Ventilation heterogeneity is a major determinant of airway hyperresponsiveness in asthma, independent of airway inflammation

Sue R Downie, Cheryl M Salome, Sylvia Verbanck, Bruce Thompson, Norbert Berend, Gregory G King

Background: Airway hyperresponsiveness is the ability of airways to narrow excessively in response to inhaled stimuli and is a key feature of asthma. Airway inflammation and ventilation heterogeneity have been separately shown to be associated with airway hyperresponsiveness. A study was undertaken to establish whether ventilation heterogeneity is associated with airway hyperresponsiveness independently of airway inflammation in subjects with asthma and to determine the effect of inhaled corticosteroids on this relationship.

Methods: Airway inflammation was measured in 40 subjects with asthma by exhaled nitric oxide, ventilation heterogeneity by multiple breath nitrogen washout and airway hyperresponsiveness by methacholine challenge. In 18 of these subjects with uncontrolled symptoms, measurements were repeated after 3 months of treatment with inhaled beclomethasone dipropionate.

Results: At baseline, airway hyperresponsiveness was independently predicted by airway inflammation (partial r² = 0.20, p < 0.001) and ventilation heterogeneity (partial r² = 0.39, p < 0.001). Inhaled corticosteroid treatment decreased airway inflammation (p = 0.002), ventilation heterogeneity (p = 0.009) and airway hyperresponsiveness (p < 0.001). After treatment, ventilation heterogeneity was the sole predictor of airway hyperresponsiveness (r² = 0.64, p < 0.001).

Conclusions: Baseline ventilation heterogeneity is a strong predictor of airway hyperresponsiveness, independent of airway inflammation in subjects with asthma. Its persistent relationship with airway hyperresponsiveness following anti-inflammatory treatment suggests that it is an important independent determinant of airway hyperresponsiveness. Normalisation of ventilation heterogeneity is therefore a potential goal of treatment that may lead to improved long-term outcomes.
derivation of these indices is explained in the online supplement available at http://thorax.bmj.com/supplemental. Using this MBNW analysis, ventilation heterogeneity in conductive and acinar lung zones has been shown to be abnormal in asthma and to be more sensitive than spirometry for detecting early changes to the peripheral airways associated with cigarette smoking.

We hypothesised that there would be an association between ventilation heterogeneity and AHR and have tested whether this association is independent of airway inflammation in a cross-sectional study of subjects with asthma of a wide range of severity. In a subgroup of these subjects with clinically uncontrolled asthma, we further tested the relationship between airway inflammation, ventilation heterogeneity and AHR following 3 months of treatment with ICS.

METHODS
Study design
Subjects in whom asthma had been diagnosed by a respiratory physician according to the NIH guidelines were enrolled in the study. At the baseline visit all subjects underwent the following tests in the order shown: (1) atopic status was determined by skin prick testing with mean weal diameters \( \geq 4 \) mm regarded as positive; (2) Juniper asthma control questionnaire; (3) the fraction of nitric oxide in exhaled breath \((F_{ENO})\) was used as a marker of airway inflammation; (4) MBNW was performed to measure ventilation heterogeneity; and (5) methacholine challenge was undertaken to determine AHR. A subgroup of patients with at least mild persistent asthma, as determined by GINA guidelines, were given ICS for 3 months with either chlorofluorocarbon beclomethasone dipropionate (CFC-BDP) 750 \( \mu \)g twice daily via Autohaler or hydrofluoroalkane beclomethasone dipropionate (HFA-BDP) 400 \( \mu \)g twice daily via Autohaler (clinically equivalent doses). Since our aim was to investigate overall treatment effects, all treatment data were pooled for analyses.

Written informed consent was obtained from all subjects and the study was approved by the human ethics review committee of the South-Western Area Health Service. This study is registered on the Australian Clinical Trials Registry (ACTRN012605000317695).

