Adding salmeterol to an inhaled corticosteroid: long term effects on bronchial inflammation in asthma

J G Koopmans, R Lutter, H M Jansen, J S van der Zee

Background: Addition of the long acting β2 agonist salmeterol to inhaled corticosteroids leads to better symptomatic asthma control than increasing the dose of inhaled corticosteroids. However, little is known about the long term effects of adding salmeterol on the asthmatic inflammatory process, control of which is considered important for the long term outcome of asthma.

Methods: After a 4 week fluticasone run-in period, 54 patients with allergic asthma were randomised to receive twice daily treatment with fluticasone 250 μg with or without salmeterol 50 μg for 1 year in a double blind, parallel group design (total daily dose of fluticasone 500 μg in both treatment groups). Primary outcomes were sputum eosinophil numbers and eosinophil cationic protein concentrations. Secondary outcomes were neutrophil associated sputum parameters and a respiratory membrane permeability marker. The effects on allergen induced changes were determined before and at the end of the treatment period.

Results: Adding salmeterol to fluticasone resulted in improved peak expiratory flow, symptom scores, rescue medication usage, and bronchial hyperresponsiveness (p < 0.05 for all). There was no sustained effect on sputum cell differential counts and cytokine concentrations during the treatment period or on changes induced by allergen challenge at the end of treatment (p > 0.05). However, adding salmeterol significantly reduced sputum ratios of α2-macroglobulin and albumin during the treatment period (p = 0.001).

Conclusions: The addition of salmeterol to fluticasone produces no sustained effect on allergen induced cellular bronchial inflammation but leads to a significant improvement in size selectivity of plasma protein permeation across the respiratory membrane. This may contribute to the improved clinical outcome seen in patients with allergic asthma when a long acting β2 agonist is combined with inhaled corticosteroids.
mation of maintenance treatment with fluticasone/salmeterol versus fluticasone. Primary outcomes were sputum eosinophil numbers and eosinophil cationic protein (ECP) concentrations. Secondary outcomes were serum levels of interleukin (IL)-5, size selectivity of the respiratory membrane as measured by the ratio of sputum α2-macroglobulin to albumin, sputum neutrophil numbers, IL-8, and myeloperoxidase (MPO). In addition, we measured lung function parameters, peak expiratory flows, symptom scores, and rescue medication usage. The overall differences in outcome parameters over the 1 year treatment period were analysed, as well as differences in allergen induced changes between the two treatment groups.

METHODS

An extended version of the Methods section is available in the online data supplement at http://www.thoraxjnl.com/supplemental.

Study participants

Non-smoking patients with mild to moderate persistent allergic asthma were enrolled. The participating patients were recruited via advertisements and from the Outpatient Department of Pulmonology at the Academic Medical Center (AMC). The study was approved by the institutional ethics and research committees and all subjects gave written informed consent.

Study design

The study was of a double blind, randomised, two armed parallel design (fig 1). After a 2 week steroid washout period, a 4 week run-in period with fluticasone 250 µg twice daily, and a baseline bronchial allergen challenge, eligible patients were randomised to receive twice daily treatment with either fluticasone (250 µg) or fluticasone in a combination inhaler with salmeterol (250/50 µg) for 1 year. Patients were provided with rescue salbutamol 200 µg (GlaxoSmithKline, Zeist, The Netherlands). All drugs were administered via a dry powder inhaler (Diskus).

Outcome measures

Primary outcomes were sputum eosinophil numbers and eosinophil cationic protein concentrations. Secondary outcomes were neutrophil associated sputum parameters and a respiratory membrane permeability marker. In addition, lung function parameters, peak expiratory flows, symptom scores, and rescue medication usage were measured throughout the study. At the start of the washout period a full medical history, physical examination and forced expiratory volume in 1 second (FEV1) were performed. Baseline values of lung function parameters as well as primary and secondary outcomes were measured at the end of the run-in period (time point 0 months). In addition, outcomes were measured after washout and at 1, 3, 6, 9, 11 and 12 months during the randomised treatment period. Furthermore, outcomes were determined 24 hours before and 24 hours after bronchial allergen challenges which were performed at the end of the run-in period (pre-randomisation challenge) and at the end of the randomised treatment period (end of treatment challenge) (fig 1). Two week daily dairy cards, completed before and throughout the randomised treatment period, included peak expiratory flows, symptom scores, and rescue salbutamol usage. Before every visit patients abstained from rescue salbutamol for 8 hours and from the study medication for 12 hours except at 11 months of randomisation (abstaining from study medication for 36 hours).

