ORIGINAL ARTICLE

Sputum-to-serum hydrogen sulfide ratio in COPD

Junpei Saito,^{1,2,3} Alex J Mackay,⁴ Christos Rossios,^{1,2} David Gibeon,^{1,2} Patricia Macedo,^{1,2} Rudy Sinharay,^{1,2} Pankaj K Bhavsar,¹ Jadwiga A Wedzicha,³ Kian Fan Chung^{1,2}

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¹Section of Experimental Studies, National Heart and Lung Institute, Imperial College London, London, UK ²NIHR Respiratory Biomedical Research Unit at the Royal Brompton NHS Foundation Trust and Imperial College London, London, UK ³Department of Pulmonary Medicine, Fukushima Medical University, Fukushima, Japan ⁴Centre for Respiratory Medicine, University College London Medical School, Royal Free Campus, London, UK

Correspondence to

Professor K F Chung, Experimental Studies, National Heart and Lung Institute, Imperial College London, Dovehouse St, London SW3 6LY, UK; f.chung@imperial.ac.uk

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ABSTRACT

Objectives Hydrogen sulfide (H₂S) is a gas produced by respiratory cells including smooth muscle cells and may play a role as a cellular gasotransmitter. We evaluated whether H₂S levels in serum or sputum could represent a new biomarker of COPD in a cross-sectional study.

Methods H₂S levels in sputum and serum samples were measured using a sulfide-sensitive electrode in 64 patients with stable COPD (S-COPD), 29 COPD subjects during acute exacerbation (AE-COPD), 14 healthy smokers and 21 healthy non-smokers.

Results Sputum H₂S levels in AE-COPD subjects were higher than those in S-COPD, healthy smoking and nonsmoking subjects (p<0.001), but serum H₂S levels in AE-COPD were lower than those in S-COPD (p<0.001). Thus, the sputum-to-serum ratio of H₂S (H₂S ratio) in AE-COPD subjects were higher than those in stable COPD, healthy smoking and non-smoking subjects (p<0.001). In 14 COPD subjects whose H₂S ratios were measured during and after an exacerbation, the mean ratio was increased during exacerbation (p<0.05). H₂S ratio was positively correlated with St. George's Respiratory Questionnaire score, sputum neutrophils and IL-8 levels in sputum and serum (p<0.01) but inversely correlated with sputum macrophages (%), FEV₁%predicted and FEV₁/FVC (p<0.01). The cut-off level of H_2S ratio to indicate an exacerbation was >0.44 (sensitivity of 93.1% and specificity of 84.5%).

Conclusions The ratio of sputum-to-serum levels of H₂S may provide a useful marker of COPD indicative of obstructive neutrophilic inflammation and of potential ongoing exacerbation.

INTRODUCTION

COPD is one of the major causes of morbidity and mortality, likely to become the third global cause by 2020. COPD is characterised by persistent progressive airflow limitation and chronic pulmonary inflammation. Exacerbations of COPD characterised by worsening of respiratory symptoms are commonly observed in more severe disease and have been linked to lung function decline, cardiovascular events and death, and with increased pulmonary and extrapulmonary inflammation. Systemic inflammation may be a key link between COPD and comorbidities, possibly resulting from a spillover from the lung into the systemic circulation.

Hydrogen sulfide (H₂S), a gas with a typical malodorous smell, is produced by many cell types in the lungs including pulmonary arterial and airway

Key messages

What is the key question?

We evaluated whether H₂S levels in serum or sputum could represent a new biomarker of COPD.

What is the bottom line?

▶ We found that the ratio of sputum-to-serum levels of H₂S may provide a useful marker of COPD indicative of obstructive neutrophilic inflammation and of potential future risk of exacerbation.

Why read on?

 Concomitant measurement of H₂S in sputum and in serum represents a novel promising biomarker of COPD.

smooth muscle cells, primary fibroblasts and endothelial cells^{11–13} through the action of three enzymes, namely, cystathionine-γ-lyase (CSE), cystathionine-βsynthase (CBS) and 3-mercaptopyruvate sulfur transferase.¹⁴ H₂S is the third gasotransmitter along with nitric oxide (NO) and carbon monoxide (CO), with vasodilator and neurotransmitter properties. 15 H₂S may also be involved in anti-inflammatory processes and has been reported to reduce cigarette smoke-induced lung inflammation in mice. 16 It also has reducing properties and can scavenge various oxidising and nitrative species such as superoxide and peroxynitrite.¹⁷ In vivo, H₂S prevented endothelial disruption and lung vascular leakage induced by particulate air pollution through scavenging reactive oxygen species. 18

H₂S levels in the lung as well as in serum have been assayed in asthma and COPD. In COPD, higher levels of serum H2S have been reported compared with healthy non-smokers (HNS), with lower levels during an acute exacerbation. 19 In asthma, H₂S levels in both sputum and serum were strongly correlated with obstructive neutrophilic airway inflammation.²⁰ Therefore, H₂S could be considered as a potential biomarker of chronic airway diseases.²¹ The aims of our study were (i) to measure sputum and serum H₂S levels in patients with COPD during a stable period and during an exacerbation, and (ii) to examine their relationships to key markers of disease activity and neutrophilic inflammation. In addition, we evaluated whether sputum and serum levels of H2S could indicate an ongoing exacerbation.



