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# Progress in cystic fibrosis and the CF Therapeutics Development Network

Steven M Rowe,<sup>1</sup> Drucy S Borowitz,<sup>2</sup> Jane L Burns,<sup>3</sup> John P Clancy,<sup>4</sup>  
Scott H Donaldson,<sup>5</sup> George Retsch-Bogart,<sup>5</sup> Scott D Sagel,<sup>6</sup>  
Bonnie W Ramsey<sup>3</sup>

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<sup>1</sup>Department of Medicine, University of Alabama at Birmingham, Birmingham, Alabama, USA

<sup>2</sup>Department of Pediatrics, State University of New York at Buffalo, Buffalo, New York, USA

<sup>3</sup>Seattle Children's Hospital and the University of Washington, Seattle, Washington, USA

<sup>4</sup>Department of Pediatrics, Cincinnati Children's Hospital Medical Center, and the University of Cincinnati, Cincinnati, Ohio, USA

<sup>5</sup>University of North Carolina at Chapel Hill, Chapel Hill, North Carolina, USA

<sup>6</sup>Department of Pediatrics, Children's Hospital Colorado and University of Colorado, Aurora, Colorado, USA

## Correspondence to

Dr Steven M Rowe, University of Alabama at Birmingham, 1819 University Boulevard (MCLM 768), Birmingham, AL 35294, USA; [smrowe@uab.edu](mailto:smrowe@uab.edu)

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## ABSTRACT

Cystic fibrosis (CF), the most common life-shortening genetic disorder in Caucasians, affects approximately 70 000 individuals worldwide. In 1998, the Cystic Fibrosis Foundation (CFF) launched the CF Therapeutics Development Network (CF-TDN) as a central element of its Therapeutics Development Programme. Designed to accelerate the clinical evaluation of new therapies needed to fulfil the CFF mission to control and cure CF, the CF-TDN has conducted 75 clinical trials since its inception, and has contributed to studies as varied as initial safety and proof of concept trials to pivotal programmes required for regulatory approval. This review highlights recent and significant research efforts of the CF-TDN, including a summary of contributions to studies involving CF transmembrane conductance regulator (CFTR) modulators, airway surface liquid hydrators and mucus modifiers, anti-infectives, anti-inflammatories, and nutritional therapies. Efforts to advance CF biomarkers, necessary to accelerate the therapeutic goals of the network, are also summarised.

## STRUCTURE OF THE NETWORK, CFF PARTNERSHIP, INDUSTRY PARTNERSHIPS, OPERATIONAL ISSUES THAT FOSTER SUCCESS

Cystic fibrosis (CF), the most common life-shortening genetic disorder in Caucasians, affects approximately 70 000 individuals worldwide and meets the US Food and Drug Administration (FDA) definition of an orphan disease.<sup>1</sup> Historically, the limited number of patients at each clinical site led to underpowered research studies<sup>2</sup> and was a critical barrier to development and regulatory approval of new therapies. Hence, the Cystic Fibrosis Foundation (CFF) launched the CF Therapeutics Development Network (CF-TDN) in 1998 to accelerate the clinical evaluation of new CF therapies.<sup>3 4</sup> From its inception, the CF-TDN was organised to provide necessary infrastructure to promote efficient study conduct. In addition, patient safety has always been the highest priority.<sup>3</sup> From the beginning, the programme was composed of Therapeutic Development Centers (TDCs) across the USA, a clinical and data coordinating centre located at Seattle Children's Research Institute, and a number of resource centres with expertise in interpretation or performance of specialised outcomes measures (microbiology, imaging, infant lung function testing, inflammatory mediators, sweat testing, CF transmembrane conductance regulator (CFTR) detection and cytology interpretation) (see online supplementary figure S1). To maintain scientific

integrity and safety of CF-TDN studies, there is a Steering Committee, protocol review of all multi-center trials, and review of presentations and publications from CF-TDN studies (see online supplementary figure S2). The CFF-supported Data Safety Monitoring Board located at the University of Arizona has oversight of clinical trials at all centres. The TDCs are selected by a competitive, peer-reviewed application process from the 120 US CF care centres<sup>4 5</sup> and receive infrastructure support for a research coordinator and other necessary staff. As a result, TDCs have been able to retain well trained, experienced research staff teamed with site investigators. Over the past 15 years, the CF-TDN has successfully completed over 75 clinical trials along the entire developmental path from phase I to phase III registration trials. The goal to bring new therapies to the CF community has been achieved with FDA approval of several therapies (table 1).<sup>6</sup> A key factor in the success of the CF-TDN has been its partnership with industry, providing a full range of consultative services from the earliest discussions about protocol development through critical protocol review to feedback on the industry/contract research organisation experience from the study site perspective. Above all, industry sponsors value the CF-TDN's experience in the conduct of CF clinical research in the USA.

The coordinated efforts of the CF-TDN also include the development of novel outcome measures (below), use of banked specimens and data to conduct ancillary studies,<sup>7 8</sup> and development of novel approaches to study design and analyses.<sup>9</sup> In addition, the network has focused on a quality improvement (QI) initiative based upon groundbreaking work of the CF care centre programme.<sup>5 10</sup> The need for improved communications and a QI programme became essential when the CF-TDN expanded from 8 to 77 TDC sites from 1998 to 2009, encompassing a population of 19 000 patients. Communication was improved using quarterly newsletters, conference calls, annual in-person meetings, and the creation of the CF-TDN website, CF Clinical Research Net, which tracks site study activity, provides tools for site training and programme enhancement, hosts committee work, posts news and information regarding studies and network programmes, and provides access to the network database. A commitment to QI for all aspects of clinical studies has paralleled the growth in network size. Here we summarise recent progress of the network across key therapeutic areas (see CF-TDN Pipeline depicted on the front cover).

**Table 1** CF therapeutics evaluated by the TDN and approved by the FDA

Medication	Therapeutic area	FDA approval
Pancrelipase (Creon, Zenpep) (Pancreaze) (Pertzye) (Ultresa)	Pancreatic enzyme replacement therapy	2009 2010 2012 2012
Aztreonam lysate for inhalation (Cayston)	Inhaled antibiotic	2010
Ivacaftor (Kalydeco)	CFTR potentiator	2012

CF, cystic fibrosis; TDN, Therapeutics Development Network.

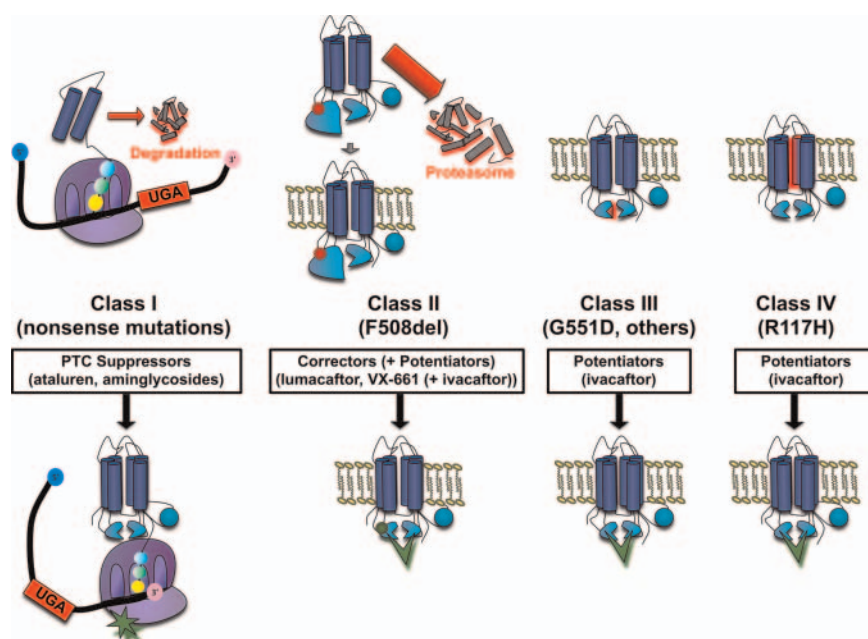
## CFTR MODULATORS

CFTR modulators have brought a personalised approach to altering the basic defect in the disease using gene class specific therapy (see reviews<sup>11–15</sup>). By grouping CFTR mutations into causative classes, specific approaches can be developed that address the underlying molecular defect on an individual patient level (figure 1), and serves as a paradigm for the treatment of other genetic diseases.<sup>16</sup> The evaluation of these novel agents required expertise in proof-of-concept clinical trial design, biomarkers of pharmacological activity, and clinical outcome measures in an entirely new therapeutic area, and were promoted by close partnerships between the pharmaceutical industry and the CFF. Recent results have firmly established that rescue of CFTR-mediated anion transport can result in marked clinical benefit if CFTR functional improvement is sufficient,<sup>6</sup> a concept likely to be a major contributor to CF-TDN efforts in the future.

