub ahead of print: 01 Oct 2020].

- 5 Gao P, Gibson PG, Baines KJ, et al. Anti-Inflammatory deficiencies in neutrophilic asthma: reduced galectin-3 and IL-1RA/IL-1β. Respir Res 2015;16:5.
- 6 Wright TK, Gibson PG, Simpson JL, et al. Neutrophil extracellular traps are associated with inflammation in chronic airway disease. *Respirology* 2016;21:467–75.
- 7 Fricker M, Gibson PG, Powell H, *et al*. A sputum 6-gene signature predicts future exacerbations of poorly controlled asthma. *J Allergy Clin Immunol* 2019;144:e11:51–60.
- 8 Zimmermann P, Ziesenitz VC, Curtis N, et al. The immunomodulatory effects of Macrolides-A systematic review of the underlying mechanisms. Front Immunol 2018;9:302.
- 9 Dienz O, Rincon M. The effects of IL-6 on CD4 T cell responses. *Clin Immunol* 2009;130:27–33.
- 10 Robinson MB, Deshpande DA, Chou J, *et al.* II-6 trans-signaling increases expression of airways disease genes in airway smooth muscle. *Am J Physiol Lung Cell Mol Physiol* 2015;309:L129–38.

Add-on Azithromycin reduces sputum cytokines in non-eosinophilic asthma: an AMAZES sub-

study

ONLINE SUPPLEMENT

SUPPLEMENTARY METHODS

Sub-study design

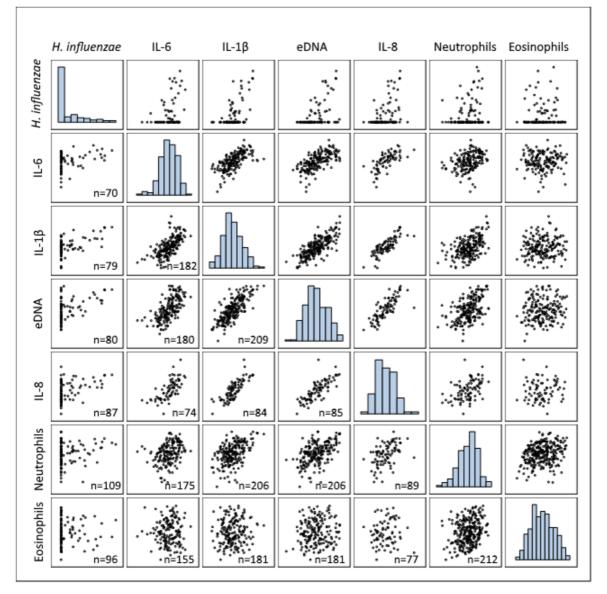
This was a pre-specified sub-study from the initial AMAZES trial protocol (Section 11.2, Appendix I) https://www.severeasthma.org.au/wp-content/uploads/2017/04/AMAZES-Protocol-V15-25.02.14-final.pdf

Inflammatory marker assessment

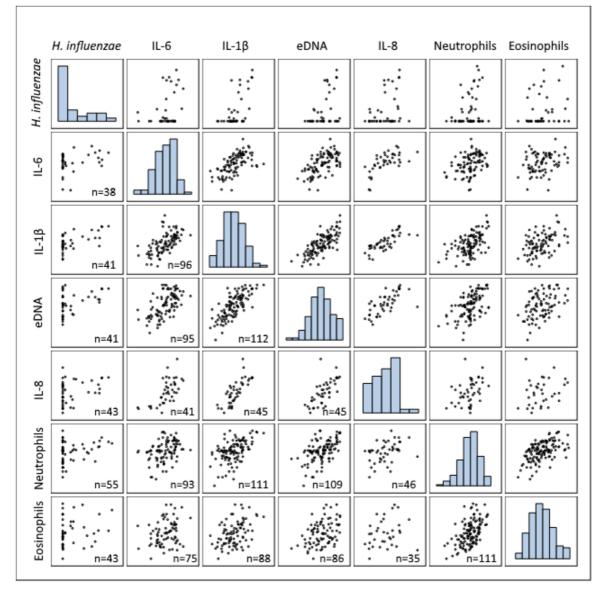
Sputum IL-6, IL-8 and IL-1 β levels were determined by ELISA according to the manufacturer's instructions (R&D Systems; Minneapolis, MS, USA). The validity of IL-6, IL-8 and IL-1 β in sputum supernatant has been reported previously.¹⁻³ The limits of detection of each assay were as following; sputum IL-6: 168.84 pg/ml; IL-8: 31.3 pg/ml; IL-1 β : 35.19 pg/ml. IL-6 and IL-1 β were assessed in all available sputum supernatant pairs. The abundance of eDNA in the cell-free sputum supernatant was quantitated using the Quant-iT PicoGreen dsDNA Assay Kit (P7589, Invitrogen, Carlsbad, CA) as per manufacturer's instructions.⁴ This assay selectively detects double-stranded DNA, which was quantitated in 10 µL of sputum supernatant against a DNA standard curve of 0–200 ng/µL.