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METHODS

Subjects

In total, 64 subjects with stable COPD (S-COPD) and 29 subiects with acute exacerbation of COPD (AE-COPD) were recruited from the clinics of the Royal Brompton and Royal Free Hospitals in London. Fourteen healthy smokers (HS) and 21 HNS were recruited from advertisement (table 1). The diagnosis of COPD was made according to the Global Initiative for Chronic Obstructive Lung Disease (GOLD) guidelines.²² Briefly, COPD subjects had a history of chronic respiratory symptoms (ie, dyspnoea, coughing, sputum or both), a smoking history of >10 pack-years and a postbronchodilator FEV₁/FVC ratio <70%. COPD subjects were categorised as severe (stages III and IV) or non-severe (stages I and II). Subjects with concomitant respiratory diseases other than COPD were excluded. HS (>10 pack-years) and HNS (≤5 pack-years) had normal lung function, no bronchial hyperresponsiveness and no history of pulmonary disease. All participants gave informed consent to a protocol approved by the Ethics Committee.

COPD exacerbation

An acute exacerbation was defined as new or increased respiratory symptoms for two or more consecutive days, with at least one major symptom (dyspnoea and sputum production) with either another major or minor symptom (wheeze, cold, sore throat and cough). All exacerbation visits occurred within 7 days after its onset. A stable state was defined as having no symptoms related to exacerbation for the preceding 6 weeks.²³

Study design

This was a cross-sectional observational study. Health status was assessed with the Medical Research Council (MRC) dyspnoea scale²⁴ and the St. George's Respiratory Questionnaire (SGRQ).²⁵ Lung function tests, 6 minute walk test (6MWT) and sputum and peripheral blood samplings were performed. In 14 COPD subjects, H₂S levels in sputum and serum were obtained during an exacerbation and during the stable state.

Sputum induction and processing

Sputum induction was performed with inhaled nebulised 4.5% NaCl solution. Sputum plugs were harvested and processed with 0.1% dithiothreitol (DTT) and sulfide antioxidant buffer added to sputum supernatant and stored at -80 °C. Cytospins were prepared for differential cell counts.

	Healthy non-smoker (n=21)	Healthy smoker (n=14)	Stable COPD (n=64)	COPD with exacerbation (n=29)
Age (years)	49.2 (9.24)	54.4 (7.31)	69.1 (8.56) [*] [†]	74.7 (9.06) [*] †
Sex (male/female)	13/8	6/8	33/31	18/11
Height (cm)	168.3 (7.80)	165.8 (7.96)	166.7 (8.32)	167.4 (9.06)
Weight (kg)	67.7 (17.6)	66.6 (14.0)	72.7 (14.2)	70.8 (13.0)
BMI	23.8 (5.10)	24.2 (4.46)	26.1 (4.41)	25.2 (4.02)
Smoking (pack-years), median (IQR)	N/A	23.0 (19.3–31.8)	39.8 (27.5–55.0)	37.4 (19.7–51.3)
GOLD stage (I/II/III/IV))	N/A	N/A	5/36/19/4	2/14/12/1
MRC score (1–5)	N/A	N/A	2.57 (0.98)	3.46 (1.04) [¶]
Baseline SPO2 (%)	97.4 (1.51)	97.2 (1.14)	94.8 (2.26)* [†]	94.8 (2.48)* †
SGRQ score				
Symptom score	3.91 (6.22)	25.8 (20.1)*	58.6 (22.3)* [†]	N/D
Activity score	1.97 (3.70)	23.4 (22.3)*	61.1 (24.9)* [†]	N/D
Impact score	0.26 (1.00)	4.91 (7.49)	28.9 (16.9)* [†]	N/D
Total score	1.30 (1.26)	13.6 (12.9)	43.3 (19.1)* [†]	N/D
6 minute walk distance(m)	594 (36.5)	497 (64.0)	414 (82.1)* [†]	N/D
FEV ₁ (%pred)	99.8 (12.1)	89.3 (11.4)	55.6 (16.6)* [†]	55.1 (17.4)* [†]
FVC (%pred)	103.0 (11.2)	98.3 (13.2)	86.6 (20.2)* [†]	89.3 (21.9)* [†]
FEV ₁ /FVC (%)	82.0 (5.42)	76.4 (5.16)	52.4 (12.7)* [†]	49.7 (9.30)* [†]
TL _{CO} (%)	88.0 (16.8)	75.5 (15.9)	62.4 (17.6)*	N/D
K _{CO} (%)	91.5 (12.2)	82.3 (14.8)	73.5 (20.1)*	N/D
WBC, median (IQR)	5.60 (5.00-7.70)	6.80 (5.50-8.65)	7.10 (6.00-9.24)	7.95 (6.87–9.63)*
Neutrophil, median (IQR)	3.40 (2.80-4.10)	4.10 (2.90-5.10)	7.10 (6.00-9.24)*	7.95 (6.87–9.63)* ^{† ¶}
CRP, median (IQR)	0.50 (0.50-1.00)	2.00 (0.50-5.50)	3.00 (2.00-10.0)	13.5 (6.25–28.5)* [†] ¶
Sputum macrophages (%), median (IQR)	85.8 (64.7–87.8)	45.8 (19.9-58.9)*	30.6 (17.5-49.0)*	18.7 (8.50–24.5)* [†]
Sputum neutrophils (%), median (IQR)	12.3 (6.67–33.5)	52.5 (37.9–79.2)*	63.0 (43.4–79.0)*	80.5 (72.8–91.3)* [†]
Sputum eosinophils (%), median (IQR)	0.70 (0.22-1.00)	0.68 (0.21-1.00)	1.44 (0.50-2.52)	0 (0–1.25)
Maintenance therapy				
SABA or SAMA, n (%)	N/A	N/A	13 (20.3)	7 (24.1)
LABA or LAMA, n (%)	N/A	N/A	1 (1.6)	0 (0)
ICS+(LABA or LAMA), n (%)	N/A	N/A	16 (25)	8 (27.6)
ICS+LABA+LAMA, n (%)	N/A	N/A	34 (53.1)	14 (48.3)