Subjects
Subjects with asthma were recruited by advertising throughout the University of Sydney. Inclusion criteria for all subjects with asthma were: (1) no smoking within the last 6 months and <10 pack-years smoking history; (2) no current lung disease other than asthma; (3) no oral prednisone use in the last 4 weeks; and (4) no respiratory tract infection in the last 4 weeks. Additional inclusion criteria for entry into the 3 month treatment period were: (1) asthma symptoms on at least three occasions a week over the previous month; (2) AHR defined as a provoking dose of methacholine causing a 20% fall in FEV\(_1\) of \( \leq 6.1 \) \( \mu \)mol; and (3) taking no more than 800 \( \mu \)g/day CFC-BDP equivalent of ICS.

Two cohorts of healthy non-asthmatic subjects were recruited from our research staff and the University of Sydney. The first cohort was recruited for the sensitivity and specificity analysis of a parameter from the MBNW test in detecting AHR, and the second cohort was recruited for testing the repeatability of the MBNW indices. Healthy non-asthmatic subjects were eligible to participate if they had (1) no history of or medication use relating to chronic respiratory disease; (2) no smoking within the last 6 months and <10 pack-years smoking history; and (3) no respiratory tract infection in the last 4 weeks.

Symptoms
All the subjects with asthma completed the Juniper asthma control questionnaire at the baseline visit and after 3 months of treatment (in the treatment subgroup) to assess asthma symptoms within the previous week.

Exhaled nitric oxide
\( F_{ENO} \) was measured using an offline technique according to American Thoracic Society guidelines. The subject exhaled over 5–15 s into a nitric oxide impermeable polyethylene bag (Scholle Industries Pty Ltd, Elizabeth West, Australia) at a constant flow rate of 0.2 l/s, monitored by rotameter (Dwyer Flowmeter Model VFASS-25, AMBIT Instruments Pty Ltd, Parramatta, Australia). The exhaled gas was analysed using a chemiluminescence analyser (Thermo Environmental Instruments Model 42C, Franklin, Massachusetts, USA). The upper limit of normal for \( F_{ENO} \) has been established as 13 ppb.

Multiple breath nitrogen washout (MBNW)
The MBNW was performed using a closed circuit bag-in-box breathing system to deliver 100% oxygen during inspiration with separate capture of exhaled breath. The nitrogen concentration was measured at the mouth with a Model 721 KacTech Nitrogen Analyser (KacTech Instruments Inc, Green Bay, Wisconsin, USA), while flow and volume were recorded from the box by a pneumotachograph. Subjects breathed 100% oxygen at a tidal volume of 1–1.3 litres until the mean expired nitrogen concentration fell to <2%. Three tests were performed with successive tests starting after alveolar nitrogen had returned to baseline. Ventilation heterogeneity indices in the conductive (S\(_{cond}\)) and acinar (S\(_{acin}\)) lung zones were derived as previously described (see calculations in online supplement available at http://thorax.bmj.com/supplemental). The upper limit of normal for S\(_{cond}\) was 0.037/l and for S\(_{acin}\) was 0.130/l, derived from the mean \( 1.96 \times SD \) of the value obtained previously in normal subjects.

The lung clearance index (LCI) was calculated by dividing the cumulative expired volume (CEV) during the washout by the patient’s functional residual capacity (FRC) determined from the washout. Details of a repeatability study for S\(_{cond}\), S\(_{acin}\) and LCI obtained from an independent group of patients with asthma and healthy non-asthmatic patients are given in the online supplement available at http://thorax.bmj.com/supplemental.

