

Adverse effects of salmeterol in asthma: a neuronal perspective

Marek Lommatzsch¹, Yvonne Lindner¹, Anke Edner¹, Kai Bratke¹, Michael Kuepper¹ and
Johann Christian Virchow¹

¹Department of Pneumology, University of Rostock, Germany

Corresponding author:

PD Dr. Marek Lommatzsch

Abteilung für Pneumologie

Klinik und Poliklinik für Innere Medizin

Universität Rostock

Ernst-Heydemann-Str. 6, 18057 Rostock, Germany

Tel: +49 (0) 381 / 494 - 7461, Fax: +49 (0) 381 / 494 - 7392

E-mail: marek.lommatzsch@med.uni-rostock.de

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ABSTRACT

Background: Regular use of inhaled β_2 -agonists has been associated with a paradoxical loss of asthma control and a deterioration of airway hyperresponsiveness, but the underlying mechanism is unknown. The neurotrophin Brain-derived neurotrophic factor (BDNF) has recently been identified as a mediator of airway hyperresponsiveness in asthma.

Methods: Eighteen patients with mild allergic asthma who did not use any regular anti-asthmatic therapy inhaled the long-acting β_2 -agonists salmeterol for 2 weeks followed by 2 weeks of combination therapy with salmeterol and the corticosteroid fluticasone. Airway responsiveness to histamine and BDNF concentrations in blood were assessed prior to entry, after 14 days of salmeterol therapy and after 14 days of combination therapy. In a separate experiment, salmeterol effects on BDNF release by human peripheral blood mononuclear cells were assessed.

Results: Monotherapy with salmeterol significantly increased BDNF concentrations in serum and platelets. This increase was abolished by the addition of fluticasone to the treatment. The findings were confirmed *in vitro*: salmeterol increased the release of BDNF by mononuclear cells, and this was inhibited by co-incubation with fluticasone. Increased BDNF concentrations in serum and platelets correlated with the deterioration of airway hyperresponsiveness following salmeterol monotherapy. In contrast, there was no association between β_2 -receptor polymorphisms and changes in airway responsiveness.

Conclusion: Increased BDNF concentrations may underly the adverse effects of salmeterol monotherapy on airway responsiveness in asthma.

Clinical trial registered with www.clinicaltrials.gov: NCT00736801.

INTRODUCTION

Asthma is characterised by airway inflammation, airway hyperresponsiveness (AHR) and a reversible airflow limitation¹. Inhaled corticosteroids (ICS) are the treatment of choice for asthma. In more severe asthma, international guidelines recommend that ICS can be combined with inhaled long-acting β_2 -agonists (LABA) such as salmeterol². Monotherapy with β_2 -agonists has not been recommended due to accumulating evidence suggesting a loss of control and an excess mortality in asthma with this treatment³. Several studies reported that unbalanced use of short-acting sympathomimetic bronchodilators as well as long-acting β_2 -agonists can deteriorate asthma control, and increase exacerbations and hospital admissions, most likely as a class effect of β_2 -agonists⁴⁻⁸.

Well-controlled clinical studies have demonstrated that regular inhalation of short acting β_2 -agonists such as fenoterol, albuterol, and terbutaline increases airway responsiveness to histamine or methacholine^{4, 9, 10}. This effect is not attributable to a β_2 -receptor subsensitisation^{11, 12}. In addition, the regular use of albuterol has been shown to increase the allergen-induced early¹³ and late asthmatic response¹⁴. Carefully conducted studies on the effect of regular use of LABA in patients with asthma have only been performed in children where regular monotherapy with salmeterol also led to an increase in AHR^{15, 16}. Furthermore, regular inhalation of short (terbutaline) as well as long-acting β_2 -agonists (salmeterol) led to a tolerance of the bronchoprotective actions of both drugs against non-specific bronchoconstrictor stimuli^{17, 18}. In two more recent large scale trials, salmeterol treatment was even associated with excess mortality in asthma^{19, 20}. A trend towards excess mortality in

asthma has recently also been reported for formoterol²¹. The mechanism, however, by which the regular inhalation of β_2 -agonists contributes to increased airway responsiveness and a loss in asthma control is unclear.

The neurotrophin Brain-Derived Neurotrophic Factor (BDNF), a crucial regulator of neuronal activity in the adult²², has been linked to several features of asthma. BDNF is upregulated in allergic airway inflammation and induces AHR and airway obstruction in an animal model of allergic asthma, via an increase of neuronal sensitivity and activity in the airways²³⁻²⁶. In patients with asthma, systemic concentrations of BDNF are increased and these concentrations correlate with AHR²⁷. Following local allergen challenge, endobronchial BDNF levels increase significantly in patients with asthma²⁸. In addition, there is evidence in human asthma that corticosteroids prevent allergen-induced increases in AHR²⁹ and reduce BDNF concentrations^{27, 30, 31}. However, there is no information on the effects of β_2 -agonists on BDNF concentrations in asthma. In this report, we investigate the effect of a monotherapy with a long-acting β_2 -agonist on BDNF concentrations and airway responsiveness in patients with asthma.

METHODS

Study design

The study was performed between September and December 2006 in Rostock (Germany). Patients were recruited by newspaper advertisements. Patients were eligible when they met the following criteria: age > 18 years, a physician's diagnosis of allergic asthma, a documented sensitisation to aero-allergens (pollen, animal hair or house dust mite), no regular treatment (only short-acting inhalers on demand were allowed), no history of or evidence for other chronic disease than asthma, no history of smoking. Prior to inclusion, recruited patients were assessed in the Department of Pneumology (University of Rostock, Germany). Recruited patients were included in the study if they met the following criteria: a pre-bronchodilator forced expiratory volume in the first second (FEV₁) > 80% of the predicted value, a provocative concentration of histamine causing a 20% fall in FEV₁ (PC₂₀) of < 8 mg histamine/ml, and the absence of any signs or symptoms of an infection. After inclusion into the study, blood was collected and patients were instructed in the use of the inhalation device. Patients were asked to inhale salmeterol xinafoate 50 μ g (Serevent Discus®, GSK, Brentford, UK) twice in the morning and twice in the evening for 2 weeks. In the following 2 weeks, patients were asked to inhale salmeterol xinafoate 50 μ g and fluticasone propionate 250 μ g (Viani Discus®, GSK) twice in the morning and twice in the evening (**Fig. 1**). For safety reasons, patients were asked to record their peak flow daily, and to inform the monitor in case of any adverse event or symptomatic deterioration or a drop in peak expiratory flow below 3 liters / second. After 14 days of salmeterol therapy and after 14 days of combination therapy, body plethysmography, assessment of airway responsiveness and blood sampling were repeated (**Fig. 1**). The study medication was withheld for \geq 12 hours prior to lung function testing. The study was approved by the ethics committee of the Ärztekammer Mecklenburg-Vorpommern (Rostock, Germany). Participating subjects gave their written informed consent.