FEV1 and the concentration of histamine provoking a fall in FEV1 of at least 20% (PC20histamine) were measured according to guidelines.11 Standardised allergen extracts were used for the allergen challenges which were performed as described by Sterk et al11 with modifications. A single dose of

<table>
<thead>
<tr>
<th>Table 1 Characteristics of study patients</th>
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<tr>
<td></td>
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<td>------------------------------------------</td>
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<tr>
<td>Age (years)*</td>
</tr>
<tr>
<td>M/F</td>
</tr>
<tr>
<td>Inhaled corticosteroids before study</td>
</tr>
<tr>
<td>(µg/day)*</td>
</tr>
<tr>
<td>Total IgE (IU/ml)†</td>
</tr>
<tr>
<td>Specific IgE (IU/ml)†</td>
</tr>
<tr>
<td>PC20histamine start run-in (mg/ml)†</td>
</tr>
<tr>
<td>PC20histamine end run-in (mg/ml)†</td>
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<tr>
<td>FEV1; start run-in (% predicted)‡</td>
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<tr>
<td>FEV1; end run-in (% predicted)‡</td>
</tr>
<tr>
<td>Morning PEF end run-in (l/min)‡</td>
</tr>
<tr>
<td>Evening PEF end run-in (l/min)‡</td>
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<tr>
<td>Morning symptom score end run-in</td>
</tr>
<tr>
<td>(scale 0–4)</td>
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<tr>
<td>Evening symptom score end run-in</td>
</tr>
<tr>
<td>(scale 0–5)‡</td>
</tr>
<tr>
<td>Short acting β2 agonist usage</td>
</tr>
<tr>
<td>(puffs/day)‡</td>
</tr>
</tbody>
</table>

IgE, immunoglobulin E; specific IgE, serum level of specific IgE directed against the type of allergen used during allergen challenge; PC20histamine, concentration of histamine provoking a fall in FEV1 of at least 20%; FEV1, forced expiratory volume in 1 second; PEF, peak expiratory flow.

No significant differences in patient characteristics were observed at the end of the run-in period (defined as baseline) between the two groups. During the run-in period FEV1 and PC20histamine improved significantly (p<0.001).

Table 1 Characteristics of study patients

*Median (range).
†Geometric mean (geometric SD).
‡Mean (SD).
§Ifr agonist usage recorded during the 2 weeks preceding the end of run-in visit.

Figure 1 Study schedule. After a 2 week steroid washout period and a 4 week run-in period with fluticasone 250 µg twice daily, eligible asthma patients were randomised to receive twice daily treatment with fluticasone (250 µg) with or without salmeterol (50 µg) for 1 year. Baseline values were determined at the end of the run-in period. Bronchial allergen challenges were performed the day before randomisation and at the end of the randomised treatment period. Primary and secondary outcomes were measured after washout, before randomisation, at 1, 6, 9, and 12 months of randomisation, as well as 24 hours before and 24 hours after the bronchial allergen challenges (depicted by asterisks).
analyzed by measuring the relative coefficient of excretion in induced sputum (RCEs), which is the ratio of α2-macroglobulin to albumin in induced sputum. 17

Table 2 Inflammatory parameters at start and end of the run-in period

<table>
<thead>
<tr>
<th></th>
<th>Start run-in</th>
<th>End run-in</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sputum eosinophils (×10³/g)</td>
<td>2.1 (1.4)</td>
<td>0.6 (1.3)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Sputum eosinophils (%)</td>
<td>3.2 (1.3)</td>
<td>0.8 (1.3)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Sputum ECP (ng/ml)</td>
<td>97 (1.3)</td>
<td>65 (1.3)</td>
<td>0.06</td>
</tr>
<tr>
<td>Serum IL-5 (pg/ml)</td>
<td>1.1 (1.2)</td>
<td>0.6 (1.2)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Sputum neutrophils (×10³/g)</td>
<td>28.1 (1.3)</td>
<td>29.1 (1.3)</td>
<td>0.98</td>
</tr>
<tr>
<td>Sputum IL-8 (ng/ml)</td>
<td>400 (1.4)</td>
<td>338 (1.8)</td>
<td>0.62</td>
</tr>
<tr>
<td>Sputum MPO (µg/ml)</td>
<td>1308 (1.3)</td>
<td>1273 (1.3)</td>
<td>0.95</td>
</tr>
<tr>
<td>RCEs (×10³/g)</td>
<td>76 (1.2)</td>
<td>52 (1.2)</td>
<td>0.006</td>
</tr>
<tr>
<td>Sputum α2-macroglobulin (ng/ml)</td>
<td>1388 (1.3)</td>
<td>984 (1.3)</td>
<td>0.04</td>
</tr>
<tr>
<td>Sputum albumin (µg/ml)</td>
<td>24 (1.3)</td>
<td>21 (1.3)</td>
<td>0.99</td>
</tr>
</tbody>
</table>

All values are expressed as geometric mean (SE). ECP, eosinophil cationic protein; IL, interleukin; MPO, myeloperoxidase; RCEs, relative coefficient of excretion; representing the ratio of α2-macroglobulin to albumin in induced sputum.

Analysis of data

Statistical analyses were performed using SAS Version 8.2 (SAS Institute Inc, Cary, NC, USA). The study was designed to have 80% power to be able to detect a 50% difference in geometric means of the primary outcomes between the groups with a sample size of 54 subjects. Changes over the run-in period were determined using the Wilcoxon signed ranks test or, in cases of normally distributed data, the t test. Differences within and between the treatment groups were determined using mixed model ANOVA adjusted for differences at baseline. Differences in allergen induced changes were determined using ANCOVA and adjusted for baseline allergen induced changes. All p values are two tailed and levels <0.05 were considered significant.