Data are presented as mean (SD) unless otherwise indicated, *p<0.05 versus healthy non-smoker, $^{\dagger}p$ <0.05 versus healthy smoker, $^{\dagger}p$ <0.05 versus stable COPD. BMI, body mass index; CRP, C-reactive protein; GOLD, Global Initiative for Chronic Obstructive Lung Disease; ICS, inhaled corticosteroid; K_{CO} , carbon monoxide transfer coefficient; LABA, long-acting β -agonist; LAMA, long-acting muscarinic antagonist; MRC, Medical Research Council; N/A, not applicable; N/D, not done; SABA, short-acting β -agonist; SAMA, short-acting muscarinic antagonist; SGRQ, St. George's Respiratory Questionnaire; TL_{CO}, transfer factor of the lung for carbon monoxide; WBC, white blood cell.

Measurement of H₂S

 H_2S concentrations in serum and sputum were measured using a sulfide-sensitive electrode (Model 9616; Orion Research; Beverly, Massachusetts, USA). Briefly, standard solutions were made using 0.1% DTT plus phosphate-buffered saline or distilled water for sputum and serum H_2S measurement respectively in order to minimise any effect of DTT. The electrode was placed into the standard solutions for measurement of H_2S levels and the calibration standard curve constructed. Validation of H_2S measurements including accuracy, reproducibility and storage effects of samples has been reported. 20

Measurement of IL-6 and IL-8

IL-6 and IL-8 in serum and sputum supernatants were measured using a commercially available sandwich ELISA kit (R&D Systems, Minneapolis, Minnesota, USA).

Statistical analysis

The main outcome predictor was H_2S levels in sputum and serum, or the ratio of sputum to serum H_2S levels. At least 14 subjects were needed per group for a 70% power to detect a 20% difference in H_2S levels between each group with a two-sided α of 0.05.

Data are shown as mean and SD for normally distributed variables, and median and IQR for non-normally distributed variables. Comparisons of continuous variables between groups were made using Kruskall-Wallis and Mann-Whitney U test with Bonferroni correction. χ^2 or Fisher's exact test was used for the comparison of categorical data. Differences of paired samples in the same subjects were analysed by Wilcoxon matched-pairs signed rank test. Linear regression modelling was used to evaluate the simple and joint associations of H₂S levels in sputum and serum with parameters that were relevant to COPD, adjusting for age, sex, height, weight and smoking status. Then, multiple linear regression analysis, using a forward stepwise selection, was conducted to determine the independent association of H₂S levels with parameters with p value <0.05 obtained from regression modelling. Finally, receiver operating characteristic (ROC) curve was constructed to determine the predictive value of the sputum-to-serum H₂S ratio for an exacerbation. A two-tailed p value < 0.05 was considered significant.

RESULTS

Characteristics of participants

Forty-one (64.0%) in S-COPD group and 16 (55.2%) in AE-COPD group had mild-to-moderate COPD. SpO_2 at rest, FEV₁, FVC, FEV₁/FVC ratio and sputum macrophages in COPD subjects were lower than in healthy subjects (p<0.05). The 6MWT distance in S-COPD subjects was also shorter (p<0.05). Total SGRQ score and neutrophils in blood and sputum in S-COPD and AE-COPD subjects were higher than those of HNS (p<0.05).

H₂S levels in sputum and serum

H₂S levels in induced sputum from S-COPD (31.9±15.0 μM) were higher than those from HNS (12.1±6.64 μM; p<0.001). In addition, sputum H₂S levels in AE-COPD subjects (50.4±24.4 μM) were much higher than those from S-COPD, HS and HNS subjects (31.9±15.0, 12.1±6.64, 19.7±7.98 μM, respectively; p<0.001) (figure 1A). The concomitant serum H₂S levels in S-COPD (148.9±77.6 μM) were also higher than those from HNS and HS (91.0±62.2, 90.6±52.7 μM, respectively; p<0.05). However, unlike the

sputum H_2S levels, the concomitant serum H_2S levels in AE-COPD (48.9±25.5 μ M) were much lower compared with those in S-COPD (figure 1B). There was no difference in H_2S levels between non-severe and severe COPD subjects during both stable and exacerbated states (see online supplementary figure E1).

