Inflammatory cells in the airways in COPD

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Airway inflammation is central to the pathogenesis of both airway remodelling and parenchymal destruction in chronic obstructive pulmonary disease (COPD). Neutrophils, macrophages, and CD8+ T lymphocytes have been implicated in a number of studies, but a detailed profile of disease-phenotype specific inflammation has yet to emerge. The heterogeneity of the disease has hindered data interpretation while extrapolation of the results of relatively non-invasive studies to the actual pathology found in the distal lung is difficult. Moreover, prominent studies have had frequently conflicting results. Further investigations are needed to marry the different clinical phenotypes of COPD to their respective inflammatory profiles in the airways and thus improve our understanding of the pathogenesis of the disease as a whole.

Asthma has been the dominant focus of airways research interest, at least until recent times. A multitude of studies have examined its tapestry of airway inflammation and remodelling. Sophisticated theories have resulted, encompassing environmental exposures, epithelial repair, and epithelial-mesenchymal signalling.1 Therapeutic successes have followed, encouraging more interest and further investment. Chronic obstructive airways disease (COPD), on the other hand, has suffered from the unflattering contrast. COPD is generally considered a self-inflicted condition that results, in the majority, from years of smoking and the only effective treatment to date is long term oxygen therapy. Its profile of airways inflammation is, in retrospect, unsurprising. Neutrophils, macrophages, and CD8+ T lymphocytes are, after all, predictable responders to a sustained noxious insult. However, in recent years something of a renaissance has occurred. COPD inflammation research has burgeoned, new treatments are being tested,2 and interests are focused not now so much on the cell types but more on the “whys and wherefores” of their actions in COPD.

Historically, the link between airways inflammation and lung pathology in smokers had long been suspected. In the 1950s a post-mortem study of centrifugal emphysema revealed “bronchiolitis” of the airways leading into emphysematous parenchyma.3 Further studies confirmed not only that peripheral airways inflammation is associated with remodelling and destruction,4,5 but also that its severity increases in parallel with emphysema severity.6–8 They also revealed that bronchiolar inflammation and emphysema share the same anatomical distribution,6,7 that small airways disease temporally precedes emphysema,8 and that bronchiolar inflammation severity correlates with functional impairment.9–12 It was therefore hypothesised that centrifugal emphysema results from an inflammatory reaction that spreads centrifugally from the bronchioles to involve the parenchyma.9

Animal model studies and in vitro experiments have explored the roles of specific inflammatory cells in COPD, while bronchoscopy, sputum induction and lung resection studies have described airways inflammation in vivo. All have helped to confirm dominant positions in the inflammatory hierarchy for neutrophils,13–15 macrophages,16–18 and CD8+ T lymphocytes.19–21 Other inflammatory cells such as mast cells,22 eosinophils,22 and natural killer cells23 have been credited with less importance.

KEY INFLAMMATORY CELLS IN COPD

Neutrophils
Neutrophils are front line defensive cells of the immune system and a source of reactive oxygen metabolites, inflammatory cytokines, lipid mediators, antibacterial peptides, and tissue damaging enzymes.24–27 They are strongly implicated in both the generation of mucous metaplasia in chronic bronchitis and the destruction of lung tissue in emphysema.

Neutrophil products induce mucus hypersecretion by both an acute secretagogue effect and by augmentation of the bronchial mucus producing apparatus.28–30 Mucin gene expression has been proposed as the principal factor governing the differentiation of epithelial cells into goblet cells.30 Neutrophil elastase (NE) and reactive oxygen species independently increase epithelial mucin mRNA and protein expression in vitro,31–33 possibly via ligand independent transactivation of the cell surface epidermal growth factor receptor (EGFR).34 In asthma the epithelial mucin, MUC5AC, and the EGFR are co-localised in airway epithelial goblet cells.35 As severe asthma is associated with both massively increased mucus production and neutrophilic airway inflammation,36–38 it has been hypothesised that neutrophil driven goblet cell metaplasia is a component of asthma airways remodelling. In COPD, MUC5AC is the principal secreted mucin of the epithelium37 and, like

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Received 12 March 2004
Accepted 22 November 2005

Abbreviations: BAL, bronchoalveolar lavage; COPD, chronic obstructive pulmonary disease; ECP, eosinophilic cationic protein; EGFR, epidermal growth factor receptor; FEV1, forced expiratory volume in 1 second; IFN-γ, interferon γ; IL, interleukin; LT, leukotriene B4; MMP, matrix metalloproteinase; MPO, myeloperoxidase; NFκB, nuclear factor κB; nFGF, neutrophil elastase; NK, natural killer; TNF-α, tumour necrosis factor α