Haemophilus influenzae detection in raw sputum

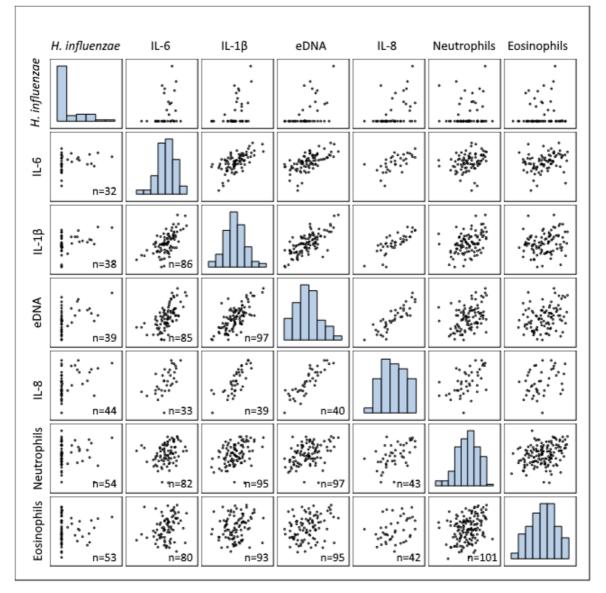
The total DNA was extracted from frozen raw sputum aliquots using a combination of physical, enzymatic, and heat-based cell lysis method that was followed by phenol-chloroform extraction and DNA recovery using EZ-10 Spin columns (Bio Basic, Inc.) as previously described.⁵ *H. influenzae* abundance was measured using a previously published, validated, species-specific qPCR assay with a standard curve of *H. influenzae* strain NTCC8468 of known quantities.^{6,7} Briefly, 1 µL of DNA extract, 0.25 µM of primers (F: ATTAAATGTTGCATCAACGC, R: GACTTTTGCCCACGCAC), 0.2 µM of target probe (FAM-ACGRTTTTACCATAGTTGCACTTTCTC-BHQ1), 17.5 µL of 2X KAPA Probe Fast qPCR Master Mix (KAPA Biosystems Inc., Wilmington, USA) and the appropriate volume of water was added to a 35 µL total reaction volume. Quantitative real-time PCR were performed on three technical replicates, at 10 µL reaction volume per replicate, on a QuantStudio 6 and 7 Flex Real-Time PCR system (Applied Biosystems, Carlsbad, USA). Cycle conditions were 50 cycles of 95 °C for 10 s and 63 °C for 30 s. Serial dilutions of a known concentration of *H. influenzae* strain NTCC8468 was run with every PCR reaction and a copy number per mL of sputum was calculated by comparing sample Ct to the standard curve.



Supplementary Figure 1: Scatter plot matrix of inflammatory mediators and *Haemophilus influenzae* in all participants. X-axis from left to right and y-axis from top to bottom display log_{10} values of *H. influenzae* (copies/mL), IL-6 (pg/mL), IL-1 β (pg/mL), extracellular DNA (ng/mL), IL-8 (pg/mL), neutrophils (count × 10⁴/mL), and eosinophils (count × 10⁴/mL)



Supplementary Figure 2: Scatter plot matrix of inflammatory mediators and *Haemophilus influenzae* in non-eosinophilic asthma. X-axis from left to right and y-axis from top to bottom display \log_{10} values of *H. influenzae* (copies/mL), IL-6 (pg/mL), IL-1β (pg/mL), extracellular DNA (ng/mL), IL-8 (pg/mL), neutrophils (count × 10⁴/mL), and eosinophils (count × 10⁴/mL)