Relationships between H₂S levels and COPD parameters

Table 2 shows the correlations between sputum H₂S, serum H₂S levels and clinically relevant parameters in S-COPD. Both sputum and serum H₂S levels positively correlated with MRC scores, total SGRQ scores, sputum neutrophils and serum IL-8 levels (p<0.01). Sputum H_2S levels, but not serum H_2S levels, showed positive correlations with sputum IL-8 and sputum IL-6 levels (p<0.01). In addition, there were negative correlations between sputum as well as serum H₂S levels and SpO₂ at rest, 6MWT distance, FEV₁%predicted, FEV₁/FVC, and sputum macrophages (p<0.01). Sputum H_2S levels, but not serum H_2S , were negatively correlated to transfer factor of the lung for carbon monoxide (TL_{CO}) and carbon monoxide diffusing capacity adjusted for alveolar volume (K_{CO}) (p<0.01). When adjusted for the potential confounding factors (age, sex, height, weight and smoking status), sputum H₂S levels were still correlated with 6MWT distance, FEV₁%predicted, FEV₁/FVC ratio, TL_{CO}, sputum macrophages (%), sputum neutrophils (%) and sputum IL-8 levels. Similarly, serum H₂S levels were significantly correlated with FEV₁%predicted and FEV₁/FVC ratio. For AE-COPD subjects, there was no significant correlation between sputum H2S, serum H2S levels and other COPD-related parameters (data not shown).

Using multiple linear regression analysis to determine the independent association with sputum and serum H_2S (table 3), sputum H_2S was associated with increased sputum neutrophil (%) and sputum IL-8 levels, and decreased FEV_1 (% predicted) ($p \le 0.05$), and serum H_2S was associated with decreased FEV_1 (% predicted) (p = 0.03). Sputum and serum H_2S were not influenced by age, sex, height, weight and smoking status.

Sputum-to-serum H₂S ratio

We examined the ratio of sputum-to-serum levels, the 'H₂S ratio'. H₂S ratio in AE-COPD (mean 1.42; 95% CI 0.89 to

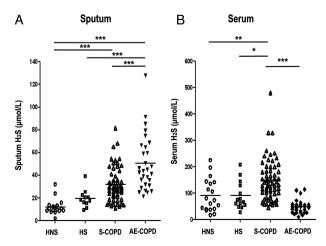


Figure 1 H₂S levels in sputum (A) and serum (B) from stable COPD subjects (S-COPD:Δ), COPD subjects with acute exacerbation (AE-COPD: ▼), healthy smoking (HS:□), and non-smoking subjects (HNS:□). Horizontal bars indicate mean. ***p≤0.001.

Table 2 Correlation analysis between H₂S in sputum and serum and other parameters related to COPD

	Sputum H ₂ S		Serum H₂S	
Variables	Crude β value (95% CI)	Adjusted† β value (95% CI)	Crude β value (95% CI)	Adjusted† β value (95% CI)
MRC score	6.006 (3.258 to 8.754)***	-	14.83 (0.781 to 28.87)*	_
SGRQ score	0.200 (0.066 to 0.333)**	_	0.685 (0.005 to 1.366)*	_
Baseline SpO ₂ (%)	-1.860 (-3.386 to -0.335)*	_	-9.415 (-15.73 to -3.098)**	-
6 MWD (m)	-0.084 (-0.122 to -0.045)***	-0.065 (-0.117 to -0.013)*	-0.218 (-0.405 to -0.031)*	_
FEV1%predicted (%)	-0.337 (-0.452 to -0.222)***	-0.272 (-0.430 to -0.114)**	-1.161 (-1.749 to -0.573)***	-1.167 (-1.989 to -0.346)**
FEV1/FVC (%)	-0.441 (-0.612 to -0.270)***	-0.341 (-0.575 to -0.106)**	-1.426 (-2.291 to -0.560)**	-1.271 (-2.522 to -0.020)*
TL _{CO} (%)	-0.333 (-0.524 to -0.143)***	-0.261 (-0.491 to -0.031)**	-0.621 (-1.661 to 0.420)	_
K _{CO} (%)	-0.275 (-0.460 to -0.099)**	_	-0.594 (-1.561 to 0.412)	-
Sputum macrophages (%)	-0.364 (-0.465 to -0.263)***	-0.331 (-0.446 to -0.216)***	-0.864 (-1.508 to -0.221)**	_
Sputum neutrophils (%)	0.353 (0.251 to 0.454)***	0.309 (0.193 to 0.424)***	0.868 (0.223 to 1.512)**	_
Serum IL8 (pg/mL)	0.409 (0.045 to 0.773)*	_	1.768 (0.184 to 3.352)*	-
Serum IL6 (pg/mL)	0.912 (-0.923 to 2.748)	_	10.62 (2.522 to 18.72)*	_
Sputum IL8 (pg/mL)	0.017 (0.009 to 0.024)***	0.013 (0.005 to 0.022)**	0.012 (-0.036 to 0.059)	-
Sputum IL6 (pg/mL)	0.038 (0.010 to 0.066)**	_	-0.075 (-0.284 to 0.134)	-

Data are presented as β value and 95% CIs. *p<0.05, **p<0.01, ***p<0.001.