CFTR modulator therapies have been directed towards specific disease-causing mutations and the molecular pathways that underlie their cause.<sup>12–16</sup> A crucial phase II trial that tested the CFTR potentiator ivacaftor (Kalydeco, VX-770, Vertex Pharmaceuticals, Boston, MA, USA) in 40 patients with CF with at least one copy of the G551D mutation, a relatively common class 3 gating mutant, in a two-part randomised

placebo-controlled design was the first to definitively establish that CFTR functional rescue detected by simultaneous improvement in nasal potential difference (NPD) and sweat chloride could result in meaningful changes in lung function.<sup>17</sup> This was rapidly followed by two phase III trials in patients with G551D CF to establish the clinical effect of ivacaftor during a sustained period. In a trial in older children and adults (aged 12 and above), forced expiratory volume in 1 s (FEV<sub>1</sub>) improved 10.5% at 24 weeks, an effect durable at 48 weeks and accompanied by a 55% reduction in the probability of experiencing a pulmonary exacerbation, a 3.1 kg weight gain (compared with 0.9 kg in the placebo group), and improved respiratory symptoms as assessed by the CFQ-R, a patient reported quality-of-life index.<sup>6</sup> As with phase II testing, sweat chloride improved by around 48 mEq/litre, and mean concentration was around 55 mEq/litre, a level below the traditional diagnostic threshold of CF (60 mEq/litre). Similar results were reported in a study of patients with G551D aged 6–12,<sup>18</sup> enabling prompt approval of the drug for patients with G551D CF aged 6 and above by the FDA and European regulatory authorities. The CF-TDN will support additional studies to evaluate the safety and pharmacokinetics of ivacaftor in younger patients with CF (aged 2–5), which could also begin to assess the effects of early CFTR rescue on the pancreas. The rapid approval and availability of ivacaftor has also enabled the G551D observational study (GOAL), which will assess the clinical effects of ivacaftor in patients with G551D CF post-approval, including the response of several mechanistic biomarkers such as sweat rate, mucociliary clearance imaging, gastrointestinal pH, and sputum measures of inflammation and microbiology; these studies are intended to advance our understanding of the effects of efficacious CFTR modulation on disease-relevant biomarkers and pathways. Since ivacaftor also exhibits robust activity in other rare but clinically relevant CFTR gating mutations, the drug is also being studied in patients with CFTR gating mutations other than G551D. Ivacaftor will also be tested in the archetype conductance mutation R117H, which could establish whether potentiation of CFTR gating is sufficient to partially ameliorate non-gating mutations, setting the stage for other studies involving rare mutations localised to the cell surface.

**Figure 1** Cystic fibrosis transmembrane conductance regulator gene mutation classes and therapeutic approaches under study by the cystic fibrosis Therapeutics Development Network (CF-TDN). Note: approaches to class V (splicing mutations that alter surface CFTR levels), and class VI (mutations in the PDZ binding domain that decrease surface stability and/or alter recycling) CFTR mutations are under development, but have not yet entered clinical testing through the TDN.



Since ivacaftor alone is not sufficient to meaningfully alter CFTR activity in patients with CF homozygous for F508del,<sup>19</sup> a class 2 mutant and the most common CFTR mutation, 'correctors' of CFTR misfolding are required to augment localisation of the F508del CFTR mutation to the cell surface. The CFTR correctors lumacaftor (VX-809, Vertex Pharmaceuticals, Cambridge, Massachusetts, USA) and VX-661 (Vertex Pharmaceuticals) have entered clinical testing within the CF-TDN for this purpose. While monotherapy with lumacaftor in adults homozygous for the F508del mutation only modestly reduced sweat chloride (8 mEq effect vs placebo at the highest dose tested (200 mg)) and did not improve lung function or NPD, evidence of sweat chloride improvement, albeit small in magnitude, suggested correction of the F508del CFTR protein was possible.<sup>20</sup> To improve upon the effects of a corrector alone, lumacaftor is presently being tested at higher doses in a three-part study alone and in combination with ivacaftor, thereby evaluating the effects of augmenting CFTR channel gating following partial restoration of CFTR to the cell surface, a strategy justified by preclinical research<sup>21 22</sup> and the known effects of the F508 deletion on gating in addition to cellular processing.<sup>23</sup> Interim results through part 2 of the study have indicated that high doses of lumacaftor combined with ivacaftor improves FEV<sub>1</sub> by 7.4%, and have recently provided the basis for a pivotal programme to evaluate combination therapy in patients with CF homozygous for F508del that will necessitate involvement in a many sites, including those beyond the network infrastructure. TDCs are also involved in testing the CFTR corrector VX-661 alone and in combination with ivacaftor as an alternate CFTR corrector to lumacaftor. In total, the combination of CFTR potentiators and correctors could result in effective therapies in a large proportion of patients with CF (table 2).

The CFTR modulator class also includes treatment strategies directed towards premature termination codons (class 1 mutations). Following the discovery that aminoglycoside antibiotics can induce readthrough premature termination codons resulting in full-length functional protein,<sup>24</sup> an initial TDN network study focused on topical administration of aminoglycosides<sup>25</sup>; while bioactivity was not detectable and conflicted with other single-centre studies,<sup>26–28</sup> this trial provided experience evaluating CFTR modulators helpful for subsequent studies, including those evaluating the small molecule ataluren (PTC124, premature termination codon therapeutics). Based on efficacy *in vitro*<sup>29</sup> and in animal models,<sup>30</sup> ataluren was studied in a series of conflicting phase II trials,<sup>31–33</sup> one of which involved the

TDN network.<sup>32</sup> Results of a long-term study showed no improvement in FEV<sub>1</sub> % predicted, the primary endpoint, but did demonstrate a small effect on lung function in a predefined subset of individuals who were not treated with inhaled antibiotics, which can alter the efficacy of ataluren.<sup>34</sup> While the clinical status of ataluren remains uncertain at present, given the strong foundation of translational readthrough as a therapeutic strategy for CF and many other genetic diseases, studies involving this compound and other readthrough approaches within the CF-TDN are likely in the future.

## AIRWAY SURFACE LIQUID DEPTH AND MUCUS MODIFICATION

The rationale supporting the development of therapeutics targeting airway surface liquid (ASL) volume and mucus properties comes from basic research on the pathogenesis of CF lung disease.<sup>35</sup> Numerous experimental models support the central role of CFTR as both a chloride channel and as a regulator of other ion channels in airway epithelia.<sup>36 37</sup> Disruption of CFTR activity, in turn, has been linked to the development of defective mucus clearance via depletion of ASL volume and mucus dehydration.<sup>38</sup> These and perhaps other distinct effects on airways defence may ultimately explain the initial, profound vulnerability to chronic bacterial airways infection that exists in CF, while also contributing to disease progression.

Approaches to increase fluid secretion and/or reduce fluid reabsorption to augment impaired mucociliary clearance in CF represent a major line of therapeutics that address fundamental physiological deficits in CF, and are not restricted to specific CFTR mutations nor the uncertainties of gene transfer. A multifaceted approach via collaborations between scientists, industry, the CF Foundation, and the Therapeutic Development Network, utilising different therapeutic targets, has been pursued to maximise the chance of success. These alternative ASL restoring therapeutics can be grouped into those that promote increases in ASL volume and mucus hydration via osmotic mechanisms (eg, hypertonic saline; Bronchitol, Pharmaxis, Frenchs Forest, Australia), by reducing fluid reabsorption from the airway lumen through inhibition of the epithelial sodium channel (eg, GS9411, Gilead Sciences, Foster City, California, USA) or by increasing chloride secretion via various non-CFTR pathways (eg, denufosal, Inspire Pharmaceuticals, Raleigh, North Carolina, USA; Moli 1901, Lantibio, Chapel Hill, North Carolina, USA; SPI-881, Sucampo, Bethesda, Maryland, USA). Within the osmotic hydrator class of therapies, hypertonic saline is already available,<sup>39 40</sup> and successful completion of phase III trials of dry-powder mannitol in the USA and abroad led to the recent approval for use in Australia and the EU, whereas FDA review is presently underway in the USA.<sup>41–43</sup> Addressing abnormal CF mucus, marked by the approval of recombinant human DNase (Pulmozyme, Genentech, San Francisco, California, USA),<sup>44</sup> has also had renewed interest, and is likely to lead to new lines of investigation supported by the network.