Spirometry and bronchial challenge
Spirometry was performed using a Sensormedics Vmax spirometer (Sensormedics Corporation, Yorba Linda, California, USA) and methacholine challenge tests were performed using the rapid method via hand-held De Vilbiss No 45 nebulisers in which doubling doses ranging from 0.05 \( \mu \)mol to 6.1 \( \mu \)mol were administered until the final dose was reached or the forced expiratory volume in 1 s (FEV\(_1\)) fell by \( >20\% \). Short acting \( \beta\)-agonists were withheld for 6 h and long-acting \( \beta\)-agonists for 24 h before testing. Response to challenge was measured by the dose response ratio (DRR), which is a continuous variable based on the two-point slope. The DRR is calculated from the final step in the challenge test as % fall in FEV\(_1\)/methacholine dose (\( \mu \)mol), and a constant of 3 is added to allow for log transformation of zero or negative values. Greater values of DRR indicate greater AHR, and a DRR value of \( >6.28\% \) fall in FEV\(_1\)/\( \mu \)mol methacholine + 3 indicates the presence of AHR. This allows inclusion of data from subjects who had a fall in FEV\(_1\) of \( <20\% \) either at baseline or following treatment.
Statistical analysis
Data were analysed using the Analyse-it for Microsoft Excel software (Analyse-It Software Ltd, Leeds, UK). DRR and FENO were log-normally distributed and were log10 transformed for all analyses. Change in DRR after treatment was expressed as change in doubling dose of methacholine (change in log DRR/0.3). The relationship between AHR (expressed as DRR) and potential predictive factors was examined using univariate analysis (with Pearson’s correlation coefficient) and multiple linear regression analyses. The sensitivity and specificity of Scond in predicting AHR was assessed by receiver operator characteristics (ROC) analysis. The paired Student’s t test was used for parametric data and the Wilcoxon signed rank test for non-parametric data to compare outcomes after treatment. Data are presented as means with 95% confidence intervals (CI) unless otherwise specified.

RESULTS
Subjects
Forty subjects with asthma participated in the baseline study; 28 had suboptimally controlled asthma and were eligible for the 3-month treatment study and 24 of these agreed to participate. Four subjects from the treatment subgroup withdrew from the study during treatment and the results of two subjects were excluded from the analysis owing to an upper respiratory tract infection at their post-treatment visit. Eighteen subjects were therefore included in the treatment subgroup analysis.

Baseline results of entire asthma group
The characteristics of all 40 patients with asthma at baseline are shown in table 1. AHR was present in 31 of the 40 subjects, and log DRR correlated positively with airway inflammation (log FENO, Scond (fig 1A) and LCI and negatively with % predicted FEV1/FVC ratio (table 2). Log DRR was not correlated with Sacin (p = 0.67). Log FENO was not correlated with Scond (r = 0.29, p = 0.07), Sacin (r = 0.20, p = 0.22) or LCI (r = 0.14, p = 0.40) at baseline.

In a multiple linear regression model which included the significant factors in table 2, log DRR was significantly predicted by both log FENO (partial r² = 0.20, p < 0.001, β coefficient 0.9, 95% CI 0.5 to 1.3) and Scond (partial r² = 0.39, p < 0.001, β coefficient 8.5, 95% CI 4.8 to 12.2). LCI, % predicted FEV1 and FEV1/FVC ratio were not significant predictors of log DRR (p = 0.90, p = 0.25 and p = 0.97, respectively) when included separately or together in this model.

Inhaled corticosteroid treatment subgroup
The characteristics of the 18 subjects with asthma in the treatment subgroup are shown in table 1. Following treatment there were significant improvements in DRR, FENO, Scond, Sacin, LCI, FEV1 (% predicted) and the Juniper symptom score (table 1), but AHR was still present in 10 of the 18 subjects. The univariate correlations at baseline in this subgroup were similar to those in the group as a whole (table 2, fig 1B). However, LCI, % predicted FEV1 and the FEV1/FVC ratio were not correlated with log DRR either before or after treatment in the treatment subgroup (table 2).

In a multiple linear regression model, log DRR in the treatment subgroup was predicted solely by Scond both before (r² = 0.38, p = 0.004, β coefficient 8.3, 95% CI 3.1 to 13.6) and after treatment (r² = 0.64, p < 0.001, β coefficient 10.9, 95% CI 6.8 to 15). Log FENO, LCI, % predicted FEV1 and the FEV1/FVC ratio were not significant predictors of log DRR either before (p = 0.18, p = 0.90, p = 0.24, p = 0.99, respectively) or after treatment (p = 0.38, p = 0.30, p = 0.25, p = 0.42, respectively) in the multivariate regression model.

