Anti-inflammatory effects of PGE$_2$ in the lung: role of the EP$_4$ receptor subtype

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**ABSTRACT**

**Background** Asthma and chronic obstructive pulmonary disease (COPD) are chronic inflammatory diseases of the airway. Current treatment options (long acting β-adrenergic agonists and glucocorticosteroids) are not optimal as they are only effective in certain patient groups and safety concerns exist regarding both compound classes. Therefore, novel bronchodilator and anti-inflammatory strategies are being pursued. Prostaglandin E$_2$ (PGE$_2$) is an arachidonic acid-derived eicosanoid produced by the lung which acts on four different G-protein coupled receptors (EP$_1$–EP$_4$) to cause an array of beneficial and deleterious effects. The aim of this study was to identify the EP receptor mediating the anti-inflammatory actions of PGE$_2$ in the lung using a range of cell-based assays and in vivo models.

**Methods and results** It was demonstrated in three distinct model systems (innate stimulus, lipopolysaccharide (LPS); allergic response, ovalbumin (OVA); inhaled pollutant, cigarette smoke) that mice lacking functional EP$_4$ (Ptger4$^{-/-}$) receptors had higher levels of airway inflammation, suggesting that endogenous PGE$_2$ was suppressing inflammation via EP$_4$ receptor activation. Cell-based assay systems (murine and human monocytes/alveolar macrophages) demonstrated that PGE$_2$ inhibited cytokine release from LPS-stimulated cells and that this was mimicked by an EP$_4$ (but not EP$_1$–EP$_3$) receptor agonist and inhibited by an EP$_4$ receptor antagonist. The anti-inflammatory effect occurred at the transcriptional level and was via the adenylyl cyclase/cAMP/ cAMP-dependent protein kinase (PKA) axis.

**Conclusion** This study demonstrates that EP$_4$ receptor activation is responsible for the anti-inflammatory activity of PGE$_2$ in a range of disease relevant models and, as such, could represent a novel therapeutic target for chronic airway inflammatory conditions.

**INTRODUCTION**

Asthma and chronic obstructive pulmonary disease (COPD) are respiratory diseases with an increasing global prevalence that represent a social and economic burden for both industrialised and developing countries. The World Health Organization states that the number of patients with asthma is 300 million and predicts that this figure will rise to 400 million by 2025,$^1$ while there are 600 million people with COPD worldwide and the disease is predicted to be the third ranked leading cause of death by 2020.$^5$ Exacerbations are common events in the lives of patients with asthma and COPD. These episodes are often associated with infections by viruses or bacteria and cause worsening of symptoms, which can be fatal. Often these heightened symptoms are far less responsive to normal treatments and are associated with increased healthcare costs and societal impact. There is therefore an urgent need to develop safe and effective therapies for these respiratory diseases.

It has been recognised for many years that prostaglandin E$_2$ (PGE$_2$) is produced by many cell types within the lung.$^3$ Furthermore, exogenous PGE$_2$ has been shown to have bronchodilator and anti-inflammatory properties in rodent models$^5$ and human subjects.$^3$ Thus, PGE$_2$ could represent an effective treatment, but it is associated with airway irritation and coughing$^6$ and has a very short half-life and, as such, has an inappropriate pharmacokinetic profile for general clinical use.

The biological actions of PGE$_2$ are predominantly mediated by the activation of the four EP receptors, EP$_1$–EP$_4$. We and others have recently shown that the bronchodilator properties of PGE$_2$...
in human airways are mediated via activation of the EP4 receptor,\(^8\) whereas the undesirable triggering of airway sensory nerves appears to be via its activity at the EP2 receptor.\(^9\) However, it is still not clear which receptor is involved in mediating the anti-inflammatory effects of PGE\(_2\). Identification of a receptor mediating the anti-inflammatory and bronchodilator effects of PGE\(_2\) without the airway irritant activity would harness the beneficial effects while avoiding the undesirable properties.