1.96) was higher than that in S-COPD subjects (mean 0.27; 95% CI 0.22 to 0.32), HNS (mean 0.20; 95% CI 0.11 to 0.28) and HS (mean 0.24; 95% CI 0.17 to 0.31) (p<0.001) subjects (figure 2A). For further confirmation of this difference, paired samples of sputum and serum during exacerbation as well as stable state from 14 COPD subjects were obtained. $\rm H_2S$ ratio during exacerbation (mean 1.54; 95% CI 0.58 to 2.49) was increased compared with that during stable state in all 14 participants (mean 0.28, 95% CI 0.19 to 0.36) (figure 2B) (p<0.01).

Sputum-to-serum IL-8 and IL-6 ratios

We determined whether the sputum-to-serum ratios of the levels of IL-6 and IL-8 could also be altered during exacerbations. IL-8 ratio in AE-COPD (mean 54.8; 95% CI 46.3 to 63.4) was lower than that in HS (p<0.01), but was not different from S-COPD and HNS (figure 3A). There was no significant difference in IL-6 ratio between the groups (figure 3B). Both serum and sputum IL-6 levels were increased in AE-COPD compared with S-COPD, while only serum IL-8 was increased (see online supplementary figure E2).

Table 3 Independent parameters associated with H₂S in sputum and serum using multiple linear regression analysis*

			0.563
499 –0.	900 to -0.099	0.016	
271 0.0	66 to 0.476	0.011	
016 0.0	004 to 0.028	0.012	
			0.263
621 –1.1	79 to -0.063	0.029	
	.271 0.0 .016 0.0	271 0.066 to 0.476 016 0.004 to 0.028	271 0.066 to 0.476 0.011 016 0.004 to 0.028 0.012

H₂S ratio, bacterial sputum cultures and onset of exacerbations

 H_2S ratio in AE-COPD with negative bacterial sputum cultures (mean 2.33; 95% CI 0.65 to 4.01) was higher than that in AE-COPD with positive sputum cultures (mean 1.03; 95% CI 0.52 to 1.54) (p<0.05), S-COPD with negative (mean 0.28; 95% CI 0.21 to 0.36) and S-COPD with positive bacteria (mean 0.25; 95% CI 0.14 to 0.36) (p<0.001). There was no difference in H_2S ratio between S-COPD with and without bacteria (figure 4A).

There was a negative correlation between H_2S ratio and the time interval from exacerbation onset to a hospital visit, that is, the greater the H_2S ratio, the shorter the time of presentation to hospital (r=-0.45; p=0.02) (figure 4B).

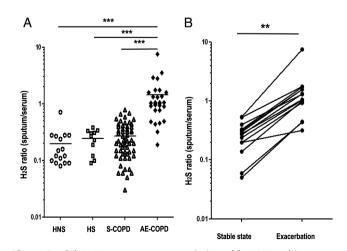


Figure 2 (A) H₂S sputum-to-serum ratio in stable COPD subjects (S-COPD: Δ), COPD subjects with acute exacerbation (AE-COPD: \blacktriangledown), healthy smoking (HS: \bigcirc) and non-smoking subjects (HNS: \bigcirc). Horizontal bars indicate mean. ***p≤0.001. (B) Difference in H₂S sputum-to-serum ratio between stable and exacerbation state. At acute exacerbation, H₂S ratio increased in comparison with that in stable state. **p≤0.01.

[†]Adjusted for age, gender, height, weight, and smoking status.

⁶MWD, 6 minute walk distance; K_{CO}, carbon monoxide diffusing capacity adjusted for alveolar volume; K_{CO}, TL_{CO}/Alveolar volume (Va); MRC, Medical Research Council; SGRQ,

St. George's Respiratory Questionnaire; TL_{CO}, Transfer factor of the lung for carbon monoxide.

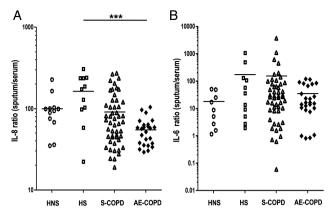


Figure 3 IL-8 sputum-to-serum ratio (A) and IL-6 sputum-to-serum ratio (B) from stable COPD subjects (S-COPD:△), COPD subjects with acute exacerbation (AE-COPD:▼), healthy smoking (HS:□) and non-smoking subjects (HNS:○). Horizontal bars indicate mean.
***p≤0.001.

H₂S ratio as indicative of an exacerbation

ROC curves were constructed to determine the cut-off level of H_2S ratio for indicating an acute exacerbation from stable COPD (figure 5). The optimal cut-off level of H_2S ratio for indicating an exacerbation was ≥ 0.44 (area under the curve; 0.946, sensitivity of 93.1% and specificity of 84.5%, p<0.001).