The magnitude of improvement in AHR correlated with both the reductions in FENO and Scond (F = 3.94, p = 0.04 for the multivariate model), but there was no correlation between the reductions in Scond and FENO2 following treatment (r = 0.07, p = 0.77). There were no differences in the magnitude of improvements in FENO (p = 0.26), Scond (p = 0.81) or DRR (p = 0.90) between subjects treated with HFA-BDP or CFC-BDP (corresponding post-treatment data are represented by squares and triangles in fig 1B).

Scond as a predictor of AHR
Based on the correlation between AHR and Scond, we combined the patients with asthma from this study (n = 40) with a cohort of 17 healthy non-asthmatic subjects (their characteristics are summarised in table E1 in the supplementary data available online at http://thorax.bmj.com/supplemental) to test whether Scond was a significant predictor of the presence of AHR using ROC analysis (fig 2). The area under the ROC curve was 0.88.

Table 1 Characteristics of study subjects

<table>
<thead>
<tr>
<th>Feature</th>
<th>Entire asthma group (n = 40)</th>
<th>ICS treatment asthma subgroup (n = 18)</th>
<th>Absolute difference†</th>
<th>p Value†</th>
</tr>
</thead>
<tbody>
<tr>
<td>M/F</td>
<td>20/20</td>
<td>9/9</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Mean (range) age (years)</td>
<td>32.1 (18–66)</td>
<td>33.9 (18–63)</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Atopic (Y/N)</td>
<td>40/0</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>FENO (ppb)*</td>
<td>15.4 (12.2 to 19.4)</td>
<td>14.1</td>
<td>8.0</td>
<td>6.5 (1.5 to 11.6)</td>
</tr>
<tr>
<td>Scond (/l)</td>
<td>0.055 (0.03)</td>
<td>0.067</td>
<td>0.052</td>
<td>0.02 (0.01 to 0.03)</td>
</tr>
<tr>
<td>Sacin (/l)</td>
<td>0.146 (0.02)</td>
<td>0.159</td>
<td>0.142</td>
<td>0.02 (0.00 to 0.03)</td>
</tr>
<tr>
<td>LCI (CEV/FRC)</td>
<td>8.16 (1.1)</td>
<td>8.54</td>
<td>7.78</td>
<td>0.8 (0.4 to 1.1)</td>
</tr>
<tr>
<td>DRR (% fall FEV1/µmol methacholine+3)*</td>
<td>18.9 (12.4 to 28.8)</td>
<td>22.0</td>
<td>10.1</td>
<td>1.1 (0.6 to 1.6)</td>
</tr>
<tr>
<td>FEV1 (% predicted)</td>
<td>82.4 (11.7)</td>
<td>80.1</td>
<td>85.9</td>
<td>5.8 (3.2 to 8.4)</td>
</tr>
<tr>
<td>FEV1/FVC ratio</td>
<td>0.76 (0.08)</td>
<td>0.73</td>
<td>0.75</td>
<td>0.02 (0.00 to 0.04)</td>
</tr>
<tr>
<td>β agonist use (times/day)</td>
<td>1.4 (2.4)</td>
<td>1.95</td>
<td>0.54</td>
<td>0.8 (–0.0 to 1.6)</td>
</tr>
<tr>
<td>BDP equivalent dose (µg/day)</td>
<td>320 (449.6)</td>
<td>270</td>
<td>1500</td>
<td>–</td>
</tr>
<tr>
<td>Juniper symptom score</td>
<td>1.28 (0.84)</td>
<td>1.48</td>
<td>0.81</td>
<td>0.7 (0.4 to 0.9)</td>
</tr>
</tbody>
</table>

Values are mean (SD) unless otherwise stated.
*Geometric mean and 95% CI.
†Comparison between values before and after treatment.
‡Doubling dose difference in DRR.
BDP, beclomethasone dipropionate; CEV, cumulative expired volume; DRR, dose response ratio; FEV1, forced expiratory volume in 1 s; FENO, fraction of nitric oxide in exhaled breath; FRC, functional residual capacity; FVC, forced vital capacity; ICS, inhaled corticosteroid; LCI, lung clearance index; MBNW, multiple breath nitrogen washout; Scond, ventilation heterogeneity in the conducting airways; Sacin, ventilation heterogeneity in acinar lung zone; ppb, parts per billion.
Ventilation heterogeneity and airway hyperresponsiveness in asthma