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Demonstrated NE in emphysematous tissue, increased protease/antiprotease imbalance in the lung. To support emerged that the neutrophil was the perpetrator of a deficiency was first linked with the disease. The theory patients with COPD, and a correlation between emphysema products of elastase activity in urine and plasma from this, animal model studies have shown that administration of severity and elastase levels in peripheral blood neutrophils. Figure 1 demonstrates epithelial MUC5AC immunohistochemical staining in a bronchial biopsy specimen taken from a smoker with COPD, indicating its clear expression.

The neutrophil's standing as a potential key cell in emphysema has been assured since inherited α1-antitrypsin deficiency was first linked with the disease. The theory emerged that the neutrophil was the perpetrator of a protease/antiprotease imbalance in the lung. To support this, animal model studies have shown that administration of purified NE produces emphysema, while deficiency of endogenous NE affords protection against emphysema following exposure to cigarette smoke. Human studies have demonstrated NE in emphysematous tissue, increased products of elastase activity in urine and plasma from patients with COPD, and a correlation between emphysema severity and elastase levels in peripheral blood neutrophils. In vitro, peripheral blood neutrophils from subjects with severe asthma and other hypersecretive diseases such as bronchiectasis and cystic fibrosis, neutrophilic airways inflammation is a key disease feature. Figure 1 demonstrates epithelial MUC5AC immunohistochemical staining in a bronchial biopsy specimen.

Macrophages account for the majority of inflammatory cells recovered by airway lavage, regardless of whether or not the subjects are non-smokers, healthy smokers or smokers with airways disease. Compounds released by macrophages include reactive oxygen species, chemotactic factors, inflammatory cytokines, smooth muscle constrictors, mucus gland activators, and extracellular matrix proteins. Also included is an array of matrix metalloproteinase enzymes (MMPs). These can, when combined, degrade a similar spectrum of proteins to neutrophil enzymes and they are also believed to facilitate leucocyte migration and infiltration into injured tissues. Although macrophages do not transcribe the NE gene, their ability to internalise the enzyme has led to the proposal that macrophage released NE can add further to the proteolytic potential of the cell.

Although macrophages have the theoretical potential to induce mucus hypersecretion via products with secretagogue activity such as leukotriene B4 (LTB4) and interleukin 1 (IL-1), most studies have focused on their possible role in emphysema, particularly via MMP production. In animal models administration of aerosolised alveolar macrophages produces emphysema, as does overexpression of MMP-1. Use of a broad spectrum MMP inhibitor attenuates both the initial inflammatory response and the severity of emphysema in guinea pigs exposed to cigarette smoke. In vitro, cultured alveolar macrophages taken from subjects with COPD express increased amounts of both MMP-1 and MMP-9 while, in vivo, immunoreactivity for MMP-2 and MMP-9 is demonstrably increased in COPD.

MMP-12, in particular, has been a focus of increasing attention in emphysema. In mice, deficiency of MMP-12 appears to be protective against cigarette smoke induced lung destruction. In vivo, airway and alveolar macrophage MMP-12 expression in COPD subjects is enhanced compared with healthy controls. Recent evidence, moreover, suggests that the role of MMP-12 may go beyond matrix degradation. In mouse models its presence is required for cigarette smoke induced neutrophil influx into the lung, possibly via release of the pro-inflammatory cytokine tumour necrosis...
factor α (TNF-α) with subsequent upregulation of vascular adhesion molecule VCAM-1. This supports the view that neutrophils and macrophages are co-dependent on one another to produce an elastolytic inflammatory response in airways exposed to cigarette smoke.

Studies of emphysematous lung tissue from human subjects have shown a direct relationship between alveolar macrophage density in the parenchyma and the severity of lung destruction. Moreover, numerous investigations have found macrophage numbers to be increased in the bronchial submucosa, bronchial glands, and small airways epithelium of subjects with COPD. Macrophage counts in induced sputum or bronchoalveolar lavage (BAL) fluid in COPD vary depending on whether relative or absolute cell counts are used. Relative macrophage counts are often reduced, a reflection, most likely, of increased neutrophil percentages. Absolute counts, on the other hand, are increased both in sputum and BAL fluid.