The concept of targeting sodium hyperabsorption, and thereby improving ASL hydration, first emerged more than three decades ago.<sup>45 46</sup> The earliest attempts at manipulating sodium transport in the airways of patients with CF utilised inhaled amiloride.<sup>47–49</sup> Treatment with amiloride was ultimately unsuccessful, likely due to inadequate potency and duration of action. Since then, newer agents with improved pharmacokinetic and pharmacodynamic properties have been developed, and early phase trials have been conducted to test

**Table 2** CFTR-based therapies completed or in progress within the Therapeutics Development Network

CFTR mutation	Proportion of patients with causative mutation (%)	Therapeutic approach	Status
G551D/other	4	Ivacaftor	FDA approved, age ≥6 Age 3–5 planned
Non-G551D gating/other	1	Ivacaftor	Phase II/III
R117H/other	5	Ivacaftor	Phase III
F508del/ F508del	49	Lumacaftor + ivacaftor VX-661 + ivacaftor	Phase II; phase III planned Phase II
PTC/other	10	Ataluren	Phase III (primary endpoint negative)

CF, cystic fibrosis; CFTR, CF transmembrane conductance regulator; PTC, premature termination codon.

their safety.<sup>50</sup> Additional research with the latest molecules should be available soon for testing within the CF-TDN.

Stimulation of chloride secretion in the absence of CFTR can be accomplished in vitro and in vivo using agents that stimulate alternative chloride channels, including TMEM16a and others activated by calcium.<sup>51</sup> Testing of an orally dosed prostone (cobiprostone, Sucampo) was conducted by the CF-TDN nearly 10 years ago. Although this study did not demonstrate a clear chloride secretory response, a related compound was eventually approved as a chloride secretagogue for the treatment of chronic constipation, and this study along with other early efforts informed the network on the use of NPD and other techniques now commonly used in CF-TDN network studies.<sup>25–52</sup> Moli1901 (Lantibio) showed early promise as a chloride transport activator in US and European studies,<sup>53</sup> but has not been pursued further due to formulation issues. Denufosal (Inspire Pharmaceuticals), a P2Y<sub>2</sub> agonist designed to restore chloride transport and increase mucociliary clearance, also yielded promising results in early phase studies,<sup>54–55</sup> but ultimately failed after the conduct of two phase III trials.<sup>56–57</sup> Reflection on lessons learned and research into the potential cause of this failure has been undertaken and will educate the search for additional ASL modifying agents.

### ANTI-INFECTIVES

Chronic bacterial infections of the CF airways with resistant organisms, including multidrug resistant (MDR) *Pseudomonas aeruginosa* and other intrinsically resistant organisms such as *Burkholderia cepacia* complex, *Stenotrophomonas maltophilia* and *Achromobacter* species remain a major therapeutic challenge in CF.<sup>58</sup> For example, the current rate of MDR *P aeruginosa* among patients with CF is 9.8%.<sup>59</sup> Because of the severity and relentless nature of CF airway infections and the need to develop new treatment approaches to alleviate the development of resistance, an important focus of TDN-supported studies has been in the area of anti-infectives. Unfortunately, few new antimicrobials directed against *P aeruginosa* have been developed in the past two decades. Thus, recent anti-infective strategies have used currently available antimicrobials in new ways. In addition, novel approaches to treatment have been developed, including suppression of chronic infections and early therapy to eradicate infections prior to irreversible infection.

A sentinel agent in the development of new antimicrobial strategies for treatment of *P aeruginosa* in CF was tobramycin inhalation solution (TOBI, Novartis Pharmaceuticals, East Hanover, New Jersey, USA). Inhalation of antibiotics allows high concentrations of active drug at the site of infection, while minimising side effects. Inhaled tobramycin was tested in two trials including a total of 520 subjects over the age of 6 years. The trials demonstrated improvement in lung function and decrease in bacterial colony counts, and supported the drug's FDA approval in 1994. Although early in the formation of the network, the concept of inhaled suppressive antibiotics and the actual study design of the inhaled tobramycin trials have served as a model for other antimicrobial agents tested by TDCs. Aztreonam lysinate for inhalation (AZLI, Cayston, Gilead Sciences) was initially developed through support of the CFF and the clinical trials were performed through the TDN. Since its approval by the FDA, AZLI has been rapidly used by the CF population, demonstrating the intense need for new antimicrobial therapies. To extend efficacy to alternative organisms, a phase IV trial of AZLI for patients with CF and *B cepacia* complex infections was recently completed and Gilead is conducting an ongoing phase IV CFF Patient

Registry-based study of antibiotic resistance. Other agents that have been formulated for airway administration and are currently in the CF Foundation Therapeutics Pipeline include tobramycin inhaled powder (TOBI Podhaler, Novartis Pharmaceuticals), inhaled levofloxacin (Aeroquin Axcan Pharmaceuticals, San Diego, California, USA), and inhaled liposomal amikacin (Arikace, Insmad, Monmouth Junction, New Jersey, USA), all of which are in phase III trials and offer improved convenience, delivery, and durability. Similarly, ciprofloxacin inhaled powder (Bayer Schering Pharma AG, Berlin-Wedding, Germany), has recently completed a phase II trial in CF, but has not yet begun phase III testing.

Another example of using currently available antimicrobials is the novel use of azithromycin in the management of CF airway infections. Although active in vitro against *Staphylococcus aureus* and *Haemophilus influenzae*, azithromycin is not considered active against the most typical gram-negative CF pathogens. However, studies developed through Cystic Fibrosis Foundation Therapeutics have demonstrated clinical response to three times a week administration of azithromycin. This was initially shown in a large clinical trial in patients with CF and *P aeruginosa* infection who were over the age of 6 and subsequently confirmed in younger children and those uninfected with *P aeruginosa*.<sup>60–61</sup> The exact mechanism of action has not been demonstrated but patients responded with an improvement in FEV<sub>1</sub> and a decreased frequency of pulmonary exacerbations.

Studies within the TDN have examined the role of early antibiotic therapy in eradicating early infection and delaying the establishment of chronic infections in young children.<sup>62–64</sup> The Early *Pseudomonas* Infection Control (EPIC) trial was a multicentre randomised trial comparing one-drug or two-drug therapy (TOBI with or without oral ciprofloxacin) with either cycled or culture-dependent administration to eradicate early *P aeruginosa* infection. It demonstrated equal efficacy and safety in all arms without emergence of multiple antibiotic resistant organisms.<sup>65–66</sup>

A new antimicrobial soon to begin phase II testing is KB001 (KaloBios Pharmaceuticals, South San Francisco, California, USA), a monoclonal antibody fragment directed against the PcrV protein of the type 3 secretion system of *P aeruginosa*.<sup>67</sup> Other antimicrobial therapies on the horizon include trials for the management of methicillin-resistant *S aureus* and non-tuberculous mycobacteria, and intravenous administration of gallium, an agent that is not a conventional antibiotic but has antimicrobial properties and is currently used by radiologists for nuclear medicine scans.

### ANTI-INFLAMMATORY THERAPIES

Neutrophil-dominated airway inflammation is a hallmark of CF lung disease (see previous reviews<sup>68–71</sup>); thus, evaluating drugs that target inflammation is a major emphasis of the network. Corticosteroids and high-dose ibuprofen, each broad spectrum inhibitors of pro-inflammatory signalling, were two of the first anti-inflammatory drugs studied in CF. Both demonstrated clinical benefit,<sup>72–74</sup> but side effects and other considerations have limited their use.<sup>75–77</sup> However, the evidence that these anti-inflammatory agents can slow the progression of CF lung disease, particularly in children, suggests that strategies to modulate lung inflammation can be beneficial.