Clinical and laboratory procedures

Pulmonary function, airway responsiveness to histamine, blood cell counts and BDNF concentrations were assessed as previously described³². Monocyte-enriched human peripheral blood mononuclear cells were isolated and cultured as described³² and stimulated with Tumor necrosis factor α (TNF- α , 50 ng/ml), in the presence or absence of salmeterol xinafoate and/or fluticasone propionate (GSK), for 24 hours. Because fluticasone propionate

was dissolved in alcohol, resulting in 0.01% alcohol in culture, 0.01% alcohol was added to control and salmeterol xinafoate cultures. BDNF concentrations measured in supernatants were corrected for the percentage of non-viable cells to exclude artefacts due to corticosteroid-induced apoptosis, as described³². Polymorphisms of the β_2 -receptor were analysed in blood containing ethylenediaminetetraacetic acid (EDTA) using polymerase chain reaction by a commercial laboratory (IMGM Laboratories, Martinsried, Germany).

Statistical analysis

Data were analysed using SPSS (Chicago, IL, USA). Most parameters were non-normally distributed. Correlation analyses between the changes in BDNF concentrations and PC₂₀ after 14 days of salmeterol therapy and after 14 days of combination therapy, and correlation analyses between β_2 -receptor polymorphisms and the changes of the PC₂₀ after 14 days of salmeterol therapy were performed using the Spearman's correlation coefficient. Lung function parameters, platelet counts and BDNF concentrations prior to entry, after 14 days of salmeterol therapy and after 14 days of combination therapy were compared using the signed ranks Wilcoxon test for related samples. Means of BDNF concentrations in cell culture supernatants after 24 hours of incubation with TNF- α alone and TNF- α plus salmeterol xinafoate, fluticasone propionate or salmeterol xinafoate/fluticasone propionate were compared using analysis of variance (ANOVA with SPSS). Probability values of $p < 0.05$ were regarded as statistically significant.

RESULTS

Patient characteristics

A total of 35 patients were recruited (**Fig. 1**). Of these, 14 patients did not meet the inclusion criteria ($n = 11$ had a PC₂₀ of $> 8\text{mg/ml}$, and $n = 3$ had a pre-bronchodilator FEV₁ $< 80\%$ of the predicted value). Of the remaining 21 patients, which were included in the study, 3 patients did not complete the study (one patient withdrew consent during salmeterol therapy due to an unacceptable subjective increase in asthma symptoms; two patients erroneously inhaled the study medication in the morning prior to lung function testing, and were excluded from the study). The baseline characteristics of the 18 patients which completed the study protocol (**Fig. 1**) are given in **Table 1**. Three patients (16%) reported mild adverse effects (subjective discomfort or worsened dyspnea) during salmeterol monotherapy, whereas none of the patients reported adverse effects during combination therapy. There were no severe adverse effects (leading to hospitalisations or death) during the study.

Lung function and airway hyperresponsiveness

The FEV₁ (% predicted) and peak expiratory flow (PEF, % predicted) did not change significantly following 14 days of salmeterol monotherapy (**Fig. 2**). Therapy with salmeterol and fluticasone led to a numerical increase in median FEV₁ and PEF values, as compared to the baseline and to salmeterol monotherapy. However, the differences were not statistically significant (**Fig. 2**). Although there was no statistically significant change in AHR when the whole patient population was analysed, twelve of the 18 patients (67 %) demonstrated an increase in AHR as measured by lower PC₂₀ values to histamine following salmeterol monotherapy compared to baseline (**Fig. 3**). In contrast, when treated with a combination of salmeterol and fluticasone, nearly all patients displayed an increase in PC₂₀ values (**Fig. 3**). AHR improved with combination therapy when compared to baseline in 15 patients (83%), and when compared to values obtained after salmeterol monotherapy in 16 patients (89%). This resulted in a statistically significant increase in the overall PC₂₀ following the addition of fluticasone to salmeterol therapy (**Fig. 3**).

Pat. No.	Age	Sex	Years since diagnosis	PC ₂₀ (mg/ml)	FEV ₁ (% pred.)	Allergies	β ₂ -receptor Polymorphisms	
							arg16gly	gln27glu
1	20	M	13	4.0	91.9	P, D, A	Gly / Gly	Gln / Gln
2	19	M	5	3.8	81.8	D, A	Gly / Gly	Glu / Glu
3	20	M	8	4.5	105.7	P, D	Arg / Gly	Gln / Gln
4	18	M	5	0.2	84.2	P	Arg / Gly	Gln / Glu
5	36	F	5	5.0	103.6	P, D, A	Gly / Gly	Gln / Glu
6	19	F	11	3.3	102.2	P, D, A	Gly / Gly	Gln / Gln
7	39	F	6	4.2	86.8	P, A	Arg / Arg	Gln / Gln
8	20	F	10	3.8	109.1	P, D	Arg / Gly	Gln / Glu
9	42	F	10	5.5	115.3	P	Arg / Gly	Gln / Glu
10	44	M	8	0.4	82.5	D	Arg / Gly	Gln / Glu
11	18	M	3	2.8	95.8	P	Gly / Gly	Glu / Glu
12	29	M	3	5.5	88.7	P, A	Gly / Gly	Glu / Glu
13	18	M	6	0.8	89.9	D, A	Arg / Gly	Gln / Gln
14	38	M	28	0.8	92.0	P, D	Arg / Gly	Gln / Glu
15	26	F	10	1.5	93.2	P, D	Arg / Gly	Gln / Glu
16	19	M	10	1.4	97.3	P, D, A	Gly / Gly	Glu / Glu
17	19	F	8	8.0	87.0	P, D, A	Arg / Gly	Gln / Glu
18	20	F	15	2.7	106.0	P	Arg / Gly	Gln / Glu
Median	20		8	3.6	92.6			

Table 1. Baseline patient characteristics

Abbreviations denote: Pollen (P), House dust mite (D), Animal hair (A), Male (M), Female (F), Forced expiratory volume in the first one second (FEV₁) in percent of the predicted value (% pred.), provocative concentration of histamine causing a 20% fall in FEV₁ (PC₂₀), Arginine (Arg), Glycine (Gly), Glutamine (Gln), Glutamic acid (Glu).

BDNF concentrations in serum, platelets and plasma

There was no statistically significant difference in BDNF concentrations in plasma at baseline (median: 0.09 ng/ml), after salmeterol monotherapy (median: 0.08 ng/ml) and after combination therapy (median: 0.08 ng/ml). In contrast, there was a statistically significant increase in BDNF concentrations in serum and platelets after salmeterol monotherapy compared to baseline. Both serum and platelet BDNF concentrations decreased significantly after 14 days of combination therapy (**Fig. 4**). There were no statistically significant changes in platelet counts at baseline (median: $232 \times 10^6/\text{ml}$), after salmeterol monotherapy (median: $247 \times 10^6/\text{ml}$) and after combination therapy (median: $255 \times 10^6/\text{ml}$).

Association of BDNF with changes in PC₂₀

Changes in BDNF concentrations in serum and platelets were correlated with the changes in PC₂₀ values following salmeterol monotherapy (**Fig. 5**). Although BDNF levels decreased and PC₂₀ values increased significantly following combination therapy with salmeterol and fluticasone, changes in BDNF levels were no longer correlated with the changes in PC₂₀ following combination therapy ($p > 0.05$ for serum and platelet BDNF concentrations).

Effect of salmeterol on BDNF secretion by mononuclear cells

To substantiate the *in vivo* effects of salmeterol and combination therapy on BDNF concentrations, monocyte-enriched peripheral blood mononuclear cells were isolated from 22 healthy volunteers, and stimulated with TNF- α for 24 hours. Fluticasone significantly suppressed BDNF secretion, whereas salmeterol significantly increased BDNF secretion, as compared to medium control (**Fig. 6**). BDNF secretion after co-incubation with salmeterol and fluticasone was not significantly different from medium control ($p = 0.62$), and significantly lower than BDNF secretion after incubation with salmeterol alone (**Fig. 5**).