DISCUSSION

In patients with COPD, sputum H₂S levels were elevated in S-COPD and AE-COPD compared with healthy smoking and non-smoking subjects; however, serum levels were higher in stable COPD but not during an acute exacerbation when compared with serum levels in healthy subjects. Chronic cigarette smoking itself did not influence serum or sputum levels. Sputum H₂S levels increased and serum H₂S levels dropped during acute exacerbations of COPD, as confirmed in patients where concomitant sputum and serum levels were measured at baseline and during an exacerbation. Both sputum and serum H₂S levels correlated inversely with the degree of airflow obstruction,

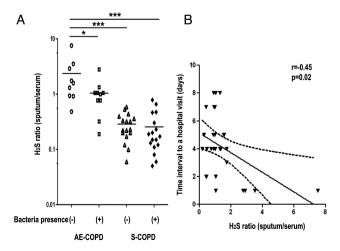


Figure 4 (A) H_2S sputum-to-serum ratio in AE-COPD and S-COPD with and without bacterial presence; AE-COPD without bacteria (\bigcirc), AE-COPD with bacteria (\bigcirc), S-COPD without bacteria (\triangle) and S-COPD with bacteria (\spadesuit). Horizontal bars indicate mean. (B) Relationship between H_2S ratio and the time interval from exacerbation onset to a hospital visit.

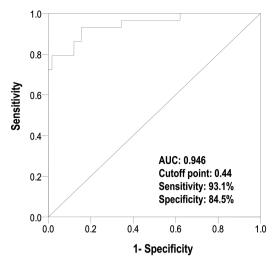


Figure 5 Receiver operator characteristics curve for H₂S sputum-to-serum ratio to predict the future risks of acute exacerbation.

suggesting that H₂S levels are closely related to physiological characteristics of COPD. There were positive relationships between sputum neutrophils (%), serum IL-8 and H₂S levels in sputum and serum, indicating that H₂S levels may reflect neutrophilic airway inflammation. Furthermore, H₂S levels in sputum and serum were associated with SpO₂ at rest, 6MWT distance, MRC score and SGRQ scores, indicating that H₂S may also be a marker of the degree of physical and quality-of-life impairment.

This is the first study to examine the levels of H₂S in sputum and serum during the stable and acute exacerbation phases of COPD. Previous studies have reported serum H2S levels in COPD and during chest infections 19 27 and in agreement with our results, serum H₂S levels in patients with S-COPD were found to be higher than those in HNS, with lower levels in AE-COPD. 19 Interestingly, the changes in sputum and serum H₂S levels between stable and exacerbation were entirely opposite with an increase in sputum H2S levels and concomitant decrease in serum H₂S levels during acute exacerbation. Thus, the sputum-to-serum H₂S ratio increased sixfold during an exacerbation and a ratio >0.44 was indicative of an exacerbation. By comparison, the sputum-to-serum IL-6 and IL-8 ratios did not differentiate stable COPD and COPD during an exacerbation from non-smoking and smoking healthy subjects. This lack of differentiation is explained by the fact that serum IL-6 and IL-8 levels increased during an exacerbation in contrast to the reduction in serum H₂S levels; in addition, the increase in sputum cytokines during exacerbation was either negligible or small.

The role of H₂S in the lungs is currently unclear. H₂S possesses anti-inflammatory and antioxidant effects in the lungs. In vitro, H₂S can suppress airway smooth muscle proliferation and IL-8 release, ¹² and induce vascular smooth muscle relaxation. ²⁸ In in vivo studies, aggravated airway hyperresponsiveness and increased airway inflammation occurred not only in mouse models of asthma with deficiency of H₂S-producing enzymes ²⁹ but also in rat models of cigarette-induced inflammation when CSE was blocked. ¹⁶ Therefore, H₂S may have a protective role in COPD. However, other studies have revealed that H₂S has a proinflammatory role in regulating the severity of LPS-induced sepsis, ³⁰ acute pancreatitis ³¹ and burn injury. ³² Inhibition of H₂S in animal models was protective of organ injury in endotoxaemia ³⁰ and so systemic production of H₂S might be downregulated during an exacerbation.

Endogenous H₂S synthesis occurs through the action of several enzymes including CSE and CBS. 14 In airway smooth muscle cells. CBS is the more important enzyme in generating H₂S.¹² We have preliminary data indicating that in airway smooth muscle cells from patients with COPD, there is an increase in expression of CBS. An increase in CBS may underlie the increase in H₂S found in sputum and serum of COPD subjects. The increase in serum H₂S we found in COPD subjects is in agreement with a previous study, 19 as is our finding of a reduction in serum H₂S during exacerbation. The mechanisms by which H2S levels are decreased in serum and increased in sputum are unclear. In addition to increased expression of H₂S-generating enzymes such as CBS, there could be an increase in inflammatory cells in the lungs during an exacerbation together with an increased bacterial load that could add to the increased production of H2S. However, in serum, the reduction in H₂S levels may be a reflection of increased sequestration of H₂S from the circulation into the lungs.

In our study, the higher the levels of sputum and serum H₂S, the greater were the degree of airflow obstruction and of neutrophilic inflammation. These results are similar to those found in patients with asthma where the levels of sputum and serum H₂S correlated with the degree of airflow limitation and the level of sputum neutrophilia.²⁰ In addition, in COPD, the 6MWT distance, which could be influenced by physical activity, and quality-of-life scores among many other determinants³³ ³⁴ also correlated with sputum and serum levels of H₂S indicating the potential value of measuring this diffusible marker.

Importantly, these levels may also indicate the risk of an exacerbation. A most significant observation is the decrease in the systemic levels of H₂S and an increase in the lung levels of H₂S during exacerbations that make this ratio a unique biomarker of a warning system for exacerbations.