The CF-TDN has devoted significant resources to evaluating anti-inflammatory treatments. Among those successful include azithromycin, hydroxychloroquine which has been shown to reduce pulmonary exacerbations and improve lung function in

patients with CF, without significantly affecting lower airway bacterial density<sup>60 61</sup> (see Anti-infectives section). The evidence suggests that azithromycin may act as an anti-inflammatory or immunomodulatory agent, which may also explain its efficacy in chronic obstructive pulmonary disease.<sup>78</sup> In a trial of azithromycin in patients with CF chronically infected with *P aeruginosa*, there was a modest reduction in sputum neutrophil elastase activity in favour of the azithromycin group.<sup>61</sup> In a subsequent clinical trial in patients with CF, 6–18 years of age and uninfected with *P aeruginosa*, azithromycin significantly reduced circulating neutrophil counts and systemic markers of inflammation, including C-reactive protein, serum amyloid A and calprotectin.<sup>79</sup>

Several CF-TDN-supported studies evaluating more targeted anti-inflammatory agents have been largely unsuccessful, and a few were associated with detrimental side effects. A notable example of the latter was a trial testing the leukotriene B4 receptor antagonist amelubant (Boehringer Ingelheim, Bracknell, UK) in patients with CF and mild to moderate lung disease.<sup>80</sup> The study was terminated early due to a significant increase in the frequency of pulmonary exacerbations in adults receiving amelubant. This trial gave the CF community pause, providing a potent reminder that modulating inflammation is not without risks. Based on preclinical models and associated clinical data,<sup>81</sup> it was hypothesised that treatment with interferon  $\gamma$  (IFN $\gamma$ ), a pleiotropic cytokine with immunomodulatory and antimicrobial activities, would be beneficial in CF. However, a clinical trial with inhaled IFN- $\gamma$ 1b (Actimmune, InterMune, Brisbane, California, USA) did not significantly improve lung function, sputum bacterial density or sputum biomarkers of inflammation.<sup>82</sup> The CF-TDN network also sponsored studies of 'low-hanging fruit' anti-inflammatory compounds (ie, available medications used for other inflammatory disorders), including the 3-hydroxy-3-methyl coenzyme A reductase inhibitor simvastatin, the peroxisome proliferator-activated receptor  $\gamma$  agonist pioglitazone, and the immunosuppressants methotrexate and hydroxychloroquine. Data from these small, short studies were largely negative, while methotrexate was associated with an intolerable adverse event profile.<sup>83–86</sup> These compounds are no longer being actively investigated, and indicate the complexities of a targeted anti-inflammatory approach in CF lung disease. Additionally, recent 6-month antioxidant trials of inhaled glutathione and oral N-acetylcysteine, a glutathione prodrug, did not demonstrate positive effects either on inflammation, oxidative stress, or clinical outcomes in patients with CF.<sup>87 88</sup>

Despite these disappointments, there are several ongoing studies of potential therapeutic agents which address airway inflammation. Numerous studies have demonstrated an excess burden of neutrophil-derived proteases in the CF airway and provide justification for interventions that neutralise free neutrophil elastase activity or augment local antiprotease levels. There has been renewed interest by the pharmaceutical industry to develop  $\alpha_1$ -antitrypsin therapies in CF and clinical trials using more potent and efficacious formulations are once again being planned. Antioxidant deficiencies remain a viable therapeutic target in CF. A trial of a reformulated oral multivitamin supplement (AquADEKs), enriched with several non-enzymatic antioxidants including  $\beta$ -carotene, mixed carotenoids, coenzyme Q10, mixed tocopherols and selenium, will evaluate the effects of this antioxidant 'cocktail' on antioxidant levels and markers of oxidative stress. Other novel approaches include the phosphodiesterase inhibitor sildenafil on airway inflammation and the effects of sulforaphane (found in broccoli sprouts) on Nrf2 activation. Finally, a crucial question in the new era of

CFTR modulation is whether improving CFTR activity will have a detectable impact on airway inflammation, a question addressed in the GOAL study (see CFTR modulation section). The breadth of these studies using the CF-TDN network suggest targeting inflammation in CF remains an attractive therapeutic approach, but optimising anti-inflammatory effects while minimising any detrimental impact on host defence remains a key challenge.

## NUTRITIONAL THERAPIES

Good nutrition is vital in CF,<sup>89</sup> thus the TDN has investigated interventions to support normal growth. Almost 90% of patients with CF are pancreatic insufficient<sup>90</sup> and need life-long treatment with pancreatic enzyme replacement therapy (PERT) to prevent malnutrition. Because some pancreatic enzymes products were available before passage of the 1937 Pure Food and Drug Act, drug companies had been allowed to manufacture and sell PERT without prior FDA approval.<sup>91</sup> In 1994 case reports of fibrosing colonopathy (FC) began to appear; a subsequent analysis of CF Foundation Patient Registry data showed a strong correlation between dose of PERT and the appearance of FC.<sup>92</sup> Concerns regarding PERT overuse, and other issues (eg, enteric coating and high-dose capsules) were raised as potential contributing factors. As a result of these and other problems, in 2004 the FDA issued notice that they would require a new drug application for PERT with a deadline of April 2008,<sup>93</sup> which was subsequently extended to 2010. The CF-TDN played a central role in developing study protocols and implementing studies of the efficacy of PERT in this short but urgent time window. Based on these efforts, the FDA has approved five oral delayed-release PERTs, Creon (Creon Abbott Laboratories, Abbott Park, IL, USA),<sup>94</sup> Zenpep (Zenpep Aptalis Pharma US, Inc., Birmingham, AL, USA),<sup>95 96</sup> Pancreaze (Pancreaze Janssen Pharmaceuticals, Titusville, NJ, USA),<sup>97 98</sup> Pertzeye (Pertzeye Digestive Care, Inc., Bethlehem, PA, USA)<sup>99</sup> and Ultresa (Aptalis Pharma US, Inc., Birmingham, AL, USA).

A longitudinal observational study of nutritional status in infants (BONUS) is underway, cosponsored by the National Institute of Health. With newborn screening for CF available in all 50 states as of 2010, PERT is being used in a larger number of infants, yet dose response has never been defined. A nested sub-study of BONUS will explore the efficacy of higher and lower doses of PERT in infants.

All current PERTs are biological products derived from hog pancreas. Although current FDA-approved products are no longer overfilled, have improved stability and are free of enveloped viruses, non-enveloped viruses cannot be removed from the drug substance without loss of efficacy. The CF-TDN has participated in studies of lipotamase, a novel formulation of recombinant bacterial lipase, fungal protease and amorphous amylase that is a non-biological PERT.<sup>100–103</sup> The crystalline nature of some of its components confers stability in an acid environment and makes a liquid preparation possible, providing a potential advantage.

Progression of CF lung disease is linked to oxidative stress.<sup>104</sup> Pancreatic insufficiency and a diminished bile acid pool cause malabsorption of important antioxidants including fat-soluble vitamins (vitamins A, D, E and K), carotenoids, tocopherols, selenium and coenzyme Q<sub>10</sub>.<sup>105</sup> A number of studies have shown that in patients with CF, supplementation can correct many micronutrient deficiencies, although the evidence of clinical benefit has been hampered by inconsistent study design.<sup>106</sup> The CF-TDN participated in developing an oral antioxidant-enriched multivitamin supplement that safely increased systemic

antioxidant levels<sup>107</sup> and continues efforts to explore the benefits of this nutritional approach to modify CF disease.

## BIOMARKERS

Since the inception of the CF-TDN, the development of biomarkers to enhance CF clinical trial planning and conduct has been at the forefront of network activity since alternatives to hard clinical endpoints such as mortality and hospitalisation are not viable endpoints for phase II and many phase III programmes.<sup>4</sup> Biomarkers are 'a characteristic that is objectively measured and evaluated as an indicator of a normal biologic process, pathogenic process, or pharmacologic response to a therapeutic intervention'.<sup>108</sup> As such, CF-TDN-supported research has concentrated on the testing and validation of new biomarkers, linking changes in biomarkers to established surrogate or clinical outcome measures, and refining biomarker assessment techniques largely in the context of drug development. In addition, the acquisition of high-quality longitudinal biospecimens linked to clinical data has been incorporated into several CF-TDN clinical trials, leading to the development of rich specimen banks for biomarker discovery. For example, these biobanks have been leveraged by the CF research community to help define relationships between sputum inflammatory markers (eg, interleukin-8, human neutrophil elastase) and clinical outcome measures (eg, lung function),<sup>8 109</sup> and to relate macrolide therapy to anti-inflammatory activity.<sup>79</sup> The capacity to interrogate samples obtained from multicentre studies that have been collected with common standard operating procedures also increases the probability that results can be successfully recapitulated in the clinic.

With the development of CFTR modulators, there has been a need to develop and refine biomarkers related to augmented CFTR function. From these efforts, sweat chloride has emerged as a robust, standardised and validated method to monitor the bioactivity of CFTR potentiators and correctors.<sup>6 17 19 20 110</sup> NPD has also performed reasonably well as a CFTR biomarker,<sup>111–113</sup> successfully detecting ivacaftor activity in patients with G551D CF. For unclear reasons, neither CFTR biomarker has correlated with clinical outcomes (eg, lung function, patient-reported outcomes (PROs), weight) on an individual patient basis. Ongoing CF-TDN trials are underway to better understand this gap; one example is the GOAL study, which includes the evaluation of functional biomarkers (see CFTR modulation section), banking of serum, sputum, and urine linked to clinical data, and may help future biomarker discovery.