(p<0.0001), and the cut-off value of $S_{\text{cond}} = 0.037/l$ corresponding to the upper limit of normal range gave a good combination of sensitivity and specificity (71.9% and 90.0%, respectively) for detecting the presence of AHR. Using the same cut-off value ($0.037/l$), $S_{\text{cond}}$ remained a significant predictor of the presence of AHR after treatment in our 18 subjects from the subgroup (area under the ROC curve = 0.96, p<0.0001) with a high sensitivity and specificity (90.0% and 87.5%, respectively, see fig E2 in the supplementary data available online at http://thorax.bmj.com/supplemental).

Repeatability of $S_{\text{cond}}$, $S_{\text{acin}}$ and LCI

The characteristics of the cohort of 10 subjects with asthma and 11 healthy non-asthmatic subjects who participated in the repeatability study of the MBNW indices are summarised in table E2 in the supplementary data (available online at http://thorax.bmj.com/supplemental). In subjects with asthma, $S_{\text{cond}}$, $S_{\text{acin}}$ and LCI had a good repeatability with an intraclass correlation coefficient of 0.84 (95% limits of agreement $-0.026$) for $S_{\text{cond}}$, 0.95 (95% limits of agreement $+0.027$) for $S_{\text{acin}}$, and 0.91 (95% limits of agreement $+0.735$) for LCI. The repeatability results for the healthy non-asthmatic subjects are shown in table E3 in the supplementary data (available online at http://thorax.bmj.com/supplemental).

Table 2 Pearson correlation coefficients (r) of baseline variables with airway hyperresponsiveness (dose response ratio, DRR) for the asthmatic group as a whole and for the treatment subgroup before and after treatment

<table>
<thead>
<tr>
<th>Variable</th>
<th>Baseline asthma group (n = 40)</th>
<th>Baseline treatment subgroup (n = 18)</th>
<th>Post-treatment treatment subgroup (n = 18)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>r</td>
<td>p Value</td>
<td>r</td>
</tr>
<tr>
<td>Airway inflammation (FENO)</td>
<td>0.62</td>
<td>&lt;0.001</td>
<td>0.51</td>
</tr>
<tr>
<td>Ventilation heterogeneity ($S_{\text{cond}}$)</td>
<td>0.62</td>
<td>&lt;0.001</td>
<td>0.64</td>
</tr>
<tr>
<td>Ventilation heterogeneity ($S_{\text{acin}}$)</td>
<td>0.07</td>
<td>0.67</td>
<td>0.04</td>
</tr>
<tr>
<td>Ventilation heterogeneity (LCI)</td>
<td>0.38</td>
<td>0.02</td>
<td>0.31</td>
</tr>
<tr>
<td>FEV$_1$, % predicted</td>
<td>-0.44</td>
<td>0.003</td>
<td>-0.40</td>
</tr>
<tr>
<td>FEV$_1$/FVC ratio</td>
<td>-0.47</td>
<td>0.002</td>
<td>-0.34</td>
</tr>
</tbody>
</table>

FEV$_1$, forced expiratory volume in 1 s; FNO, fraction of nitric oxide in exhaled breath; FVC, forced vital capacity; LCI, lung clearance index; $S_{\text{cond}}$, ventilation heterogeneity in conducting airways; $S_{\text{acin}}$, ventilation heterogeneity in acinar lung zone
DISCUSSION

In this study we have shown that baseline ventilation heterogeneity correlates strongly with the severity of AHR in subjects with asthma with a range of disease severity. In particular, $S_{\text{cond}}$ predicted AHR to methacholine independently of airway inflammation and airway calibre. Importantly, the strong correlation between $S_{\text{cond}}$ and AHR persisted after airway inflammation was reduced with ICS treatment. Following treatment, $S_{\text{cond}}$ was, in fact, the sole independent predictor of AHR. The improvement in $S_{\text{cond}}$ with ICS treatment suggests that it is determined, at least in part, by steroid-responsive inflammatory processes. However, the residual ventilation heterogeneity which persisted after ICS treatment suggests that non-steroid responsive or non-inflammatory factors also make an important contribution to ventilation heterogeneity. The conductive airways, where the residual ventilation heterogeneity responsible for AHR after anti-inflammatory treatment originates, clearly constitute an important therapeutic target. These findings are novel and have clinical implications in patients with asthma.