Impact of β_2 -receptor polymorphisms

In order to test if the effects of salmeterol on airway responsiveness were related to β_2 -receptor polymorphisms, the β_2 -receptor polymorphisms arg16gly (glycine for arginine in position 16) and gln27glu (glutamic acid for glutamine in position 27) were analysed (**Table 1**). As far as the arg16gly polymorphism is concerned, 7 patients (39%) were glycine homozygotes, 1 patient (6%) was an arginine homozygote and 10 patients (55%) were heterozygotes. As far as the gln27glu polymorphism is concerned, 4 patients (22%) were glutamic acid homozygotes, 5 patients (28%) glutamine homozygotes and 9 patients (50%) were heterozygotes (**Table 1**). There was no statistically significant association between the changes in PC₂₀ after salmeterol treatment and the β_2 -receptor polymorphism arg16gly ($r = 0.11$, $p = 0.65$) or the β_2 -receptor polymorphism gln27glu ($r = 0.34$, $p = 0.17$).

DISCUSSION

Inhaled β_2 -agonists provide rapid bronchodilation and have been shown to improve asthma control in a fixed combination with ICS^{33,34}. However, there is now increasing evidence for a paradoxical loss of asthma control and an increase in asthma mortality following β_2 -agonist monotherapy^{3,19,20}. An explanation for this observation is lacking. Several reports highlight that regular treatment with β_2 -agonists in patients with asthma can increase airway responsiveness^{4,9,10} due to mechanisms which are incompletely understood³⁵. There is, however, evidence that increases in airway responsiveness following β_2 -agonist treatment are at least in part explained by changes in the reactivity of airway nerves³⁶.

Mediators with a potential to change neuronal reactivity in the airways are neurotrophins such as BDNF which induce long-term changes in neuronal function and activity^{22, 37}. Animal studies have shown that the production of BDNF by leukocytes and epithelia is strongly upregulated in allergic airway inflammation^{23, 26}. In a functional study, the inhibition of endogenous BDNF reduced AHR in allergen-challenged mice, whereas administration of recombinant BDNF was sufficient to induce AHR in healthy mice²⁵. In the same study, it was demonstrated that these effects of BDNF are due to changes in the neuronal reactivity within the airways²⁵. These findings have recently been confirmed in a study with guinea-pigs³⁸. Thus, there is now accumulating evidence from animal models that BDNF enhances neuronal reactivity in the airways and contributes to neuronal dysfunction and AHR in allergic airway inflammation²⁴.

In patients with asthma, enhanced local BDNF concentrations in the lung²⁸ are mirrored by enhanced BDNF concentrations in circulating platelets²⁷. BDNF is neither produced by platelets nor by its precursors, but actively acquired by platelets³⁹. Therefore, platelet BDNF appears to be an estimate for the average BDNF secretion in organs of the human body over a period of several days³². Accordingly, platelet BDNF (but not plasma BDNF) has been shown to correlate with the severity of AHR²⁷. This association may simply reflect the fact that enhanced BDNF concentrations in the airways lead to both AHR and to an enhanced uptake of BDNF into circulating platelets. However, it is also conceivable that platelet BDNF plays a genuine role in asthma. Platelets have been shown to actively migrate into the lung and to contribute to functional changes within the airways in allergic airway inflammation^{40, 41}. Thus, it can be speculated that not only a BDNF overproduction in the airways by leukocytes and epithelia, but also an enhanced deposition of BDNF by platelets might contribute to the development of AHR in asthma.

In the present study, 14 days of treatment with salmeterol in patients with asthma led to a significant increase in platelet BDNF concentrations and this was correlated to changes in airway responsiveness. In addition, these effects were abolished by adding inhaled fluticasone to the treatment. Thus, the correlation between the changes in AHR and the changes in BDNF concentrations suggests that BDNF might indeed contribute to the development of AHR in asthma and to some of the adverse effects of a salmeterol monotherapy. The mechanisms underlying the induction of BDNF by salmeterol are as yet unclear. However, salmeterol has been shown to increase the transcription of genes with cAMP response elements in their promoters⁴². Thus, as BDNF is known to have active cAMP response elements in its promoter⁴³, salmeterol could enhance BDNF transcription via this pathway.

Concomitant treatment with fluticasone led to a significant decrease in platelet BDNF levels *in vivo* and to a significant reduction in BDNF secretion by leukocytes *in vitro*. It has already been shown that ICS such as fluticasone reduce BDNF secretion by leukocytes²⁷ as well as BDNF serum levels in patients with allergic asthma³¹. This is not specific for fluticasone because other corticosteroids (such as prednisolone and dexamethasone) also suppress BDNF secretion^{27, 30}. ICS can prevent the increase in AHR following allergen challenge²⁹, which is associated with an increase in BDNF concentrations²⁸. Accordingly, the inhibition of BDNF production by ICS as observed in this and previous studies²⁷ further suggests that BDNF might indeed participate in the pathogenesis of AHR. Therefore, some of the beneficial effects of ICS in a combination with LABA in asthma might be related to a suppression of LABA-induced BDNF overexpression. It is of note that the decrease in BDNF levels did not correlate with the increase in PC₂₀ values following combination therapy. This might be due to the fact that fluticasone does not only suppress BDNF, but does have several other effects on a variety of cell types which influence airway responsiveness.

In this study, a deterioration in AHR following salmeterol monotherapy was observed in the majority (67%) of the patients. These data are consistent with studies showing adverse effects of long-acting^{15, 16} and short-acting β_2 agonists^{4, 9, 10} on airway responsiveness in asthma. Genotyping for two β_2 -receptor polymorphisms implicated in the response to β_2 -agonists⁴⁴ or persistence of asthma⁴⁵ showed that the changes in PC₂₀ values following salmeterol monotherapy were not related to these polymorphisms. Thus, although our study is clearly underpowered to exclude other detrimental effects of β_2 -receptor polymorphisms on asthma control, the observed effects on PC₂₀ values do not appear to be related to these polymorphisms. This is in line with a recent analysis suggesting that β_2 -receptor polymorphisms do not affect the therapeutic response to LABA in patients with asthma⁴⁶. Our observation that the majority of patients with mild asthma developed an increase in AHR following salmeterol monotherapy suggests a susceptible subpopulation of patients which cannot be identified by β_2 -receptor polymorphisms or other clinical characteristics obtained in our study.

One limitation of this study is the lack of a crossover design. However, this was omitted due to ethical and safety concerns. A study arm in which patients would have left the study after regular monotherapy with a β_2 -agonist (with possibly detrimental effects on asthma control and mortality) was considered ethically inappropriate based on the strong recommendations in international guidelines that LABA should not be used as monotherapy in asthma². Thus, although this may appear overcautious in patients with mild asthma, a crossover design was rejected. This decision has been confirmed in part by our results which showed a deterioration of AHR and mild adverse effects in a substantial portion of the participants following salmeterol monotherapy. Another potential pitfall in the design of our study is the lack of a placebo arm. However, although this might have at least theoretically improved the study, it was prospectively decided that this approach would not offer advantages because the high dose of the β_2 -agonist (100 μ g of salmeterol b.i.d.) unblinds such a design due to its inherent side effects. Furthermore, in our patient population, baseline parameters are representative of a placebo arm since baseline treatment (short-acting bronchodilators on an as needed basis) was maintained unchanged throughout the study.