A critical barrier that can potentially limit the extension of new therapies to the treatment of infants and toddlers with CF (and other patients with very mild obstructive disease) is the relative lack of biomarkers sensitive to small changes in these populations. It is a widely held opinion that the greatest benefits conferred by many CF therapies would be expected in the healthiest patients, prior to the development of irreversible lung injury. As one example of success in this area, the CF-TDN developed standard operating procedures for the conduct of infant pulmonary function tests for multicentre performance and analysis.<sup>114</sup> These efforts were used to demonstrate improved lung function (change in FEV<sub>1</sub> in 0.5 s) in infants and toddlers with CF treated with 7% hypertonic saline compared with normal saline, despite no detectable difference in pulmonary exacerbation rates between the two treatment groups.<sup>115</sup> Other CF-TDN studies in progress are geared towards accelerating drug development in the infant population, including the Baby Observational and Nutritional Study

which also includes a biospecimen collection programme (see also above). Finally, TDCs have participated in non-invasive techniques to monitor mild lung disease and responses to intervention, including CT imaging<sup>55 116–119</sup> and Lung Clearance Index measurements in ivacaftor-treated patients with G551D CF. Further studies that link sensitive disease biomarkers to clinical endpoints in the setting of new and established therapies could help accelerate the development path required to bring therapies to the healthiest patients with CF.

The TDN has played an instrumental role in advancing CF PROs and helping define their role in drug development. This work was particularly instrumental during the development of AZLI.<sup>120–122</sup> PROs have also served as primary and secondary endpoints in pivotal trials involving CF therapeutics; establishing the minimal clinically important difference for the Respiratory Domain of the CF Questionnaire-Revised (CFQ-R) was critical to this effort.<sup>123</sup> This work has paved the way for the incorporation of the CFQ-R into clinical trials of many new CF treatment strategies.<sup>6 17 19 20 124 125</sup> Future studies are planned to further refine CF PROs, including the development of PROs for non-pulmonary disease manifestations, correlation of PRO results with other biomarkers, and the capture of information more frequently (eg, daily diaries compared with 2-week recall that is a limitation of the current CFQ-R).

## FUTURE DIRECTIONS FOR THE NETWORK

As novel therapies emerge and evolve, clinical trials infrastructures must respond to the expanding need for studies. The CF-TDN and its partner network, the European Society for Cystic Fibrosis (ESCF) Clinical Trials Network, must maintain high-quality study conduct, which will be optimised by providing the structure and tools for improvement through web-based, QI programmes. These programmes are available to all sites with the requirement that they participate in an assessment–prioritisation–implementation–reflection cycle and report their results annually. A competitive application process will continue to sustain an engaged network of participating sites that allows new sites to enter the network when they

**Table 3** Lessons learned by the TDN and changes implemented to address them

Lessons learned	Actions to address challenges
In an expanding network, different sites have variable experience and knowledge base regarding conduct of clinical research	Defined education programmes including self-education modules, published standard operating procedures, etc
Variable performance in study start-up time and enrolment rate across TDN sites	Established a central database to track performance, provide sites real-time feedback, annual peer comparisons of metrics and a web-based quality improvement programme
Variability in important clinical endpoints and biomarkers can adversely impact interpretation of results	Established National Resource Centres to provide expertise in particular endpoints and provide quality assurance to individual TDCs. Established and published standard operating procedures for outcomes
Increase in number of interested sponsors and therapeutics to be tested creates stress on recruitment capabilities and study throughput	Steady expansion of clinical network through competitive renewal programme that emphasises performance metrics and quality study conduct
Variable experience and understanding of CF by industry sponsors	Developed TDN sponsored consulting programme to provide consistent advice to sponsors and to enable rapid development of clinical protocols

CF, cystic fibrosis; TDC, Therapeutic Development Centre; TDN, Therapeutics Development Network.

demonstrate interest and capability, and permits adjusting network size to meet enrolment projections for studies linked to the research pipeline.

The CF-TDN will continue to work cooperatively with the ESCF Clinical Trials Network and the Canadian and Australian CF research communities to facilitate the conduct of uniform protocols across multiple countries, to enrol larger sample sizes in a shorter time period, and permit sufficient recruitment of subsets of patients with unique characteristics, such as age, genotype, infecting organism or comorbidities. Collaboration across networks also exists for combined protocol reviews, development of industry and investigator initiated studies, and working groups developing consensus around novel outcome measures and endpoint definitions. Similarly, the network has made a consistent effort to learn from challenges experienced, and address these areas in the future; some important examples are noted in table 3.

The development of novel or improved outcome measures remains a major focus for CF-TDN working groups; in particular, assessing outcome measures applicable for infants and young children as new therapies become available and decisions regarding efficacy in younger populations must be made, for example.<sup>126</sup> The existence of the CF Patient Registry enhances the conduct of clinical research in many ways, and will be a pivotal tool for implementation of comparative effectiveness research (CER) in CF. The tool has already been used in long-term observational studies conducted through the CF-TDN.<sup>127</sup> As this capability grows, it is conceivable that it will allow the expansion of future CER and observational studies to all CF care centres that participate in the registry, maximising investigative power. Combined with the robust pipeline of therapeutics, the network is well poised to make continued contributions to the rapid evolution of CF care.