**T lymphocytes**

In asthma the CD4+ T cell is the proposed orchestrator of a Th2 type immune response in the airways. In COPD the CD8+ cell is the accepted crucial lymphocyte subtype. An increase in total T cells (CD3+ cells), which includes both CD4+ and CD8+ cells, occurs in the alveolar walls in emphysema, but CD8+ cells are predominant. Increased CD8+ cell numbers are also found in both the large and small airway walls in COPD and in peripheral airway smooth muscle. Figure 3 shows CD8+ cells infiltrating the bronchial submucosa and epithelium in a bronchial biopsy specimen.

Speculation has surrounded the role of CD8+ cells in emphysema. A key function of the CD8+ cell is to combat viruses either by cytolytic of infected cells or induction of apoptosis. Collateral tissue damage is a possible consequence; in respiratory syncytial virus (RSV) infected mice, for example, excessive CD8+ cell activation results in potentially lethal pulmonary damage. In smokers infected with human immunodeficiency virus (HIV), high CD8+ cell numbers in BAL fluid have been associated with an accelerated onset of emphysema.

If CD8+ cells destroy lung parenchyma, the mechanism is uncertain. They have, in theory, the potential to damage the lung interstitium directly via release of lytic substances such as perforin and granzyme. It was observed recently that CD8+ cells recovered from the sputum of COPD patients display higher levels of perforin expression and increased cytotoxic activity than CD8+ cells taken from control subjects. Another possibility is that CD8+ cells induce structural cell apoptosis. Alveolar cell turnover does appear to increase in emphysema as increased numbers of alveolar epithelial and endothelial cells undergo both proliferation and apoptosis. Moreover, an association has been observed between apoptosing cell numbers and CD8+ T cell numbers in the alveolar walls. The possibility remains, however, that such associations are a secondary phenomenon. Given their potential suppressor function, the infiltrating CD8+ cells may actually serve to inhibit the inflammatory process. Further studies are needed to define the activity of infiltrating CD8+ cells in emphysema.

Prior activation of T cells by antigen presentation is generally required before they can infiltrate non-lymphoid tissue such as the lung. Although the nature of the proposed antigen in COPD is unknown, two hypotheses have been put forward. Enelow and colleagues demonstrated that recognition of a lung “autoantigen” by T cells can produce lung injury in the absence of a viral stimulus, and this damage was mediated or amplified by non-antigen specific inflammatory cells such as macrophages. It was therefore hypothesised that repetitive injury to the lung, as a result of chronic smoking and inflammation, results in structural alteration of self-antigens allowing them to be recognised by T cells.

Alternatively, a persistent intracellular pathogen may provide a foreign antigenic stimulus. One such candidate is adenovirus, as suggested by Retamales and colleagues who found a 41-fold increase in alveolar epithelial cells expressing the adenoviral transactivating protein EIA in subjects with severe emphysema compared with healthy smokers.

A number of molecules have been proposed to mediate pulmonary T cell infiltration in COPD. RANTES (regulated upon activation, normal T cell expressed and secreted), a potent chemoattractant that is overexpressed in exacerbations of chronic bronchitis, is believed to act synergistically with CD8+ cytolytic cells to enhance apoptosis of virally infected cells. It was therefore postulated that, when CD8+ cells predominate, recurrent viral infections with resulting increased RANTES levels may promote CD8+ cell mediated tissue damage. Th1 type cycle of inflammation has also been suggested; in COPD airways infiltrating CD8+ cells express more of the chemokine receptor CXCR3, its ligand CXCL10, and the inflammatory cytokine interferon γ (IFN-γ). This has led to the suggestion that an inflammatory insult induces increased airway expression of CXCL10 and IFN-γ, that these—via expression of IFN-γ—could induce further CXCL10 expression thus perpetuating a cycle of CD8+ inflammation.

The function of CD4+ cells in COPD is unknown. CD4+ cells, in addition to CD8+ cells, are increased in the small airway walls of smokers with severe COPD. They have the potential to contribute to the inflammatory process via production of a variety of pro-inflammatory cytokines including TNF-α and granulocyte-macrophage colony-stimulating factor (GM-CSF), the Th1 agents IFN-γ and IL-2, and the Th2 agents IL-4, IL-5, and IL-6. Th2 type cycle of inflammation, results in structural alteration of self-antigens allowing them to be recognised by T cells.