In conclusion, we show that unbalanced monotherapy with salmeterol in patients with mild asthma increases BDNF production and storage and that changes in AHR are associated with this effect. We, therefore, hypothesise that augmented BDNF concentrations explain some of the adverse effects of β_2 -agonists in asthma. Further studies in patients with more severe airflow obstruction are warranted to confirm these findings.

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COMPETING INTERESTS

The authors have no competing interests to declare.

SPONSOR DETAILS

University of Rostock and GlaxoSmithKline (GSK).

ETHICS APPROVAL

The study was approved by the local ethics committee of Rostock, Germany.

LICENCE FOR PUBLICATION

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FIGURE LEGENDS

Fig. 1. Study design

Body plethysmography, assessment of airway responsiveness to histamine and blood collection for BDNF measurements were performed prior to entry (white box), after 14 days of salmeterol therapy (light grey box) and after 14 days of combination therapy (dark grey box). *Abbreviations denote:* Forced expiratory volume in the first one second (FEV₁) in % of the predicted value (% pred.), provocative concentration of histamine causing a 20% fall in FEV₁ (PC₂₀).

Fig. 2. Lung function

Shown are FEV₁ (% predicted) values (A) and PEF (% predicted) values (B) prior to entry (white box), after 14 days of salmeterol therapy (light grey box) and after 14 days of combination therapy (dark grey box) of n = 18 patients with allergic asthma. Boxplot graphs display the median (line within the box), interquartile range (edges of the box) and the range of all values less distant than 1.5 interquartile ranges from the upper or lower quartile (vertical lines). S denotes salmeterol, F denotes fluticasone.

Fig. 3. Airway responsiveness to histamine

PC₂₀ values are shown for each patient (n = 18 patients) prior to entry (Baseline), after 14 days of salmeterol therapy and after 14 days of combination therapy. Patients with a decrease in PC₂₀ values after 14 days of salmeterol therapy are displayed with continuous lines. Patients with an increase in PC₂₀ values after 14 days of salmeterol therapy are displayed with dashed lines. S denotes salmeterol, F denotes fluticasone.

Fig. 4. BDNF concentrations in serum and platelets

Shown are BDNF concentrations in serum (ng BDNF / ml serum) and platelets (pg BDNF / 10⁶ platelets) prior to entry (white box), after 14 days of salmeterol therapy (light grey box) and after 14 days of combination therapy (dark grey box) of n = 18 patients with allergic asthma. Boxplot graphs display the median (line within the box), interquartile range (edges of the box) and the range of all values less distant than 1.5 interquartile ranges from the upper or lower quartile (vertical lines). S denotes salmeterol, F denotes fluticasone.

Fig. 5. Association of BDNF with changes in PC₂₀

Shown are correlations between changes in BDNF concentrations in serum (A) or platelets (B) and changes in the PC₂₀ values (histamine) after 14 days of salmeterol therapy, as compared to the baseline before therapy. Each dot represents one patient, the line is the regression line calculated with SPSS. Spearman's rank correlation coefficient (r) and the significance of the correlation (p) are given above each graph.

Fig. 6. BDNF release by leukocytes *in vitro*

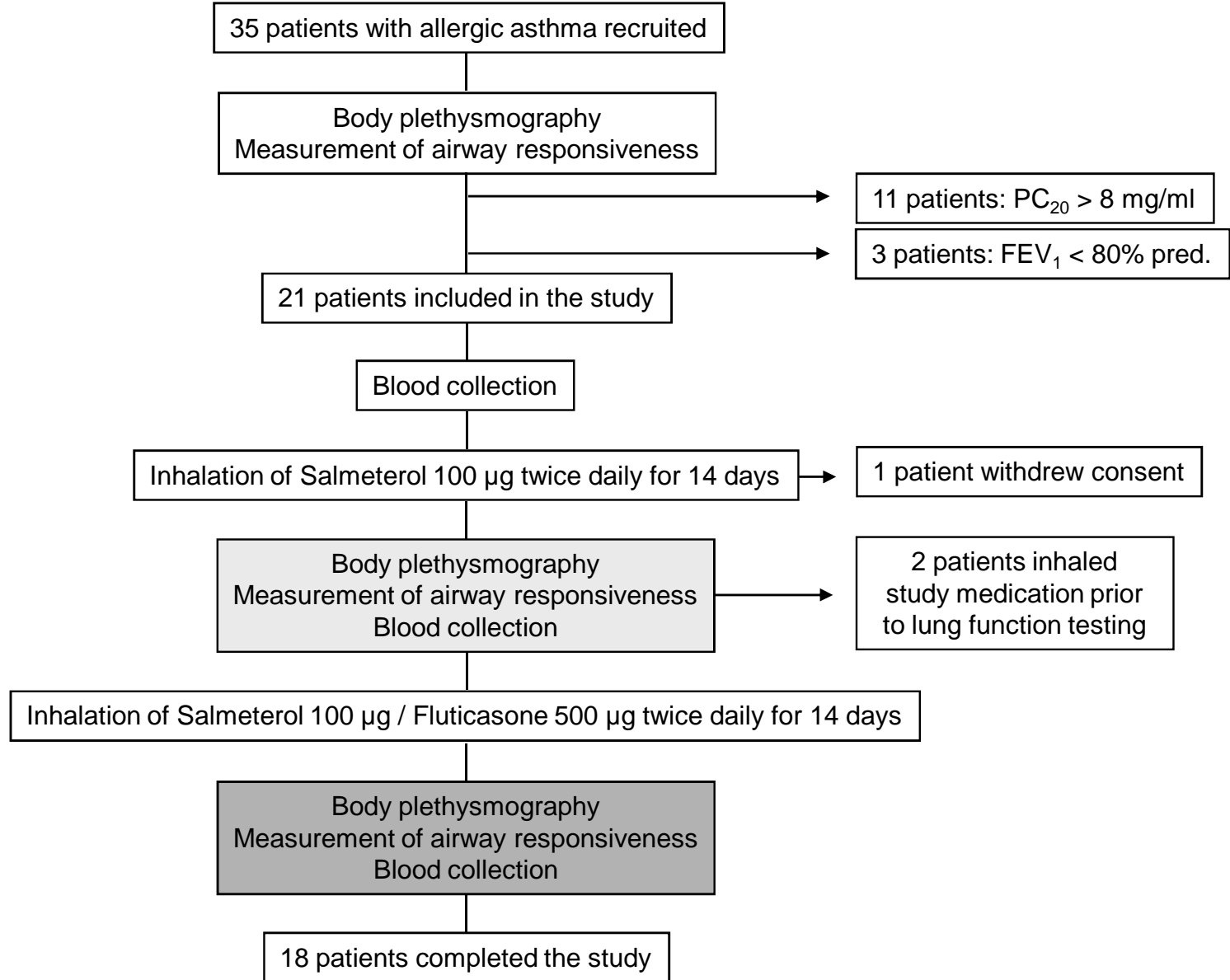
Monocyte-enriched human peripheral blood mononuclear cells of n = 22 healthy volunteers were stimulated with Tumor necrosis factor α (TNF- α , 50 ng/ml) for 24 hours. Shown are BDNF concentrations (mean \pm standard deviation) in supernatants of wells containing fluticasone propionate (10⁻⁷ M), salmeterol xinafoate (10⁻⁷ M) and both fluticasone propionate (10⁻⁷ M) and salmeterol xinafoate (10⁻⁷ M), as compared to BDNF concentrations in the medium control.