Supplementary Figure 3: Scatter plot matrix of inflammatory mediators and *Haemophilus influenzae* in eosinophilic asthma. X-axis from left to right and y-axis from top to bottom display log_{10} values of *H. influenzae* (copies/mL), IL-6 (pg/mL), IL-1 β (pg/mL), extracellular DNA (ng/mL), IL-8 (pg/mL), neutrophils (count × 10⁴/mL), and eosinophils (count × 10⁴/mL)

	All			Non-eosinophilic		Eosinophilic			
	Observations	Estimate (95% CI)	p- value	Observations	Estimate (95% CI)	p-value	Observations	Estimate (95% CI)	p-value
log ₁₀ (H.influenzae)	AZM: 23 Placebo: 30	-0.309)	0.007	AZM: 11 Placebo: 14	-2.142 (-3.352 to -0.932)	0.001	AZM: 12 Placebo: 16	0.017 (-0.940 to 0.975)	0.971
log_{10} (IL-6)	F lacebo. 80	-0.234 (-0.416 to -0.052)		r lacebo. 41	-0.235 (-0.446 to -0.023)	0.029	AZM: 39 Placebo: 43	-0.292 (-0.57 to -0.015)	0.039
log_{10} (IL-1 β)	AZM: 108 Placebo: 100	-0.268 (-0.423 to -0.114)	0.001	AZM: 57 Placebo: 47	-0.393 (-0.627 to -0.158)	0.001	AZM: 44 Placebo: 50	-0.130 (-0.341 to 0.081)	0.227
$log_{10}(eDNA)$	AZM: 103 Placebo: 98	-0.139 (-0.229 to -0.049)	0.002	AZM: 54 Placebo: 45	-0.215 (-0.351 to -0.080)	0.002	AZM: 42 Placebo: 50	-0.075 (-0.198 to 0.048)	0.234
log_{10} (Neutrophils)	AZM: 92 Placebo: 98	-0.175 (-0.431 to 0.082)	0.183	AZM: 52 Placebo: 47	-0.453 (-0.909 to 0.004)	0.052	AZM: 40 Placebo: 51	0.042 (-0.188 to 0.273)	0.719
log ₁₀ (Eosinophils)	AZM: 92 Placebo: 98	-0.024 (-0.507 to 0.459)	0.922	AZM: 52 Placebo: 47	-0.267 (-1.020 to 0.486)	0.487	AZM: 40 Placebo: 51	0.410 (-0.149 to 0.969)	0.151

Supplementary Table 1. Azithromycin (AZM) effects on inflammatory mediators and *Haemophilus influenzae* in all, eosinophilic and non-eosinophilic asthma. Estimated difference (95% CI) between AZM and placebo at week 48 is shown.

* All models adjust for the baseline level. Negative values of the difference indicate lower post treatment levels in the AZM group. Eosinophilic asthma was defined as sputum eosinophils \geq 3%.

References

- 1. Gao P, Gibson PG, Baines KJ, et al. Anti-inflammatory deficiencies in neutrophilic asthma: reduced galectin-3 and IL-1RA/IL-1β. *Respiratory research*. 2015;16(1):5.
- 2. Fu JJ, McDonald VM, Gibson PG, Simpson JL. Systemic Inflammation in Older Adults With Asthma-COPD Overlap Syndrome. *Allergy Asthma Immunol Res.* 2014;6(4):316-324.
- 3. Neveu WA, Allard JL, Raymond DM, et al. Elevation of IL-6 in the allergic asthmatic airway is independent of inflammation but associates with loss of central airway function. *Respiratory research*. 2010;11:28.
- 4. Wright TK, Gibson PG, Simpson JL, McDonald VM, Wood LG, Baines KJ. Neutrophil extracellular traps are associated with inflammation in chronic airway disease. *Respirology*. 2016;21(3):467-475.
- 5. Taylor SL, Leong LEX, Choo JM, et al. Inflammatory phenotypes in patients with severe asthma are associated with distinct airway microbiology. *Journal of Allergy and Clinical Immunology*. 2018;141(1):94-103.e115.
- 6. Taylor SL, Leong LEX, Mobegi FM, et al. Long-Term Azithromycin Reduces Haemophilus influenzae and Increases Antibiotic Resistance in Severe Asthma. *Am J Respir Crit Care Med.* 2019.
- 7. Reddington K, Schwenk S, Tuite N, et al. Comparison of Established Diagnostic Methodologies and a Novel Bacterial smpB Real-Time PCR Assay for Specific Detection of Haemophilus influenzae Isolates Associated with Respiratory Tract Infections. *Journal of clinical microbiology*. 2015;53(9):2854-2860.