We have used a sulfide-ion selective electrode to assay H₂S that usually provides levels in the µM range, while other techniques have detected lower levels.²⁷ ³⁵ We have added a sulfide-antioxidant buffer to our samples prior to storage and subsequent assay in order to prevent the oxidation of sulfur compounds and avoid the volatilisation of H₂S.²⁰ Thus, these levels represent the total sulfide pool (rather than solely as H₂S) derived by hydroxyl replacement on cysteine residues in the blood and sputum protein pool. We performed a spiking experiment with different concentrations of H2S added to serum samples and found a tight correlation between the measured and predicted H₂S levels.²⁰ In terms of short-term reproducibility, serum H₂S levels from nine healthy volunteers taken twice over a week period had a mean coefficient of variation of 6.34%.²⁰ However, the short-term and long-term reproducibility of sputum H₂S measurements in healthy or COPD subjects is not known.

Other than the inflammatory milieu, other factors may influence the levels of H₂S. First, the effect of COPD medications, particularly anticholinergic, β-adrenergic agonists and inhaled or oral corticosteroids on the production of H₂S, is not known. Corticosteroids may inhibit H₂S production in macrophages partly through the inhibition of the H₂S-producing enzyme, CSE.³⁶ ³⁷ However, there was no difference in H₂S levels between subjects taking and not taking inhaled corticosteroids (data not shown). Fluticasone propionate inhalation had no effect on the levels of sputum H₂S in patients with COPD.³⁸ Second, bacteria in the upper and lower airways may contribute to H₂S production, particularly that the lower airways contain bacteria with an altered bacteriome in COPD.³⁹ In addition, H₂S produced by bacteria may sustain bacterial growth and

suppress their sensitivity to antibiotics.⁴⁰ In our exacerbating patients where conventional sputum cultures were positive for bacteria, the sputum-to-serum ratios were significantly lower than those with negative bacterial cultures, thus indicating the contribution of lower airway bacterial infections to H₂S levels.

In summary, concomitant measurements of H_2S in sputum and in serum may provide a novel biomarker of COPD. H_2S levels may be valuable as a marker of neutrophilic inflammation, chronic airflow obstruction, physical activity and of an acute exacerbation. The clinical utility and validity of measuring the sputum-to-serum ratios of H_2S will need to be tested in larger COPD cohorts.

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REFERENCES

- 1 Murray CJ, Lopez AD. Alternative projections of mortality and disability by cause 1990–2020: Global Burden of Disease Study. *Lancet* 1997;349:1498–504.
- 2 Chung KF, Adcock IM. Multifaceted mechanisms in COPD: inflammation, immunity, and tissue repair and destruction. Eur Respir J 2008;31:1334–56.
- 3 Donaldson GC, Hurst JR, Smith CJ, et al. Increased risk of myocardial infarction and stroke following exacerbation of COPD. Chest 2010;137:1091–7.
- 4 Donaldson GC, Seemungal TA, Bhowmik A, et al. Relationship between exacerbation frequency and lung function decline in chronic obstructive pulmonary disease. *Thorax* 2002;57:847–52.
- Wedzicha JA, Seemungal TA, MacCallum PK, et al. Acute exacerbations of chronic obstructive pulmonary disease are accompanied by elevations of plasma fibrinogen and serum IL-6 levels. *Thromb Haemost* 2000;84:210–15.
- 6 Hurst JR, Donaldson GC, Perera WR, et al. Use of plasma biomarkers at exacerbation of chronic obstructive pulmonary disease. Am J Respir Crit Care Med 2006;174:867–74.
- Hurst JR, Perera WR, Wilkinson TM, et al. Systemic and upper and lower airway inflammation at exacerbation of chronic obstructive pulmonary disease. Am J Respir Crit Care Med 2006;173:71–8.
- 8 Hurst JR, Perera WR, Wilkinson TM, et al. Exacerbation of chronic obstructive pulmonary disease: pan-airway and systemic inflammatory indices. Proc Am Thorac Soc 2006;3:481–2.
- 9 Divo M, Cote C, de Torres JP, et al. Comorbidities and risk of mortality in patients with chronic obstructive pulmonary disease. Am J Respir Crit Care Med 2012;186:155–61.
- Barnes PJ, Celli BR. Systemic manifestations and comorbidities of COPD. Fur Respir J 2009:33:1165–85.
- Baskar R, Li L, Moore PK. Hydrogen sulfide-induces DNA damage and changes in apoptotic gene expression in human lung fibroblast cells. FASEB J 2007;21:247–55.
- Perry MM, Hui CK, Whiteman M, et al. Hydrogen sulfide inhibits proliferation and release of IL-8 from human airway smooth muscle cells. Am J Respir Cell Mol Biol 2011;45:746–52.
- 13 Olson KR, Whitfield NL, Bearden SE, et al. Hypoxic pulmonary vasodilation: a paradigm shift with a hydrogen sulfide mechanism. Am J Physiol Regul Integr Comp Physiol 2010;298:R51–60.
- 14 Wang R. Physiological implications of hydrogen sulfide: a whiff exploration that blossomed. *Physiol Rev* 2012;92:791–896.
- 15 Wang R. Two's company, three's a crowd: can H2S be the third endogenous gaseous transmitter? FASEB J 2002;16:1792–8.
- 16 Chen YH, Wang PP, Wang XM, et al. Involvement of endogenous hydrogen sulfide in cigarette smoke-induced changes in airway responsiveness and inflammation of rat lung. Cytokine 2011;53:334–41.
- 17 Whiteman M, Armstrong JS, Chu SH, et al. The novel neuromodulator hydrogen sulfide: an endogenous peroxynitrite 'scavenger'? J Neurochem 2004;90:765–8.
- 18 Wang T, Wang L, Zaidi SR, et al. Hydrogen sulfide attenuates particulate matter-induced human lung endothelial barrier disruption via combined reactive oxygen species scavenging and Akt activation. Am J Respir Cell Mol Biol 2012;47:491–6.