REFERENCES

1. **Tattersfield AE**, Knox AJ, Britton JR, *et al.* Asthma, *Lancet* 2002, **360**:1313-1322
2. **Bateman ED**, Hurd SS, Barnes PJ, *et al.* Global strategy for asthma management and prevention: GINA executive summary, *Eur Respir J* 2008, **31**:143-178
3. **Hasford J**, Virchow JC. Excess mortality in patients with asthma on long-acting beta2-agonists, *Eur Respir J* 2006, **28**:900-902
4. **Taylor DR**, Sears MR, Herbison GP, *et al.* Regular inhaled beta agonist in asthma: effects on exacerbations and lung function, *Thorax* 1993, **48**:134-138
5. **Crane J**, Pearce N, Flatt A, *et al.* Prescribed fenoterol and death from asthma in New Zealand, 1981-83: case-control study, *Lancet* 1989, **333**: 917-922
6. **Inman WH**, Adelstein AM. Rise and fall of asthma mortality in England and Wales in relation to use of pressurised aerosols, *Lancet* 1969, **294**: 279-285
7. **Spitzer WO**, Suissa S, Ernst P, *et al.* The use of beta-agonists and the risk of death and near death from asthma, *N Engl J Med* 1992, **326**:501-506
8. **Mann M**, Chowdhury B, Sullivan E, *et al.* Serious asthma exacerbations in asthmatics treated with high-dose formoterol, *Chest* 2003, **124**:70-74
9. **Drazen JM**, Israel E, Boushey HA, *et al.* Comparison of regularly scheduled with as-needed use of albuterol in mild asthma. Asthma Clinical Research Network, *N Engl J Med* 1996, **335**:841-847
10. **Kraan J**, Koeter GH, vd Mark TW, *et al.* Changes in bronchial hyperreactivity induced by 4 weeks of treatment with antiasthmatic drugs in patients with allergic asthma: a comparison between budesonide and terbutaline, *J Allergy Clin Immunol* 1985, **76**:628-636
11. **van Schayck CP**, Graafsma SJ, Visch MB, *et al.* Increased bronchial hyperresponsiveness after inhaling salbutamol during 1 year is not caused by subsensitization to salbutamol, *J Allergy Clin Immunol* 1990, **86**:793-800
12. **Sears MR**: Adverse effects of beta-agonists, *J Allergy Clin Immunol* 2002, **110**:S322-328
13. **Cockcroft DW**, McParland CP, Britto SA, *et al.* Regular inhaled salbutamol and airway responsiveness to allergen, *Lancet* 1993, **342**:833-837
14. **Gauvreau GM**, Jordana M, Watson RM, *et al.* Effect of regular inhaled albuterol on allergen-induced late responses and sputum eosinophils in asthmatic subjects, *Am J Respir Crit Care Med* 1997, **156**:1738-1745
15. **Simons FE**. A comparison of beclomethasone, salmeterol, and placebo in children with asthma. Canadian Beclomethasone Dipropionate-Salmeterol Xinafoate Study Group, *N Engl J Med* 1997, **337**:1659-1665
16. **Verberne AA**, Frost C, Roorda RJ, *et al.* One year treatment with salmeterol compared with beclomethasone in children with asthma. The Dutch Paediatric Asthma Study Group, *Am J Respir Crit Care Med* 1997, **156**:688-695
17. **O'Connor BJ**, Aikman SL, Barnes PJ. Tolerance to the nonbronchodilator effects of inhaled beta 2-agonists in asthma, *N Engl J Med* 1992, **327**:1204-1208
18. **Cheung D**, Timmers MC, Zwinderman AH, *et al.* Long-term effects of a long-acting beta 2-adrenoceptor agonist, salmeterol, on airway hyperresponsiveness in patients with mild asthma, *N Engl J Med* 1992, **327**:1198-1203
19. **Castle W**, Fuller R, Hall J, *et al.* Serevent nationwide surveillance study: comparison of salmeterol with salbutamol in asthmatic patients who require regular bronchodilator treatment, *BMJ* 1993, **306**:1034-1037