To achieve this aim we profiled the inflammatory status of EP receptor knockout (KO) mice in an array of preclinical respiratory disease model systems (endotoxin: innate; allergen: asthma-like; and smoke: COPD-like). To investigate the signalling mechanisms involved, we used in vitro murine and human cell-based assay systems. In vitro assays confirmed the in vivo data which suggested that EP\(_4\) receptor activation is responsible for the anti-inflammatory activity of PGE\(_2\) and that the effect was at the level of cytokine gene transcription via an adenylyl cyclase (AC)/cAMP/cAMP-dependent protein kinase (PKA) axis.

**MATERIALS AND METHODS**

**Role of EP receptors in murine respiratory models**

Breeding pairs of C57bl/6 wild type and EP receptor gene (Ptger1–Ptger4) KO mice were purchased from Harlan (Bicester, Oxon, UK) and provided by Dr Shuh Narumiya, Kyoto University, respectively. Ptger1–Ptger3 KO mice were backcrossed on the C57bl/6 at least eight times. Ptger4\(^--\)/\(^--\) mice do not survive on the C57bl/6 background\(^10\) and were backcrossed on a mixed background of 129/Ola\(\times\)C57bl/6. Corresponding wild type colonies were generated. Male mice (18–20 g) were generated from established breeding colonies maintained internally. Experiments were performed under Home Office project licence (PPL 70/7212) and procedures adhered to the Animals (Scientific Procedures) Act 1986 and according to the ARRIVE guidelines.\(^11\)

Mice were challenged with inhaled lipopolysaccharide (LPS) (innate response), ovalbumin (OVA) (allergic: asthma-like) or cigarette smoke (COPD-like). To determine if this phenomenon was due to the genetic background of the mice, we assessed EP receptor KO mice in a range of preclinical respiratory disease model systems (endotoxin: innate; allergen: asthma-like; and smoke: COPD-like). To investigate the signalling mechanisms involved, we used in vitro murine and human cell-based assay systems. In vitro assays confirmed the in vivo data which suggested that EP\(_4\) receptor activation is responsible for the anti-inflammatory activity of PGE\(_2\) and that the effect was at the level of cytokine gene transcription via an adenylyl cyclase (AC)/cAMP/cAMP-dependent protein kinase (PKA) axis.

**RESULTS**

**Role of EP receptors in murine respiratory models of airway inflammation**

Wild type mice challenged with the various disease relevant stimuli evoked the expected inflammatory responses associated with increases in a particular cell type. In particular, increases in airway neutrophils were observed in the LPS and cigarette smoke models and eosinophils after antigen challenge (figure 1). In the LPS and allergen models, inflammatory cell infiltration was significantly increased in the EP\(_4\) receptor (Ptger4\(^--\)/\(^--\)) KO mice (with no change in EP1\(_3\), EP2 KO mice) compared with the wild type control, suggesting a protective role for EP\(_4\) receptor activation (figure 1). To determine if this phenomenon was due to the genetic background of the mice, we assessed EP receptor KO mice in a range of preclinical respiratory disease model systems (endotoxin: innate; allergen: asthma-like; and smoke: COPD-like). To investigate the signalling mechanisms involved, we used in vitro murine and human cell-based assay systems. In vitro assays confirmed the in vivo data which suggested that EP\(_4\) receptor activation is responsible for the anti-inflammatory activity of PGE\(_2\) and that the effect was at the level of cytokine gene transcription via an adenylyl cyclase (AC)/cAMP/cAMP-dependent protein kinase (PKA) axis.

**Data analysis and statistics**

Data are presented as mean±SEM and statistical significance was assumed when p<0.05. N numbers and statistical tests used were the Mann–Whitney test when two datasets were compared and Kruskal–Wallis ANOVA followed by Dunn’s multiple comparison for three or more datasets. The specific test used is indicated in the figure legends.