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## REFERENCES

1. **US Food and Drug Administration.** FDA definition 'orphan disease'. <http://www.fda.gov/ForIndustry/DevelopingProductsforRareDiseasesConditions/HowtoapplyforOrphanProductDesignation/DefinitionofDiseasePrevalence/default.htm> (accessed 23 Aug 2012)
2. **Pattishall EN.** Negative clinical trials in cystic fibrosis research. *Pediatrics* 1990;**85**:277–81.
3. **Goss CH, Mayer-Hamblett N, Williams J, et al.** The cystic fibrosis foundation therapeutics development network: a national effort by the cystic fibrosis foundation to build a clinical trials network. *Child Health Care* 2008;**37**:5–20.
4. **Goss CH, Mayer-Hamblett N, Kronmal RA, et al.** The cystic fibrosis therapeutics development network (CF TDN): a paradigm of a clinical trials network for genetic and orphan diseases. *Adv Drug Deliv Rev* 2002;**54**:1505–28.
5. **Marshall BC, Penland CM, Hazle L, et al.** Cystic fibrosis foundation: achieving the mission. *Respir Care* 2009;**54**:788–95; discussion 95.
6. **Ramsey BW, Davies J, McElvaney NG, et al.** A CFTR potentiator in patients with cystic fibrosis and the G551D mutation. *N Engl J Med* 2011;**365**:1663–72.
7. **Emond MJ, Louie T, Emerson J, et al.** Exome sequencing of extreme phenotypes identifies DCTN4 as a modifier of chronic *Pseudomonas aeruginosa* infection in cystic fibrosis. *Nat Genet* 2012;**44**:886–9.
8. **Mayer-Hamblett N, Aitken ML, Accurso FJ, et al.** Association between pulmonary function and sputum biomarkers in cystic fibrosis. *Am J Respir Crit Care Med* 2007;**175**:822–8.
9. **Mayer-Hamblett N, Kronmal RA.** Improving the estimation of change from baseline in a continuous outcome measure in the clinical trial setting. *Contemp Clin Trials* 2005;**26**:2–16.
10. **Quon BS, Goss CH.** A story of success: continuous quality improvement in cystic fibrosis care in the USA. *Thorax* 2011;**66**:1106–8.
11. **Clancy JP, Jain M.** Personalized medicine in cystic fibrosis: dawning of a new era. *Am J Respir Crit Care Med* 2012.
12. **Sloane PA, Rowe SM.** Cystic fibrosis transmembrane conductance regulator protein repair as a therapeutic strategy in cystic fibrosis. *Curr Opin Pulm Med* 2010;**16**:591–7.
13. **Davis PB, Yasothan U, Kirkpatrick P.** Ivacaftor. *Nat Rev Drug Discov* 2012;**11**:349–50.
14. **Corbyn Z.** Promising new era dawns for cystic fibrosis treatment. *Lancet* 2012;**379**:1475–6.
15. **Kaiser J.** Personalized medicine. New cystic fibrosis drug offers hope, at a price. *Science* 2012;**335**:645.
16. **Rowe SM, Miller S, Sorscher EJ.** Cystic fibrosis. *N Engl J Med* 2005;**352**:1992–2001.
17. **Accurso FJ, Rowe SM, Clancy JP, et al.** Effect of VX-770 in persons with cystic fibrosis and the G551D-CFTR mutation. *N Engl J Med* 2010;**363**:1991–2003.
18. **Aherns R, Rodriguez S, Yen K, et al.** VX-770 in subjects 6 to 11 years with cystic fibrosis and the G551D-CFTR mutation. *Ped Pulm Suppl* 2011;**46**:A198.
19. **Flume PA, Liou TG, Borowitz DS, et al.** Ivacaftor in subjects with cystic fibrosis who are homozygous for the F508del-CFTR mutation. *Chest* 2012. Published Online First: 1 March 2012. doi: 10.1378/chest.111-2672
20. **Clancy JP, Rowe SM, Accurso FJ, et al.** Results of a phase IIa study of VX-809, an investigational CFTR corrector compound, in subjects with cystic fibrosis homozygous for the F508del-CFTR mutation. *Thorax* 2011;**67**:12–18.
21. **Van Goor F, Hadida S, Grootenhuys PD, et al.** Correction of the F508del-CFTR protein processing defect in vitro by the investigational drug VX-809. *Proc Natl Acad Sci U S A* 2011;**108**:18843–8.
22. **Van Goor F, Hadida S, Grootenhuys PD, et al.** Rescue of CF airway epithelial cell function in vitro by a CFTR potentiator, VX-770. *Proc Natl Acad Sci U S A* 2009;**106**:18825–30.
23. **Wang F, Zeltwanger S, Hu S, et al.** Deletion of phenylalanine 508 causes attenuated phosphorylation-dependent activation of CFTR chloride channels. *J Physiol* 2000;**524**(Pt 3):637–48.
24. **Bedwell DM, Kaenjak A, Benos DJ, et al.** Suppression of a CFTR premature stop mutation in a bronchial epithelial cell line. *Nat Med* 1997;**3**:1280–4.
25. **Clancy JP, Rowe SM, Bekok Z, et al.** No detectable improvements in cystic fibrosis transmembrane conductance regulator by nasal aminoglycosides in patients with cystic fibrosis with stop mutations. *Am J Respir Cell Mol Biol* 2007;**37**:57–66.
26. **Clancy JP, Bekok Z, Ruiz F, et al.** Evidence that systemic gentamicin suppresses premature stop mutations in patients with cystic fibrosis. *Am J Respir Crit Care Med* 2001;**163**:1683–92.
27. **Wilschanski M, Yahav Y, Yaacov Y, et al.** Gentamicin-induced correction of CFTR function in patients with cystic fibrosis and CFTR stop mutations. *N Engl J Med* 2003;**349**:1433–41.
28. **Sermet-Gaudelus I, Renouil M, Fajac A, et al.** In vitro prediction of stop-codon suppression by intravenous gentamicin in patients with cystic fibrosis: a pilot study. *BMC Med* 2007;**5**:5–14.
29. **Welch EM, Barton ER, Zhuo J, et al.** PTC124 targets genetic disorders caused by nonsense mutations. *Nature* 2007;**447**:87–91.
30. **Du M, Liu X, Welch EM, et al.** PTC124 is an orally bioavailable compound that promotes suppression of the human CFTR-G542X nonsense allele in a CF mouse model. *Proc Natl Acad Sci U S A* 2008;**105**:2064–9.
31. **Kerem E, Hirawat S, Armoni S, et al.** Effectiveness of PTC124 treatment of cystic fibrosis caused by nonsense mutations: a prospective phase II trial. *Lancet* 2008;**372**:719–27.
32. **Clancy JP, Konstan MW, Rowe SM, et al.** A phase II study of PTC124 in CF patients harboring premature stop mutations. *Ped Pulmonol Suppl* 2006;**41**(Suppl 29):abstract 269.
33. **Sermet-Gaudelus I, Boeck KD, Casimir GJ, et al.** Ataluren (PTC124) induces cystic fibrosis transmembrane conductance regulator protein expression and activity in children with nonsense mutation cystic fibrosis. *Am J Respir Crit Care Med* 2010;**182**:1262–72.
34. **Konstan M, Accurso F, De Boeck K, et al.** Results of the phase 3 study of ataluren in nonsense mutation cystic fibrosis (nmCF). *J Cyst Fibros* 2012;**11**(Suppl 1).
35. **Matsui H, Grubb BR, Tarran R, et al.** Evidence for periciliary liquid layer depletion, not abnormal ion composition, in the pathogenesis of cystic fibrosis airways disease. *Cell* 1998;**95**:1005–15.
36. **Stutts MJ, Canessa CM, Olsen JC, et al.** CFTR as a cAMP-dependent regulator of sodium channels [see comments]. *Science* 1995;**269**:847–50.
37. **Tarran R, Loewen ME, Paradiso AM, et al.** Regulation of murine airway surface liquid volume by CFTR and Ca<sup>2+</sup>-activated Cl<sup>-</sup> conductances. *J Gen Physiol* 2002;**120**:407–18.
38. **Tarran R, Button B, Picher M, et al.** Normal and cystic fibrosis airway surface liquid homeostasis: the effects of phasic shear stress and viral infections. *J Biol Chem* 2005;**280**:35751–9.