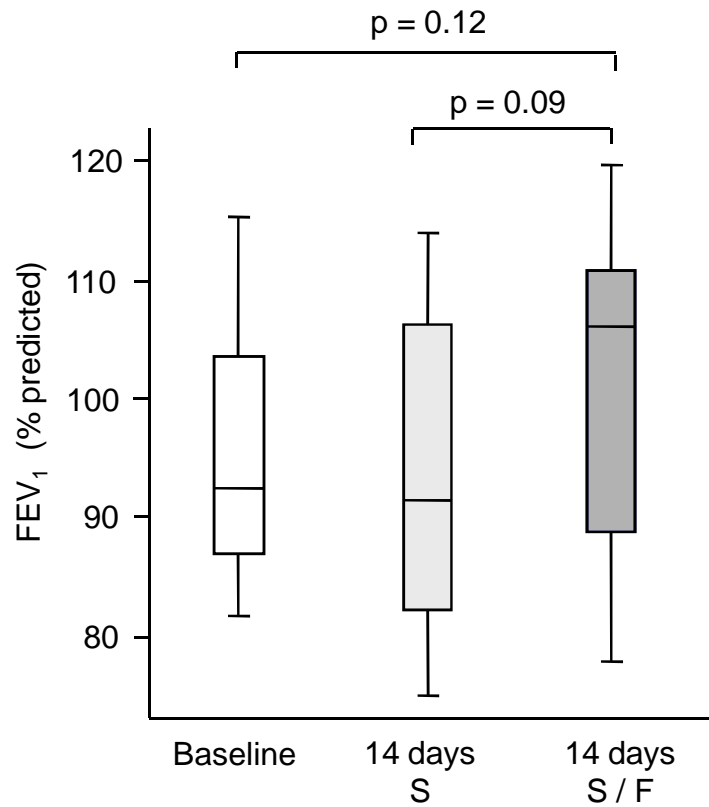
20. **Nelson HS**, Weiss ST, Bleecker ER, *et al.* The Salmeterol Multicenter Asthma Research Trial: a comparison of usual pharmacotherapy for asthma or usual pharmacotherapy plus salmeterol, *Chest* 2006, **129**:15-26
21. **Sears MR**, Ottosson A, Radner F, *et al.* Long-acting beta-agonists: a review of formoterol safety data from asthma clinical trials, *Eur Respir J* 2009, **33**:21-32
22. **Huang EJ**, Reichardt LF. Neurotrophins: roles in neuronal development and function, *Annu Rev Neurosci* 2001, **24**:677-736
23. **Braun A**, Lommatzsch M, Mannsfeldt A, *et al.* Cellular sources of enhanced brain-derived neurotrophic factor production in a mouse model of allergic inflammation, *Am J Respir Cell Mol Biol* 1999, **21**:537-546
24. **Lommatzsch M**, Braun A, Renz H. Neurotrophins in allergic airway dysfunction: what the mouse model is teaching us, *Ann N Y Acad Sci* 2003, **992**:241-249
25. **Braun A**, Lommatzsch M, Neuhaus-Steinmetz U, *et al.* Brain-derived neurotrophic factor (BDNF) contributes to neuronal dysfunction in a model of allergic airway inflammation, *Br J Pharmacol* 2004, **141**:431-440
26. **Hahn C**, Islamian AP, Renz H, *et al.* Airway epithelial cells produce neurotrophins and promote the survival of eosinophils during allergic airway inflammation, *J Allergy Clin Immunol* 2006, **117**:787-794
27. **Lommatzsch M**, Schloetcke K, Klotz J, *et al.* Brain-derived neurotrophic factor in platelets and airflow limitation in asthma, *Am J Respir Crit Care Med* 2005, **171**:115-120
28. **Virchow JC**, Julius P, Lommatzsch M, *et al.* Neurotrophins are increased in bronchoalveolar lavage fluid after segmental allergen provocation, *Am J Respir Crit Care Med* 1998, **158**:2002-2005
29. **Dolovich J**, Hargreave FE, Jordana M, *et al.* Late-phase airway reaction and inflammation, *J Allergy Clin Immunol* 1989, **83**:521-524
30. **Lommatzsch M**, Klotz J, Virchow JC. Postnatal dexamethasone for lung disease of prematurity, *N Engl J Med* 2004, **350**:2715-2718.
31. **Noga O**, Hanf G, Schaper C, *et al.* The influence of inhalative corticosteroids on circulating Nerve Growth Factor, Brain-Derived Neurotrophic Factor and Neurotrophin-3 in allergic asthmatics, *Clin Exp Allergy* 2001, **31**:1906-1912
32. **Lommatzsch M**, Zingler D, Schuhbaeck K, *et al.* The impact of age, weight and gender on BDNF levels in human platelets and plasma, *Neurobiol Aging* 2005, **26**:115-123
33. **Pauwels RA**, Lofdahl CG, Postma DS, *et al.* Effect of inhaled formoterol and budesonide on exacerbations of asthma. Formoterol and Corticosteroids Establishing Therapy (FACET) International Study Group, *N Engl J Med* 1997, **337**:1405-1411
34. **Bateman ED**, Boushey HA, Bousquet J, *et al.* Can guideline-defined asthma control be achieved? The Gaining Optimal Asthma Control study, *Am J Respir Crit Care Med* 2004, **170**:836-844
35. **Cockcroft DW**, Davis BE. Mechanisms of airway hyperresponsiveness, *J Allergy Clin Immunol* 2006, **118**:551-559
36. **Keir S**, Page C, Spina D. Bronchial hyperresponsiveness induced by chronic treatment with albuterol: Role of sensory nerves, *J Allergy Clin Immunol* 2002, **110**:388-394
37. **Zaidi SI**, Jafri A, Doggett T, *et al.* Airway-related vagal preganglionic neurons express brain-derived neurotrophic factor and TrkB receptors: implications for neuronal plasticity, *Brain Res* 2005, **1044**:133-143
38. **Benedich Kahn L**, Gustafsson LE, Olgart Hoglund C. Brain-derived neurotrophic factor enhances histamine-induced airway responses and changes levels of exhaled nitric oxide in guinea pigs in vivo, *Eur J Pharmacol* 2008, **595**:78-83

39. **Fujimura H**, Altar CA, Chen R, *et al.* Brain-derived neurotrophic factor is stored in human platelets and released by agonist stimulation, *Thromb Haemost* 2002, **87**:728-734
40. **Pitchford SC**, Momi S, Baglioni S, *et al.* Allergen induces the migration of platelets to lung tissue in allergic asthma, *Am J Respir Crit Care Med* 2008, **177**:604-612
41. **Pitchford SC**, Riffo-Vasquez Y, Sousa A, *et al.* Platelets are necessary for airway wall remodeling in a murine model of chronic allergic inflammation, *Blood* 2004, **103**:639-647
42. **Edwards MR**, Haas J, Panettieri RA, *et al.* Corticosteroids and beta2 agonists differentially regulate rhinovirus-induced interleukin-6 via distinct Cis-acting elements, *J Biol Chem* 2007, **282**:15366-15375
43. **Shieh PB**, Ghosh A. Molecular mechanisms underlying activity-dependent regulation of BDNF expression, *J Neurobiol* 1999, **41**:127-134
44. **Wechsler ME**, Lehman E, Lazarus SC, *et al.* beta-Adrenergic receptor polymorphisms and response to salmeterol, *Am J Respir Crit Care Med* 2006, **173**:519-526
45. **Hall IP**, Blakey JD, Al Balushi KA, *et al.* Beta2-adrenoceptor polymorphisms and asthma from childhood to middle age in the British 1958 birth cohort: a genetic association study, *Lancet* 2006, **368**:771-779
46. **Blecker ER**, Postma DS, Lawrance RM, *et al.* Effect of ADRB2 polymorphisms on response to longacting beta2-agonist therapy: a pharmacogenetic analysis of two randomised studies, *Lancet* 2007, **370**:2118-2125