**Anti-inflammatory role of EP receptors in cell-based assay systems**

In three different disease relevant in vivo models of airway inflammation we found that the inflammatory response was increased in EP\(_4\) receptor KO mice, suggesting an anti-inflammatory role for endogenous prostaglandins via activation...
of EPα receptors. In order to perform more in-depth investigations using pharmacological tools without the appropriate pharmacokinetic profile for in vivo studies and in order to provide translational data, we performed studies in human cell-based assays. To investigate the anti-inflammatory potential of PGE2, we examined the effect of exogenously added PGE2 on

Figure 1  Role of EP receptors in murine respiratory models. (A, B) Eosinophils in lavage fluid from mice sensitised and challenged with ovalbumin (OVA), wild type controls versus EP1–3 knockout (KO) mice (A) or wild type controls versus EP3 KO mice (B). (C, D) Neutrophils in lavage fluid from mice challenged with lipopolysaccharide (LPS), wild type controls versus EP1–3 KO mice (C) or wild type controls versus EP2 KO mice (D). (E) Neutrophils in lavage fluid from mice exposed to cigarette smoke, wild type controls versus EP4 KO mice. Data shown are mean±SEM (n=8). #Statistical significance between control challenge and stimulation (p<0.05, Mann–Whitney test). *Statistical significant between wild type and KO mice (p<0.05, Mann–Whitney test).

Figure 2  Effect of prostaglandin E2 (PGE2) on cytokine release in cell-based assays. Effect of increasing concentrations of PGE2 on levels of tumour necrosis factor α (TNFα) (A, C) and interleukin 6 (IL-6) (B, D) in culture medium from mouse (J774; A, B) and human (THP-1; C, D) THP-1 cells stimulated with vehicle (saline) or lipopolysaccharide (LPS; 0.1 μg/mL). Data shown are mean±SEM of three experimental runs performed in duplicate. #Statistical significance between vehicle and stimulation (p<0.05, Mann–Whitney test). *Statistical significance between stimulation and PGE2-treated cells (p<0.05, Kruskal–Wallis ANOVA followed by Dunn’s multiple comparison test).
inflammatory responses to one of the stimuli used in the murine model, namely LPS. All experiments were performed in the presence of a broad spectrum cyclo-oxygenase inhibitor to block endogenous prostanoid production which may complicate data interpretation. PGE₂ produced a concentration-related inhibition of LPS-induced cytokine production in mouse and human monocytes (figure 2). To investigate which receptor(s) were central to the anti-inflammatory effect seen with PGE₂, we repeated the study using selective EP agonists and found that only the EP₄ receptor agonist (ONO-AE1-329) modulated cytokine levels at concentrations commensurate with the reported nanomolar potency of this ligand at the EP₄ receptor (figure 3D, H). In a parallel experiment, the same EP receptor agonists did not trigger cytokine release under non-stimulated conditions (see online supplementary figure S3). Interestingly, EP₄ receptor mRNA expression was the predominant EP receptor subtype expressed at the mRNA level in these cells and primary human alveolar macrophages (see online supplementary figure S4). These data suggest that the EP receptor expression profile and the predominance of EP₄ explains why the anti-inflammatory activity of PGE₂ is mediated via the EP₄ receptor.

To confirm that the EP₄ receptor is central to the anti-inflammatory properties of PGE₂, we performed experiments in the presence of selective EP₂ and EP₄ receptor antagonists (PF-04418948 and ONO-AE3-208, respectively) and focused on tumour necrosis factor α (TNFα) production as an example cytokine. The data clearly show that the EP₄ receptor antagonist blocks the anti-inflammatory activity of both the selective EP₄ receptor agonist and PGE₂ itself (figure 4C, D), with no effect of the EP₂ receptor antagonist.

In order to investigate whether the production of TNFα was regulated at the transcriptional or translational level, we had to...
determine a suitable time point to measure mRNA production. The 2 h time point was selected to measure mRNA and protein production was assessed 22 h after stimulation (figure 5A). The EP4 receptor agonist modulated both the mRNA and protein levels, which suggests that activation of the receptor modulates cytokine production at the transcriptional level (figure 5C, D).