**Natural killer (NK) T lymphocytes**

NK cells are a distinct population of specialised cytoytic lymphocytes that target transformed or virus infected cells. Although the function of NK cells in COPD has not been widely studied, one investigation suggested that...
they are found in increased numbers in the large airway submucosa of smokers with COPD, with the hypothesis that excessive NK cell recruitment in COPD occurs due to repeated viral or bacterial infections. However, a study measuring lymphocyte subpopulations in the alveolar walls of non-smokers, healthy smokers, and smokers with emphysema found no increase in NK cell numbers in subjects with emphysema.13

Eosinophils and mast cells

Although eosinophils and mast cells are important effector cells in asthma, neither has been ascribed a prominent role in COPD. In exacerbations of COPD, airway wall eosinophilia has been observed at a level similar to that found in asthma.46 47 with an associated increase in RANTES expression by airway subepithelial and epithelial cells.46 In stable COPD, increased eosinophil counts have been found in sputum,74 BAL fluid,41 and in the airway wall.22 54 95 Increased levels of eosinophil cationic protein (ECP) in BAL fluid46 and induced sputum47 96 have also been observed. However, some studies have failed to confirm that any airway eosinophilia occurs in COPD,13 17-19 22 99 while others have suggested that the eosinophils, even if they are present, are not activated.40 41 The mechanism underlying eosinophilic infiltration seems to differ from asthma in that the key chemokines IL-4, IL-5 and eotaxin do not appear to be overexpressed in COPD airways.43 It has been hypothesised that any influx of these cells is merely a by-product of smoking induced inflammation. A possible mediator is the chemokine IL-8. Although normally associated with neutrophil chemotaxis, IL-8 exerts a chemotactic effect on primed eosinophils100 101 and correlates with BAL fluid levels of ECP in chronic bronchitis.12

Some reports have shown that increased numbers of eosinophils in sputum74 102 103 and BAL fluid104 taken from COPD subjects are predictive of a clinical response to steroid treatment. These results might suggest that eosinophils play a role in a clinical subset of COPD that has inflammatory features in common with that of asthma. However, the two diseases remain, in general, phenotypically dissimilar as even subjects with asthma who have fixed airflow obstruction have significantly higher percentages of eosinophils and lower percentages of neutrophils in both sputum and BAL fluid than COPD patients with similar disease severity.105

The ability of mast cells to release an array of mediators has invited speculation regarding their possible role in COPD inflammation. This could include neutrophil recruitment via the release of chemotactic factors, tissue injury by the actions of the secreted enzymes tryptase, chymase and elastase, and mucus hypersecretion via the potent secretagogue action of mast cell chymase.106 Some studies of COPD airways have revealed increased mast cell numbers in the airway wall.107 108 In addition, one study which found that mast cell degranulation in the bronchial gland layer appeared more marked in subjects with chronic bronchitis than in healthy controls suggested that increased mast cell activity may also be present.108 These findings are countered, however, by reports displaying no evidence of mast cell abundance in either the airways or parenchyma in COPD subjects.56 57 73 94 95

OBSTACLES TO RESEARCH INTO COPD INFLAMMATION

A detailed profile of disease phenotype specific inflammation has yet to emerge in COPD. There are many reasons for this, but chief among them may be the very nature of the condition itself. COPD is a heterogeneous disease, so the pathological significance of inflammatory cell measurements is often difficult to divine. The clinical severity varies considerably, asymptomatic disease is common,109 110 and interobserver variability exists in the detection of physical signs allowing mild cases to go undetected.111 Moreover, none of the common clinical tests give a complete picture of the disease. Spirometric tests are inadequate for the detection of early COPD,112 113 airflow obstruction severity is a poor predictor of symptom severity,114 and impairment of carbon dioxide gas transfer is not specific to emphysema and may occur by separate mechanisms in smokers.115-119

Linkage of lung function readings to the underlying pathology is hampered by the fact that both peripheral airway remodelling120 121 and emphysema17 122 result in airflow obstruction while emphysema is commonly found in smokers who have not developed airflow obstruction at all.123 Thus, characterisation of research subjects based only on clinical impression and lung function—as was the case with many studies18 20 95—fails to provide a clear insight into the underlying pathology. Neutrophil studies provide a prime example of the importance of distinguishing COPD phenotypes. Sputum neutrophil counts correlate with the severity of airflow obstruction in COPD,15 19 and smokers with severe emphysema have neutrophilic inflammation in the alveolar walls and air spaces.47 However, in mild emphysema there is neither BAL nor alveolar neutrophilia, even though BAL fluid levels of neutrophil degranulation products and the neutrophil chemokine IL-8 are increased.49 50 Mild emphysema would often go undetected on lung function testing so, without the use of CT scanning, such patients would be classified as healthy smokers.