39. **Elkins MR**, Robinson M, Rose BR, *et al*. A controlled trial of long-term inhaled hypertonic saline in patients with cystic fibrosis. *N Engl J Med* 2006;**354**:229–40.
40. **Donaldson SH**, Bennett WD, Zeman KL, *et al*. Mucus clearance and lung function in cystic fibrosis with hypertonic saline. *N Engl J Med* 2006;**354**:241–50.
41. **Bilton D**, Robinson P, Cooper P, *et al*. Inhaled dry powder mannitol in cystic fibrosis: an efficacy and safety study. *Eur Respir J* 2011;**38**:1071–80.
42. **Aitken ML**, Bellon G, De Boeck K, *et al*. Long-term inhaled dry powder mannitol in cystic fibrosis: an international randomized study. *Am J Respir Crit Care Med* 2012;**185**:645–52.
43. **Minasian C**, Wallis C, Metcalfe C, *et al*. Comparison of inhaled mannitol, daily rhDNase and a combination of both in children with cystic fibrosis: a randomised trial. *Thorax* 2010;**65**:51–6.
44. **Fuchs HJ**, Borowitz DS, Christiansen DH, *et al*. Effect of aerosolized recombinant human DNase on exacerbations of respiratory symptoms and on pulmonary function in patients with cystic fibrosis. The Pulmozyme Study Group. *N Engl J Med* 1994;**331**:637–42.
45. **Willumsen NJ**, Boucher RC. Transcellular sodium transport in cultured cystic fibrosis human nasal epithelium. *Am J Physiol* 1991;**261**(2 Pt 1):C332–41.
46. **Knowles M**, Gatzky J, Boucher R. Increased bioelectric potential difference across respiratory epithelia in cystic fibrosis. *N Engl J Med* 1981;**305**:1489–95.
47. **Knowles MR**, Church NL, Waltner WE, *et al*. A pilot study of aerosolized amiloride for the treatment of lung disease in cystic fibrosis. *N Engl J Med* 1990;**322**:1189–94.
48. **App EM**, King M, Helfesrieder R, *et al*. Acute and long-term amiloride inhalation in cystic fibrosis lung disease. A rational approach to cystic fibrosis therapy. *Am Rev Respir Dis* 1990;**141**:605–12.
49. **Graham A**, Hasani A, Alton EW, *et al*. No added benefit from nebulized amiloride in patients with cystic fibrosis. *Eur Respir J* 1993;**6**:1243–8.
50. **Quijana F**, Doran J, Smith R, *et al*. Safety and pharmacokinetics of a single dose of 552-02 following aerosolized administration to patients with cystic fibrosis. *Pediatr Pulmonol* 2005;(Suppl 28):A216.
51. **Mason SJ**, Paradiso AM, Boucher RC. Regulation of transepithelial ion transport and intracellular calcium by extracellular ATP in human normal and cystic fibrosis airway epithelium. *Br J Pharmacol* 1991;**103**:1649–56.
52. **Ahrens RC**, Standaert TA, Launspach J, *et al*. Use of nasal potential difference and sweat chloride as outcome measures in multicenter clinical trials in subjects with cystic fibrosis. *Pediatr Pulmonol* 2002;**33**:142–50.
53. **Zeitlin PL**, Boyle MP, Guggino WB, *et al*. A phase I trial of intranasal Moli1901 for cystic fibrosis. *Chest* 2004;**125**:143–9.
54. **Deterding R**, Retsch-Bogart G, Milgram L, *et al*. Safety and tolerability of denufosal tetrasodium inhalation solution, a novel P2Y2 receptor agonist: results of a phase 1/phase 2 multicenter study in mild to moderate cystic fibrosis. *Pediatr Pulmonol* 2005;**39**:339–48.
55. **Deterding RR**, Lavange LM, Engels JM, *et al*. Phase 2 randomized safety and efficacy trial of nebulized denufosal tetrasodium in cystic fibrosis. *Am J Respir Crit Care Med* 2007;**176**:362–9.
56. **Accurso FJ**, Moss RB, Wilmott RW, *et al*. Denufosal tetrasodium in patients with cystic fibrosis and normal to mildly impaired lung function. *Am J Respir Crit Care Med* 2011;**183**:627–34.
57. **Ratjen F**, Durham T, Navratil T, *et al*. Long term effects of denufosal tetrasodium in patients with cystic fibrosis. *J Cyst Fibros* 2012. Published Online First: 8 June 2012. doi: 10.1016/j.jcf.2012.05.003
58. **Mogayzel PJ Jr**, Flume PA. Update in cystic fibrosis 2010. *Am J Respir Crit Care Med* 2011;**183**:1620–4.
59. Cystic Fibrosis Foundation Patient Registry. *2010 Annual Data Report*, 2011.
60. **Saiman L**, Anstead M, Mayer-Hamblett N, *et al*. Effect of azithromycin on pulmonary function in patients with cystic fibrosis uninfected with *Pseudomonas aeruginosa*: a randomized controlled trial. *JAMA* 2010;**303**:1707–15.
61. **Saiman L**, Marshall BC, Mayer-Hamblett N, *et al*. Azithromycin in patients with cystic fibrosis chronically infected with *Pseudomonas aeruginosa*: a randomized controlled trial. *JAMA* 2003;**290**:1749–56.
62. **Gibson RL**, Emerson J, Mayer-Hamblett N, *et al*. Duration of treatment effect after tobramycin solution for inhalation in young children with cystic fibrosis. *Pediatr Pulmonol* 2007;**42**:610–23.
63. **Gibson RL**, Emerson J, McNamara S, *et al*. Significant microbiological effect of inhaled tobramycin in young children with cystic fibrosis. *Am J Respir Crit Care Med* 2003;**167**:841–9.
64. **Treggiari MM**, Rosenfeld M, Retsch-Bogart G, *et al*. Approach to eradication of initial *Pseudomonas aeruginosa* infection in children with cystic fibrosis. *Pediatr Pulmonol* 2007;**42**:751–6.
65. **Mayer-Hamblett N**, Kronmal RA, Gibson RL, *et al*. Initial *Pseudomonas aeruginosa* treatment failure is associated with exacerbations in cystic fibrosis. *Pediatr Pulmonol* 2012;**47**:125–34.
66. **Treggiari MM**, Retsch-Bogart G, Mayer-Hamblett N, *et al*. Comparative efficacy and safety of 4 randomized regimens to treat early *Pseudomonas aeruginosa* infection in children with cystic fibrosis. *Arch Pediatr Adolesc Med* 2011;**165**:847–56.
67. **Francois B**, Luyt CE, Dugard A, *et al*. Safety and pharmacokinetics of an anti-PcrV PEGylated monoclonal antibody fragment in mechanically ventilated patients colonized with *Pseudomonas aeruginosa*: a randomized, double-blind, placebo-controlled trial. *Crit Care Med* 2012;**40**:2320–6.
68. **Chmiel JF**, Berger M, Konstan MW. The role of inflammation in the pathophysiology of CF lung disease. *Clin Rev Allergy Immunol* 2002;**23**:5–27.
69. **Sagel SD**, Chmiel JF, Konstan MW. Sputum biomarkers of inflammation in cystic fibrosis lung disease. *Proc Am Thorac Soc* 2007;**4**:406–17.
70. **Nichols D**, Chmiel J, Berger M. Chronic inflammation in the cystic fibrosis lung: alterations in inter- and intracellular signaling. *Clin Rev Allergy Immunol* 2008;**34**:146–62.
71. **Elizur A**, Cannon CL, Ferkol TW. Airway inflammation in cystic fibrosis. *Chest* 2008;**133**:489–95.
72. **Eigen H**, Rosenstein BJ, FitzSimmons S, *et al*. A multicenter study of alternate-day prednisone therapy in patients with cystic fibrosis. Cystic Fibrosis Foundation Prednisone Trial Group. *J Pediatr* 1995;**126**:515–23.
73. **Konstan MW**, Byard PJ, Hoppel CL, *et al*. Effect of high-dose ibuprofen in patients with cystic fibrosis. *N Engl J Med* 1995;**332**:848–54.
74. **Lands LC**, Milner R, Cantin AM, *et al*. High-dose ibuprofen in cystic fibrosis: Canadian safety and effectiveness trial. *J Pediatr* 2007;**151**:249–54.
75. **Oermann CM**, Sockrider MM, Konstan MW. The use of anti-inflammatory medications in cystic fibrosis: trends and physician attitudes. *Chest* 1999;**115**:1053–8.
76. **Lai HC**, FitzSimmons SC, Allen DB, *et al*. Risk of persistent growth impairment after alternate-day prednisone treatment in children with cystic fibrosis. *N Engl J Med* 2000;**342**:851–9.
77. **Konstan MW**, VanDevanter DR, Rasouliyan L, *et al*. Trends in the use of routine therapies in cystic fibrosis: 1995–2005. *Pediatr Pulmonol* 2010;**45**:1167–72.
78. **Wenzel RP**, Fowler AA III, Edmond MB. Antibiotic prevention of acute exacerbations of COPD. *N Engl J Med*. [Review] 2012;**367**:340–7.
79. **Ratjen F**, Saiman L, Mayer-Hamblett N, *et al*. Effect of azithromycin on systemic markers of inflammation in cystic fibrosis patients uninfected with *Pseudomonas aeruginosa*. *Chest* 2012. Published Online First: 17 May 2012. doi:10.1378/chest.12-0628
80. **Konstan MW**, Doring G, Lands LC, *et al*. Results of a phase II clinical trial of BILL 284 BS (an LTb4 receptor antagonist) for the treatment of CF lung disease. *Ped Pulmonol Suppl* 2005;**28**(Suppl 7.4):125–6.
81. **Chmiel JF**, Konstan MW. Anti-inflammatory medications for cystic fibrosis lung disease: selecting the most appropriate agent. *Treat Respir Med* 2005;**4**:255–73.
82. **Moss RB**, Mayer-Hamblett N, Wagener J, *et al*. Randomized, double-blind, placebo-controlled, dose-escalating study of aerosolized interferon gamma-1b in patients with mild to moderate cystic fibrosis lung disease. *Pediatr Pulmonol* 2005;**39**:209–18.
83. **Oermann C**, Katz M, Wheeler C, *et al*. A pilot study evaluating the potential use of low-dose methotrexate as an anti-inflammatory for cystic fibrosis lung disease [abstract]. *Pediatr Pulmonol Suppl* 2007;**30**:292–3.
84. **Kraynack N**, Chmiel J, Xue W, *et al*. Effect of simvastatin on exhaled nitric oxide and inflammatory markers in sputum in patients with cystic fibrosis [abstract]. *Pediatr Pulmonol Suppl* 2008;**31**:300.
85. **Konstan M**, Hilliard K, Bucur C, *et al*. Effect of pioglitazone on sputum markers of inflammation in cystic fibrosis [abstract]. *Pediatr Pulmonol Suppl* 2008;**31**:310.
86. **Williams B**, Robinette M, Slovis B, *et al*. Hydroxychloroquine—pilot study of anti-inflammatory effects in cystic fibrosis [abstract]. *Pediatr Pulmonol Suppl* 2008;**31**:314.
87. **Tirouvanziam R**, Lymp J, Thompson V, *et al*. A multi-center, phase IIb, randomized, placebo-controlled, double-blind study of the effects of N-acetylcysteine (NAC) on redox changes and lung inflammation in cystic fibrosis patients [abstract]. *Pediatr Pulmonol Suppl* 2011:280–1.
88. **Griese M**, Hector A, Kappler M, *et al*. Inhaled glutathione in cystic fibrosis [abstract]. *J Cyst Fibros* 2012;**11**:S11.
89. **Stallings VA**, Stark LJ, Robinson KA, *et al*. Evidence-based practice recommendations for nutrition-related management of children and adults with cystic fibrosis and pancreatic insufficiency: results of a systematic review. *J Am Diet Assoc* 2008;**108**:832–9.
90. **Borowitz D**, Baker SS, Duffy L, *et al*. Use of fecal elastase-1 to classify pancreatic status in patients with cystic fibrosis. *J Pediatr* 2004;**145**:322–6.
91. **Hendelees L**, Hochhaus G, Kazerounian S. Generic and alternative brand-name pharmaceutical equivalents: select with caution. *Am J Hosp Pharm* 1993;**50**:323–9.
92. **FitzSimmons SC**, Burkhart GA, Borowitz D, *et al*. High-dose pancreatic-enzyme supplements and fibrosing colonopathy in children with cystic fibrosis. *N Engl J Med* 1997;**336**:1283–9.
93. **FDA**. Exocrine pancreatic insufficiency drug products. *Fed Regist* 2004;**69**:23410–14.
94. **Trapnell BC**, Maguiness K, Graff GR, *et al*. Efficacy and safety of Creon 24 000 in subjects with exocrine pancreatic insufficiency due to cystic fibrosis. *J Cyst Fibros* 2009;**8**:370–7.