To explore the signalling mechanisms involved, we investigated the effects of mimicking proposed EP4 receptor signalling involving the AC-PKA/EPAC pathways. The AC activator, but not its negative control (1,9 dideoxyforskolin, which shares many of the activities of forskolin but does not activate AC), caused a concentration-related inhibition of cytokine production which was of a similar magnitude to the EP4 receptor agonist (figure 6A, B). Similarly, an activator of PKA, but not EPAC, mirrored the effect of the EP4 receptor agonist (figure 6C, D). In order to determine if this proposed pathway was indeed triggered by activating the EP4 receptor, we employed a selective PKA blocker. The PKA inhibitor caused a concentration-related block of the anti-inflammatory properties of both a PKA activator and the EP4 receptor agonist (figure 7A, B). Finally, to demonstrate this

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**Figure 4** Effect of selective EP receptor antagonists on inhibitory effect of prostaglandin E2 (PGE2) and EP4 agonist-induced cytokine release. Effect of EP2 receptor agonist (ONO-AE1-329, 1 nM) (A, C) or PGE2 (1 nM) (B, D) in the presence of increasing concentrations of EP2 (PF-04418948) or EP4 (ONO-AE3-208) receptor antagonists on tumour necrosis factor α (TNFα) levels in culture medium from human (THP-1) monocytes stimulated with vehicle (saline) or lipopolysaccharide (LPS; 0.1 μg/mL). Data shown are mean±SEM of three experimental runs performed in duplicate. #Statistical significance between vehicle and stimulation (p<0.05, Mann–Whitney test). *Statistical significance between stimulation and agonist treated cells (p<0.05, Kruskal–Wallis ANOVA followed by Dunn’s multiple comparison test).

**Figure 5** Assessing the effect of EP4 receptor activation on transcription and translation of tumour necrosis factor α (TNFα). TNFα mRNA (A) and protein production (B) following treatment with vehicle (saline) or lipopolysaccharide (LPS; 0.1 μg/mL) over time in the culture medium of human (THP-1) monocytes. (C) TNFα mRNA measured 2 h post stimulation and (D) protein measured 22 h post stimulation in the presence of EP4 receptor agonist (ONO-AE1-329). Data shown are mean±SEM of three experimental runs performed in duplicate. #Statistical significance between vehicle and stimulation (p<0.05, Mann–Whitney test). *Statistical significance between stimulation and agonist treated cells (p<0.05, Mann–Whitney test).
agonist treated cells (p<0.05, Kruskal–Wallis ANOVA followed by Dunn’s multiple comparison test).

pathway was active in primary cells, we repeated the observation in human alveolar macrophages (figure 7C).

**DISCUSSION**

Current anti-inflammatory therapy for asthma and COPD is focused on the use of glucocorticoids, but their efficacy is limited in COPD and in certain patient subgroups with asthma. Furthermore, there are significant patient compliance issues and safety concerns associated with their long-term use. Existing treatment of both diseases is focused on combination therapies which include a bronchodilator together with the glucocorticoid and undesirable properties associated with PGE2 to identify a receptor subtype which could be targeted for the development of a therapeutic option, especially if it could be developed as an oral medication to avoid compliance issues.

The beneficial properties of PGE2 in the lung have long been recognised, but irritancy issues and the poor pharmacokinetic profile of endogenous bioactive lipid mediators mean that these properties are yet to be fully harnessed. Previous studies have started to define the EP receptors associated with the beneficial and undesirable properties associated with PGE2 to identify a receptor subtype which could be targeted for the development of a therapeutic. For example, EP4 receptor activation has been proposed to be involved in PGE2-induced bronchodilation of human airway smooth muscle and EP4 receptor activation to trigger airway sensory nerves producing unwanted reflexes such as cough. The aim of this study was to begin to explore which receptor(s) is required for the airway anti-inflammatory properties of PGE2.