Ventilation heterogeneity in the lungs is a well recognised feature of asthma, but its clinical significance—apart from the deleterious effects on gas exchange—has been uncertain. In our study, the main contribution to ventilation heterogeneity that was associated with AHR originated in the conductive airways. Gustafsson et al. reported a correlation between airway responsiveness to cold air challenge and ventilation heterogeneity measured by single breath washout using helium and SF6, which indicated an involvement of peripheral airways but did not assess the contribution of airway inflammation. It is impossible independently to determine ventilation heterogeneity of conductive airways from the single breath washout, making comparison with the present study difficult. However, despite methodological differences in the determination of ventilation heterogeneity and AHR, the observed correlations between ventilation heterogeneity and AHR in both studies suggest a robust association. In the present study, AHR was not correlated with baseline acinar ($S_{\text{acin}}$) ventilation heterogeneity, signalling that the important lung region with regard to the relationship between ventilation heterogeneity and AHR is specifically in the conducting airways. The weak correlation between the LCI and AHR indicates that this measure of specific ventilation distribution between relatively large lung units, which also partly contributes to $S_{\text{cond}}$, is neither sensitive nor specific as a measure of the conductive airway heterogeneity underlying AHR.

The significant reduction in ventilation heterogeneity in our subjects following anti-inflammatory ICS treatment suggests that ventilation heterogeneity is partly due to steroid-responsive airway inflammation. After treatment, residual ventilation heterogeneity persisted in the presence of normalised mean exhaled nitric oxide levels, suggesting that other inflammatory processes not related to exhaled nitric oxide, or non-inflammatory processes, could also contribute to ventilation heterogeneity. Exhaled nitric oxide correlates with sputum eosinophils, but it does not reflect the full spectrum of inflammatory and non-inflammatory processes associated with asthma. This may explain the lack of association between improvement in exhaled nitric oxide and ventilation heterogeneity during treatment. Ventilation heterogeneity could also continue to improve with further ICS treatment, as has been shown to occur with AHR which can continue to improve up to and after 12 months of treatment. It would be valuable in future studies also to assess any additional benefits of systemic treatments on ventilation heterogeneity.

The residual ventilation heterogeneity present at the end of treatment may also be caused by non-inflammatory structural changes such as airway remodelling or increased smooth muscle tone. Structural changes induced by airway remodelling are likely to be heterogeneously distributed among parallel pathways of the bronchial tree. These structural changes manifest as altered composition and organisation of the soft tissues of the airway walls and are believed to be a result of chronic or repeated episodes of acute inflammation. The changes include thickening of the airway wall, smooth muscle hyperplasia and hypertrophy, subepithelial fibrosis and mucous metaplasia. It is possible that many, if not all, of these abnormalities may contribute to ventilation heterogeneity by exaggerating the inherent heterogeneity of the airway tree and by being unevenly distributed themselves.

Increased baseline ventilation heterogeneity may cause a heterogeneous distribution of methacholine aerosol resulting in high concentrations of methacholine delivered to a small surface area of the airways. In a previous study, central-to-peripheral airway differences in the pattern of methacholine deposition in subjects with asthma did not alter AHR. However, a recent imaging study in sheep showed that the regions of lung with the highest baseline ventilation were the same regions which became constricted after methacholine, suggesting a link between parallel ventilation heterogeneity and methacholine deposition. This could partly explain the association between $S_{\text{cond}}$ (representative of parallel ventilation heterogeneity) and AHR. The exact mechanism by which ventilation heterogeneity may cause AHR remains unclear. However, a recent modelling study showed that only minor alterations in structure, function or smooth muscle activity in a virtually uniform airway tree are required to trigger extreme ventilation heterogeneity during uniformly applied bronchoconstriction.