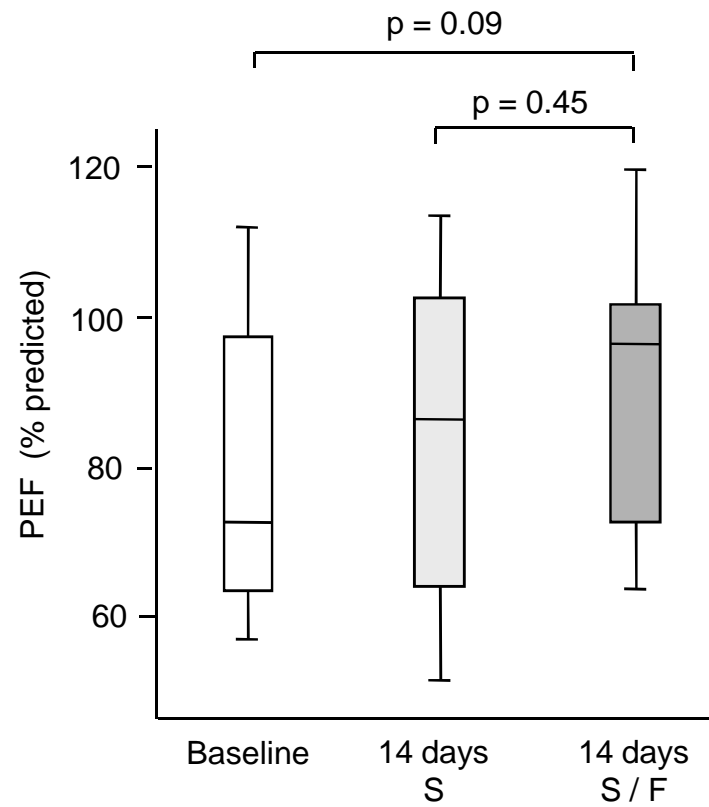


Lommatzsch et al., Fig. 1

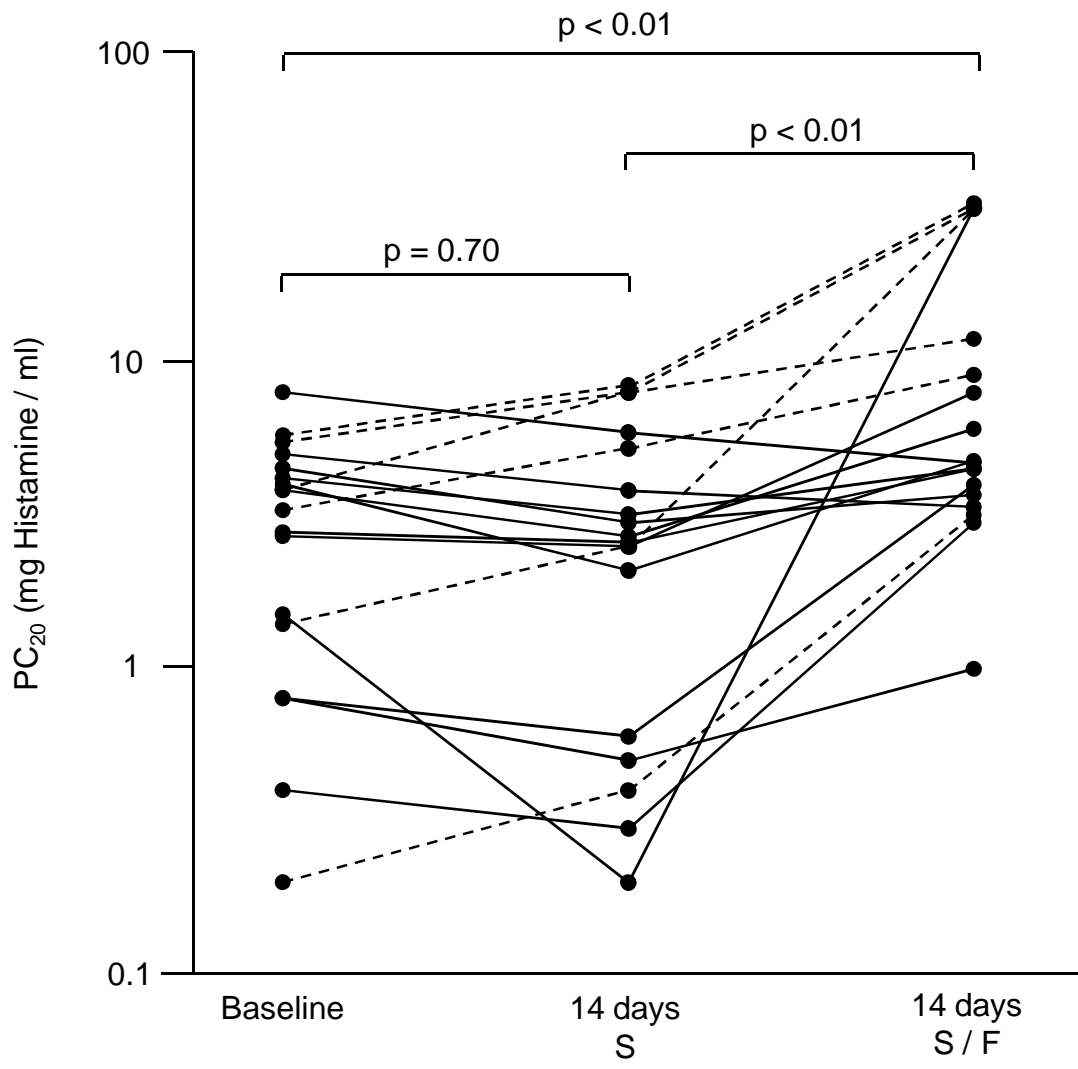
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B