A further obstacle is that the most practical investigative techniques have natural limitations. The predominant pathology is found in the not easily accessible peripheral airways and lung parenchyma. To what extent proximal airway biopsies represent events in the distal lung is uncertain. Although BAL and induced sputum sample more distally, the correlation between luminal and tissue inflammation is also unknown. Moreover, the results depend on the protocol in use. Inflammatory cell numbers vary depending on the lung region sampled, with neutrophils more numerous in the proximal bronchial tree and macrophages predominant more distally. Digital subtraction angiography has shown that BLF (fluid aspirated back from the initially instilled aliquots) samples proximal airways, whereas successive aliquots (BAL fluid) may reflect distal airway and alveolar events.124 Thus, in healthy smokers neutrophil percentages decline in successive BAL fluid aliquots.125 Sputum induction, similarly, is prone to sampling effects. In subjects with COPD, induced sputum contains higher percentages of neutrophils and lower percentages of macrophages than BAL fluid, most probably because the more proximal airways are sampled.14 126 However, increasing the duration of the induction results in a progressive reversal of these proportions, possibly due to increasingly peripheral airways sampling.127 Standardisation of induction duration is therefore important if reliable comparisons are to be made between different centres.

Prominent studies of airway inflammation in COPD have had conflicting results. Some investigations which showed CDS+ T lymphocyte infiltration of the airways128 20 have not been confirmed by others.14 15 Macrophage infiltration has similarly been both confirmed129 130 and refuted,120 while studies measuring neutrophils, eosinophils, and mast cells in airway wall and sputum have also produced inconsistent results.

The potentially crucial impact of infection and exacerbations on airways inflammation and disease severity remains unquantified. Frequent exacerbations of COPD are associated with a faster decline in forced expiratory volume in 1 second (FEV1) over time.128 Exacerbations are also associated with raised sputum counts of lymphocytes, neutrophils, and...
In the airway wall, similarly, both neutrophilia and eosinophilia are observed, as is overexpression of related chemokines IL-8, CXCL5 and RANTES.15 16 94 Bacterial infections, specifically, have been linked to increases in both neutrophil activation markers and associated inflammatory mediators including myeloperoxidase (MPO), NE, LTβ4 and IL-8.110 111 Even in the absence of acute infection, airway bacterial colonisation is well recognised in COPD.112 Such colonisation has been correlated with airway neutrophil activation,113 sputum levels of the pro-inflammatory cytokines IL-8, LTβ4 and TNF-α,114 115 airflow obstruction severity,116 and a more rapid decline in FEV1 over time in smokers.115 It has been hypothesised that chronic lower airway bacterial colonisation may facilitate the development of airflow limitation via both increased airflow in stable disease and by predisposing to more frequent exacerbations.113 117 118 Although most studies relating inflammation to clinical disease severity have excluded acute infection from their subjects on clinical grounds, airway colonisation has not been ruled out to the same extent. The effects of current smoking remain uncertain. Most investigators have preferred to record the pack year history instead. Rennard and colleagues showed that short term smoking reduction is associated with a reduction in airflow inflammation in heavy smokers.119 Willens and colleagues found a positive association between current smoking and macrophage numbers in both sputum and bronchial biopsy specimens.120 Other investigators have shown, however, that current and ex-smokers with COPD have similar degrees of inflammation in bronchial biopsy121 and induced sputum122 samples. Thus, although intuitively one would expect the ongoing insult to the airways to be relevant, the jury remains out as to whether differences in current smoking contribute to the variability of results between different studies. Thus, to date, diverse methods and, often, diverse results, have contributed to uncertainty surrounding the inflammatory profile of COPD. If future anti-inflammatory strategies are to be tested, then measurements of inflammation before and after treatment in bronchoscopic or sputum samples may be necessary. More clarity is needed as to the strength of the relationships between proximal airway inflammation, distal airway inflammation, and lung pathology.

CONCLUSION
That airway inflammation is central to the pathogenesis of COPD appears beyond doubt. However, studies to date have varied widely in terms of the depth of disease characterisation, their allowance for confounding factors such as current smoking and infection, and the investigational methodologies employed. Future studies need to clarify the relationship between each specific inflammatory cell type and each of the remodelling and destructive processes found in COPD airways. Further research combining well validated techniques—specifically, comparison of preoperative endobronchial biopsy samples with examination of resected lung specimens and careful patient phenotyping—may help.

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Funding: none.

Competing interests: none.

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