95. **Wooldridge JL**, Heubi JE, Amaro-Galvez R, *et al*. EUR-1008 pancreatic enzyme replacement is safe and effective in patients with cystic fibrosis and pancreatic insufficiency. *J Cyst Fibros* 2009;**8**:405–17.
96. **Graff GR**, Maguiness K, McNamara J, *et al*. Efficacy and tolerability of a new formulation of pancrelipase delayed-release capsules in children aged 7 to 11 years with exocrine pancreatic insufficiency and cystic fibrosis: a multicenter, randomized, double-blind, placebo-controlled, two-period crossover, superiority study. *Clin Ther* 2010;**32**:89–103.
97. **Trapnell BC**, Strausbaugh SD, Woo MS, *et al*. Efficacy and safety of PANCREAZE (R) for treatment of exocrine pancreatic insufficiency due to cystic fibrosis. *J Cyst Fibros* 2011;**10**:350–6.
98. Erratum in *J Cyst Fibros* 2011;**10**:491–2.
99. **Digestive Care**. [http://www.digestivecare.com/PDFs/NDA%20022175%20APPROVAL%2017MAY2012\\_Pi&MG.pdf](http://www.digestivecare.com/PDFs/NDA%20022175%20APPROVAL%2017MAY2012_Pi&MG.pdf) (accessed 23 Aug 2012)
100. **Borowitz D**, Goss CH, Stevens C, *et al*. Safety and preliminary clinical activity of a novel pancreatic enzyme preparation in pancreatic insufficient cystic fibrosis patients. *Pancreas* 2006;**32**:258–63.
101. **Borowitz D**, Goss CH, Limauro S, *et al*. Study of a novel pancreatic enzyme replacement therapy in pancreatic insufficient subjects with cystic fibrosis. *J Pediatr* 2006;**149**:658–62.
102. **Borowitz D**, Stevens C, Brettman LR, *et al*. Liprotamase long-term safety and support of nutritional status in pancreatic-insufficient cystic fibrosis. *J Pediatr Gastroenterol Nutr* 2012;**54**:248–57.
103. **Borowitz D**, Stevens C, Brettman LR, *et al*. International phase III trial of liprotamase efficacy and safety in pancreatic-insufficient cystic fibrosis patients. *J Cyst Fibros* 2011;**10**:443–52.
104. **Wood LG**, Fitzgerald DA, Gibson PG, *et al*. Oxidative stress in cystic fibrosis: dietary and metabolic factors. *J Am Coll Nutr* 2001;**20**(2 Suppl):157–65.
105. **Cantin AM**, White TB, Cross CE, *et al*. Antioxidants in cystic fibrosis. Conclusions from the CF antioxidant workshop, Bethesda, Maryland, November 11–12, 2003. *Free Radic Biol Med* 2007;**42**:15–31.
106. **Shamseer L**, Adams D, Brown N, *et al*. Antioxidant micronutrients for lung disease in cystic fibrosis. *Cochrane Database Syst Rev* 2010;(12):CD007020.
107. **Sagel SD**, Sontag MK, Anthony MM, *et al*. Effect of an antioxidant-rich multivitamin supplement in cystic fibrosis. *J Cyst Fibros* 2011;**10**:31–6.
108. **Mayer-Hamblett N**, Ramsey BW, Kronmal RA. Advancing outcome measures for the new era of drug development in cystic fibrosis. *Proc Am Thorac Soc* 2007;**4**:370–7.
109. **Ordonez CL**, Henig NR, Mayer-Hamblett N, *et al*. Inflammatory and microbiologic markers in induced sputum after intravenous antibiotics in cystic fibrosis. *Am J Respir Crit Care Med* 2003;**168**:1471–5.
110. **Ahrens R**, Rodriguez S, Yen K, *et al*. VX-770 in subjects 6 to 11 years with cystic fibrosis and the G551D CFTR genotype. *Pediatr Pulmonol* 2011;**34**(Suppl):283.
111. **Solomon GM**, Konstan MW, Wilschanski M, *et al*. An international randomized multicenter comparison of nasal potential difference techniques. *Chest* 2003;**124**:919–28.
112. **Boyle MP**, Diener-West M, Milgram L, *et al*. A multicenter study of the effect of solution temperature on nasal potential difference measurements. *Chest* 2003;**124**:482–9.
113. **Standaert TA**, Boitano L, Emerson J, *et al*. Standardized procedure for measurement of nasal potential difference: an outcome measure in multicenter cystic fibrosis clinical trials. *Pediatr Pulmonol* 2004;**37**:385–92.
114. **Davis SD**, Rosenfeld M, Kerby GS, *et al*. Multicenter evaluation of infant lung function tests as cystic fibrosis clinical trial endpoints. *Am J Respir Crit Care Med* 2010;**182**:1387–97.
115. **Rosenfeld M**, Ratjen F, Brumback L, *et al*. Inhaled hypertonic saline in infants and children younger than 6 years with cystic fibrosis: the ISIS randomized controlled trial. *JAMA* 2012;**307**:2269–77.
116. **Davis SD**, Brody AS, Emond MJ, *et al*. Endpoints for clinical trials in young children with cystic fibrosis. *Proc Am Thorac Soc* 2007;**4**:418–30.
117. **Davis SD**, Fordham LA, Brody AS, *et al*. Computed tomography reflects lower airway inflammation and tracks changes in early cystic fibrosis. *Am J Respir Crit Care Med* 2007;**175**:943–50.
118. **Brody AS**. Computed tomography scanning in cystic fibrosis research trials: practical lessons from three clinical trials in the United States. *Proc Am Thorac Soc* 2007;**4**:350–4.
119. **Moss RB**, Rodman D, Spencer LT, *et al*. Repeated adeno-associated virus serotype 2 aerosol-mediated cystic fibrosis transmembrane regulator gene transfer to the lungs of patients with cystic fibrosis: a multicenter, double-blind, placebo-controlled trial. *Chest* 2004;**125**:509–21.
120. **Retsch-Bogart GZ**, Quittner AL, Gibson RL, *et al*. Efficacy and safety of inhaled aztreonam lysine for airway pseudomonas in cystic fibrosis. *Chest* 2009;**135**:1223–32.
121. **McCoy KS**, Quittner AL, Oermann CM, *et al*. Inhaled aztreonam lysine for chronic airway Pseudomonas aeruginosa in cystic fibrosis. *Am J Respir Crit Care Med* 2008;**178**:921–8.
122. **Oermann CM**, Retsch-Bogart GZ, Quittner AL, *et al*. An 18-month study of the safety and efficacy of repeated courses of inhaled aztreonam lysine in cystic fibrosis. *Pediatr Pulmonol* 2011;**45**:1121–34.
123. **Quittner AL**, Modi AC, Wainwright C, *et al*. Determination of the minimal clinically important difference scores for the Cystic Fibrosis Questionnaire-Revised respiratory symptom scale in two populations of patients with cystic fibrosis and chronic Pseudomonas aeruginosa airway infection. *Chest* 2009;**135**:1610–18.
124. **Geller DE**, Flume PA, Staab D, *et al*. Levofloxacin inhalation solution (MP-376) in patients with cystic fibrosis with Pseudomonas aeruginosa. *Am J Respir Crit Care Med* 2011;**183**:1510–16.
125. **Konstan MW**, Geller DE, Minic P, *et al*. Tobramycin inhalation powder for P aeruginosa infection in cystic fibrosis: the EVOLVE trial. *Pediatr Pulmonol* 2011;**46**:230–8.
126. **Rosenfeld M**, Ratjen F, Brumback L, *et al*. Inhaled hypertonic saline in infants and children younger than 6 years with cystic fibrosis: the ISIS randomized controlled trial inhaled hypertonic saline in children with CF. *JAMA* 2012;1–9.
127. **Rosenfeld M**, Emerson J, McNamara S, *et al*. Baseline characteristics and factors associated with nutritional and pulmonary status at enrollment in the cystic fibrosis EPIC observational cohort. *Pediatr Pulmonol* 2010;**45**:934–44.