One of the strengths of our study was the wide range of asthma severity in the group of subjects tested at baseline. Indices of ventilation heterogeneity from the MBNW were robust, repeatable and comparable with values obtained in other subjects with asthma. The strong predictive capacity of $S_{\text{cond}}$ for the presence of AHR, as inferred from our ROC analysis (fig 2), is reinforced because the upper limit of the normal range for $S_{\text{cond}}$ (0.037/l) corresponded to a high combination of sensitivity and specificity for detecting the presence of AHR in the present study. This raises the possibility that $S_{\text{cond}}$ could at least partly be used to predict AHR as part of the clinical management of asthma. MBNW tests can be performed in patients whose lung function is too low to permit bronchial challenge and has been successfully administered in non-sedated children with a number of airway diseases.

In summary, we have shown for the first time that ventilation heterogeneity in the conducting airways is a significant predictor of AHR in subjects with asthma, independent of airway inflammation as measured by exhaled nitric oxide, both before and after ICS treatment. Inflammatory and non-inflammatory processes which could alter the structural features within the airway tree may contribute to ventilation heterogeneity. Based on the strength of the correlations in this study and on the results of computational modelling, it is possible that ventilation heterogeneity is an important contributor to AHR in asthma. The index $S_{\text{cond}}$ derived from the MBNW is a measure of conducting airway function that is likely to be useful in the clinical management of asthma and for the assessment of new treatment strategies and drugs in this disease. These findings suggest that greater understanding of the determinants of ventilation heterogeneity may lead to more efficacious asthma treatment and can provide an additional tool for monitoring treatment, resulting in improved long-term outcomes for patients with asthma.
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Additional data are available in the online supplement at http://thorax.bmj.com/supplemental.

Authors’ affiliations
Sue R Downie, Cheryl M Salome, Norbert Berend, Gregory G King, Woolcock Institute of Medical Research, Camperdown, Sydney, Australia
Sylvia Verbanck, Respiratory Division, Academic Hospital, Vrije Universiteit Brussels, 1090 Brussels, Belgium
Bruce Thompson, Department of Allergy, Immunology, and Respiratory Medicine, The Alfred Hospital and Monash University, Melbourne, Australia
Norbert Berend, Gregory G King, Department of Respiratory Medicine, Royal North Shore Hospital, Sydney, Australia

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Competing interests: The authors have the following potential conflicts of interest to declare: Cheryl M Salome states that her research group has received travel sponsorship from Boehringer Ingelheim, GlaxoSmithKline and AstraZeneca but these do not pose any conflict of interest for this paper. Norbert Berend serves on advisory boards for GlaxoSmithKline, Boehringer Ingelheim and AstraZeneca, and The Woolcock Institute of Medical Research of which he is the Director receives research grants from GlaxoSmithKline. Gregory King has received travel sponsorship from GlaxoSmithKline to attend the ATS ASM 2003 (approximately $9000) and a GlaxoSmithKline meeting in 2003 (approximately $11000), travel sponsorship from AstraZeneca to attend ATS ASM 2004 (approximately $1000) and two AstraZeneca scientific meetings (approximate combined value $2000), an honorarium paid to his research institute from GlaxoSmithKline to speak at a sponsored conference in South East Asia ($3000) in 2005, a travel grant from AstraZeneca to attend the ATS 2005 ($46000) and a travel grant from GlaxoSmithKline to attend the ATS 2006 ($45000). A proportion of Dr King’s research work is conducted at the Woolcock Institute of Medical Research which receives unrestricted grants from AstraZeneca, GlaxoSmithKline, Boehringer Ingelheim and Vita Medical for such work. The Woolcock Institute of Medical Research also has a consultancy agreement with Pfizer, Boehringer Ingelheim, AstraZeneca and GlaxoSmithKline for which Dr King provides consultancy services related to asthma and COPD. His research group receives a proportion of the salary of the respiratory physicians from those companies as part of specific and general allocations of those funds for research purposes across all research groups of the Woolcock Institute of Medical Research. Dr King does not own any stocks, equity or patents that pose a conflict of interest.

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