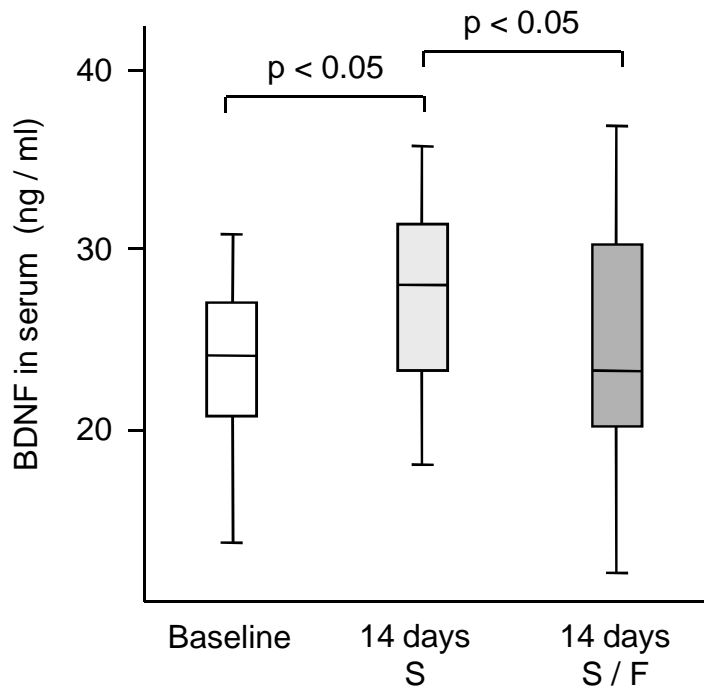


Lommatzsch et al., Fig. 2

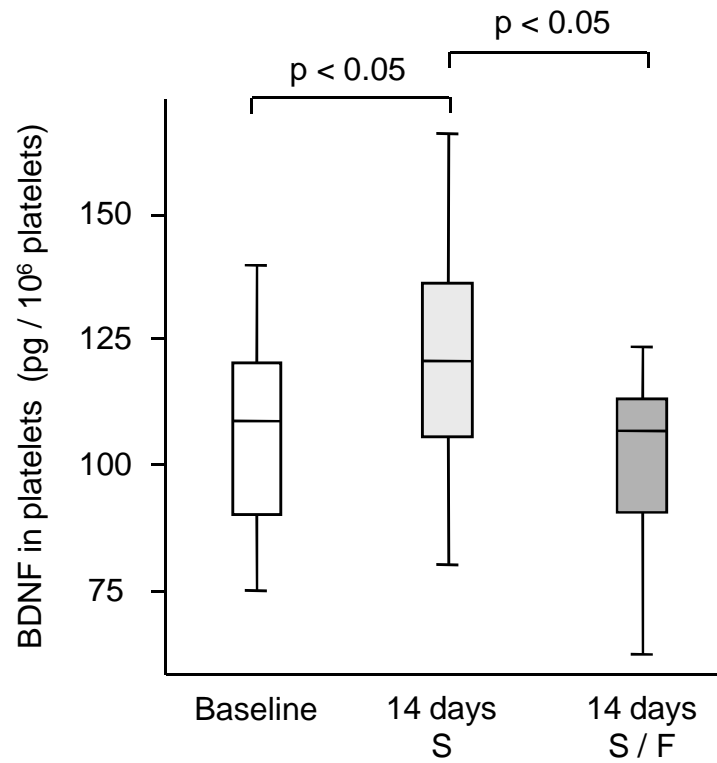


Lommatzsch et al., Fig. 3

A



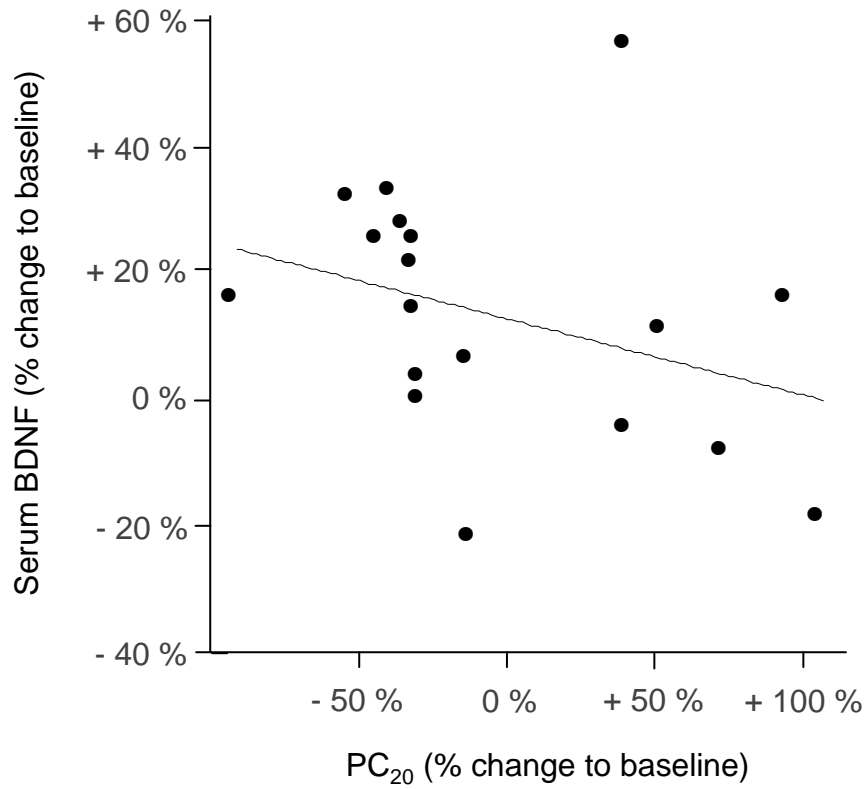
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Lommatzsch et al., Fig. 4

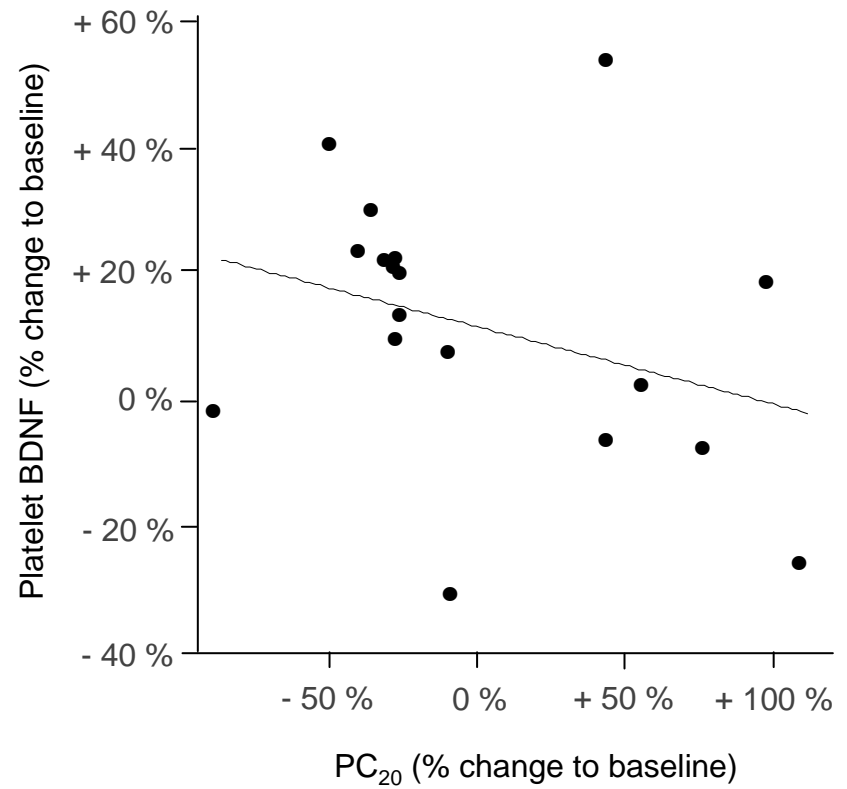
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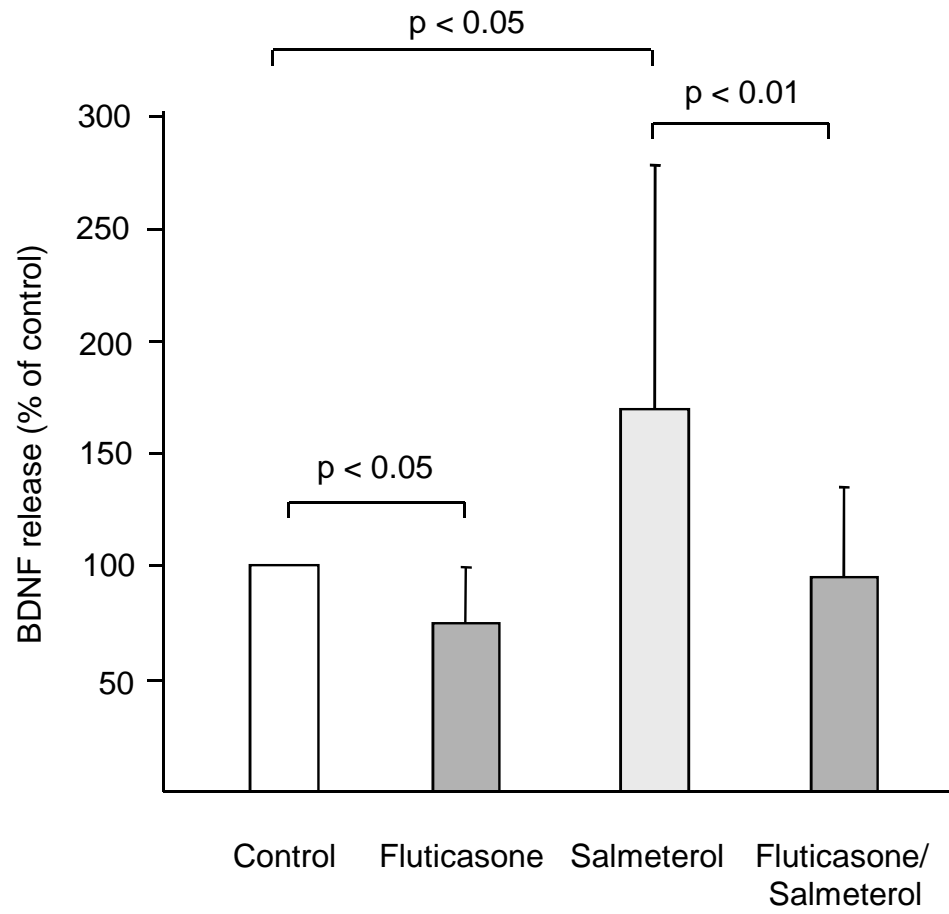
$r = -0.61, p < 0.01$



B

$r = -0.53, p < 0.05$





Lommatzsch et al., Fig. 6