The initial body of work was performed in three in vivo murine models with distinct phenotypes (modelling innate, allergic and COPD-like inflammation) and demonstrated that EP4 receptor KO mice had an enhanced cellular inflammation, suggesting an endogenous anti-inflammatory role for PGE2 acting on the EP4 receptor. Supporting these data was the observation that EP4 receptor gene expression was increased in the lungs after disease relevant challenges in standard (C57BL/6) wild type mice, suggesting that this observation was not specific to the genetic background of the mice used in the EP4 comparator studies. Consistent with these findings are the facts that most immune cell types have been reported to express EP4 receptors, and EP4 receptor activation reduces transendothelial migration of eosinophils and neutrophils and induces suppression in a murine model of aspirin-triggered allergic hyperresponsiveness. In contrast to these observations, a protective role for EP4 receptor activation has been reported in a murine asthma model. Although a beneficial role for the EP3 receptor is not supported in studies demonstrating that PGE2—but not the EP3 receptor agonist sulprostone—exhibits anti-inflammatory effects in an allergic murine model, and in other studies suggesting that EP3 receptor activation is chemotactic for lung mast cells. Alternatively, other studies have reported that EP3 receptor activation inhibits asthma-related eosinophilic trafficking. Although studies exist supporting a role for alternative receptors in mediating the anti-inflammatory effects of PGE2, often the study investigators have not had the opportunity to use the wide range of selective pharmacological tools and KO mice that we have been able to employ here. Furthermore, the majority of the published literature regarding the anti-inflammatory role of PGE2 is in agreement with our data suggesting a role for the EP4 receptor.

To provide further confirmatory data regarding the role of the EP4 receptor and to investigate the downstream signalling pathway, we used cell-based systems which are more amenable to study and where data interpretation is not limited by the pharmacokinetic limitations of tool compounds. This approach also enabled translation of findings in murine systems to human cells. Monocyte/macrophage cell types were used as the prevalent cell in the airway and the cell type most likely to have a common role in all three inflammatory responses that were modelled in the mouse. Furthermore, previous studies had demonstrated EP4 receptor gene expression in murine macrophages, the murine monocyte cell line J774 cells and human lung macrophages. In these experiments, using one of the stimuli used in the model systems (endoxin, LPS), PGE2 also had an anti-inflammatory role demonstrated as the inhibition of

![Figure 6](https://example.com/figure6.png)  
EP4 receptor activation and downstream signalling pathways. The effect of vehicle, adenylyl cyclase (AC) activator (A), AC activator negative control (B), cAMP-dependent protein kinase (PKA) activator (C), or exchange proteins activated by cAMP (EPAC) activator (D) on tumour necrosis factorα (TNFα) release from human (THP-1) monocytes stimulated with vehicle (saline) or lipopolysaccharide (LPS; 0.1 μg/mL). The EP4 receptor agonist (ONO-AE1-329, 1 nM) was included as a positive control. Data shown are mean±SEM of three experimental runs performed in duplicate. #Statistical significance between vehicle and stimulation (p<0.05, Mann–Whitney test). *Statistical significance between stimulation and agonist treated cells (p<0.05, Kruskal–Wallis ANOVA followed by Dunn’s multiple comparison test).
followed by Dunn’s stimulation and agonist treated cells (p<0.05, Kruskal–Wallis) of primary human alveolar macrophages stimulated with LPS or vehicle.

Figure 7  

| Pathways | Effect of PKA activator plus increasing concentrations of PKA inhibitor (C) in primary human alveolar macrophages stimulated with LPS or vehicle. | PKA activator plus increasing concentrations of a PKA inhibitor (RP-8-CPT-cAMPS) in THP-1 cells.
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In conclusion, these findings present a convincing dataset that supports the hypothesis that the anti-inflammatory activity of PGE2 is mediated via activation of the EP4 receptor, and that this mechanism is capable of suppressing airway inflammation that is triggered via an array of stimuli. We have shown that EP4 receptor KO mice have enhanced inflammation after innate, allergic and COPD-like conditions, and cytokine production from primary human alveolar macrophages is attenuated by an EP4 agonist in a CAMP/PKA-dependent manner. We suggest that an EP4 receptor agonist with an appropriate pharmacokinetic profile would harness the beneficial properties of PGE2 and make an effective anti-inflammatory and bronchodilator for the treatment of respiratory diseases such as asthma and COPD.

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**Contributors**  

Conception and design: MAB and MGB. Data generation, analysis and interpretation: BD, SAM, PB. Drafting the manuscript for important intellectual content: MAB, BD, MGB. All authors reviewed the manuscript and approved the final draft.

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**Competing interests**  

None.

**Ethics approval**  

Ethical approval was obtained from the Royal Brompton Harefield ethics committee.

**Provenance and peer review**  

Not commissioned; externally peer reviewed.

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**REFERENCES**