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- 19 Chen YH, Yao WZ, Geng B, et al. Endogenous hydrogen sulfide in patients with COPD. Chest 2005;128:3205–11.
- 20 Saito J, Zhang Q, Hui C, et al. Sputum hydrogen sulfide as a novel biomarker of obstructive neutrophilic asthma. J Allergy Clin Immunol 2013;131:232–4 e231–233.
- 21 Chung KF. Hydrogen sulfide as a potential biomarker of asthma. *Expert Rev Respir Med* 2014;8:5–13.
- Vestbo J, Hurd SS, Agusti AG, et al. Global strategy for the diagnosis, management, and prevention of chronic obstructive pulmonary disease: GOLD executive summary. Am J Respir Crit Care Med 2013;187:347–65.
- 23 Mackay AJ, Donaldson GC, Patel AR, et al. Usefulness of the chronic obstructive pulmonary disease assessment test to evaluate severity of COPD exacerbations. Am J Respir Crit Care Med 2012;185:1218–24.
- 24 Bestall JC, Paul EA, Garrod R, et al. Usefulness of the Medical Research Council (MRC) dyspnoea scale as a measure of disability in patients with chronic obstructive pulmonary disease. Thorax 1999;54:581–6.
- 25 Jones PW, Quirk FH, Baveystock CM, et al. A self-complete measure of health status for chronic airflow limitation. The St. George's Respiratory Questionnaire. Am Rev Respir Dis 1992;145:1321–7.
- 26 Pizzichini E, Pizzichini MM, Efthimiadis A, et al. Indices of airway inflammation in induced sputum: reproducibility and validity of cell and fluid-phase measurements. Am J Respir Crit Care Med 1996:54:308–17.
- 27 Chen YH, Yao WZ, Gao JZ, et al. Serum hydrogen sulfide as a novel marker predicting bacterial involvement in patients with community-acquired lower respiratory tract infections. Respirology 2009;14:746–52.
- 28 Wang R. Signaling pathways for the vascular effects of hydrogen sulfide. Curr Opin Nephrol Hypertens 2011;20:107–12.
- 29 Zhang G, Wang P, Yang G, et al. The inhibitory role of hydrogen sulfide in airway hyperresponsiveness and inflammation in a mouse model of asthma. Am J Pathol 2013;182:1188–95.

- O Collin M, Anuar FB, Murch O, et al. Inhibition of endogenous hydrogen sulfide formation reduces the organ injury caused by endotoxemia. Br J Pharmacol 2005:146:498–505.
- 31 Bhatia M, Wong FL, Fu D, et al. Role of hydrogen sulfide in acute pancreatitis and associated lung injury. FASEB J 2005;19:623–5.
- 32 Zhang J, Sio SW, Moochhala S, et al. Role of hydrogen sulfide in severe burn injury-induced inflammation in mice. Mol Med 2010;16:417–24.
- 33 Gimeno-Santos E, Frei A, Steurer-Stey C, et al. Determinants and outcomes of physical activity in patients with COPD: a systematic review. *Thorax* 2014;69:731–9.
- 34 Pitta F, Troosters T, Spruit MA, et al. Characteristics of physical activities in daily life in chronic obstructive pulmonary disease. Am J Respir Crit Care Med 2005;171:972–7.
- 35 Whitfield NL, Kreimier EL, Verdial FC, et al. Reappraisal of H2S/sulfide concentration in vertebrate blood and its potential significance in ischemic preconditioning and vascular signaling. Am J Physiol Regul Integr Comp Physiol 2008;294:R1930–7.
- 36 Zhu XY, Liu SJ, Liu YJ, et al. Glucocorticoids suppress cystathionine gamma-lyase expression and H2S production in lipopolysaccharide-treated macrophages. Cell Mol Life Sci 2010;67:1119–32.
- 37 Li L, Whiteman M, Moore PK. Dexamethasone inhibits lipopolysaccharide-induced hydrogen sulphide biosynthesis in intact cells and in an animal model of endotoxic shock. J Cell Mol Med 2009;13:2684–92.
- 38 Kirkham PA, Whiteman M, Winyard PG, et al. Impact of theophylline/corticosteroid combination therapy on sputum hydrogen sulfide levels in patients with COPD. Eur Respir J 2014;43:1504–6.
- 39 Hilty M, Burke C, Pedro H, et al. Disordered microbial communities in asthmatic airways. PloS ONE 2010;5:e8578.
- 40 Shatalin K, Shatalina E, Mironov A, et al. H2S: a universal defense against antibiotics in bacteria. Science 2011;334:986–90.