Table 3 Mean differences in clinical and lung function parameters in the fluticasone/salmeterol (FP/S) group compared with the fluticasone (FP) group over the 1 year treatment period

<table>
<thead>
<tr>
<th></th>
<th>Mean difference FP/S – FP</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>PEF morning (l/min)*</td>
<td>29 (9)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>PEF evening (l/min)*</td>
<td>36 (9)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Symptom score morning (scale 0–4)*</td>
<td>−0.1 (0.1)</td>
<td>0.02</td>
</tr>
<tr>
<td>Symptom score evening (scale 0–5)*</td>
<td>−0.2 (0.1)</td>
<td>0.01</td>
</tr>
<tr>
<td>Short acting β₂ agonist usage (puffs/day)*</td>
<td>−0.9 (0.3)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>FEV₁ (% predicted)*</td>
<td>2.7 (1.5)</td>
<td>0.07</td>
</tr>
<tr>
<td>PC₂₀ histamine (doubling doses)†</td>
<td>0.7 (0.3)</td>
<td>0.02</td>
</tr>
</tbody>
</table>

Values are expressed as *mean or geometric mean (SE). PEF, peak expiratory flow; FEV₁, forced expiratory volume in 1 second; PC₂₀ histamine, concentration of histamine provoking a fall in FEV₁ of at least 20%.

RESULTS

Patients

Sixty patients agreed to participate in the study. Four did not fulfill the inclusion criteria after the run-in period and two withdrew during the run-in period for personal reasons, so 54 were randomised to receive one of the two treatment regimens. Patient characteristics, including age, dose of inhaled corticosteroids prior to the study and lung function parameters, were not significantly different between the treatment groups at baseline (end of run-in) (table 1).

Four patients in the fluticasone group failed to complete the study. Two of these patients withdrew 1 month after randomisation, one because of worsening of asthma symptoms and one was lost to follow up. The other two patients withdrew after 6 and 9 months of treatment, respectively, because of personal reasons. All patients in the fluticasone/salmeterol group completed the study.

Figure 2 (A) Morning peak expiratory flow rate and (B) short acting β₂ agonist usage. Values are mean (SE). There were significant differences in morning peak expiratory flow rate and short acting β₂ agonist usage between the two treatment groups (p<0.01).
decrease in the RCEs (p = 0.006). Overall, during the 1 year treatment period adjusted geometric mean (SE) values are shown. During the run-in period there was a significant decrease in the RCEs than in the fluticasone group (p = 0.001). The relative coefficient of excretion (RCE) was significantly lower in the fluticasone/salmeterol group than in the fluticasone group (p = 0.001). The relative coefficient of excretion is the ratio of α₂-macroglobulin to albumin in induced sputum.

**Randomised treatment period**

**Lung function**

Morning and evening peak expiratory flows, asthma symptoms scores, and short acting β₂ agonist usage were significantly improved throughout the randomised treatment period in the fluticasone/salmeterol group compared with the fluticasone group (table 3, fig 2).

In the fluticasone/salmeterol group there was a trend for a higher FEV₁ (table 3). PC₂₀ histamine was significantly higher in the fluticasone/salmeterol group over the 1 year treatment period measured on visits after abstaining for 12 hours from the study medication (table 3), but there was no significant difference in PC₂₀ histamine at the visit after 11 months of randomisation when patients abstained from the study medication for 36 hours (geometric mean (SE) baseline adjusted PC₂₀ histamine 1.6 (1.2) mg/ml in fluticasone group v 1.9 (1.3) mg/ml in fluticasone/salmeterol group, p = 0.53). There were no differences in numbers or severity of exacerbations between the two treatment groups (results not shown).

**Sputum eosinophils and ECP**

The geometric mean number of eosinophils in the fluticasone/salmeterol group expressed as percentage difference from that in the fluticasone group varied significantly between visits (p = 0.019). After 1 month of randomised treatment the number of sputum eosinophils in the fluticasone/salmeterol group was −71% (95% CI −92 to −51)% compared with the fluticasone group (p = 0.048, fig 3). However, at other time points there was no significant difference between the groups (6 months: 56% (95% CI −71 to 74); 9 months −0.14% (95% CI −77 to 335); 12 months: 61% (95% CI −43 to 358)). Overall, during the 1 year treatment period there were no significant differences between the two treatment groups in either numbers or percentages of sputum eosinophils (p = 0.72 and p = 0.85, respectively, fig 3). The geometric mean ECP level in the fluticasone/salmeterol group was 5.7% lower than in the fluticasone group (p = 0.88). There were no indications that the true geometric mean ECP concentration in the fluticasone/salmeterol group should have been more than 57% lower than in the fluticasone group to be detected as significant, given the results of the study.

**Sputum neutrophils, IL-8, MPO and serum IL-5**

Sputum neutrophil numbers, sputum IL-8, sputum MPO, and serum IL-5 were not significantly different between the two treatment groups throughout the randomised treatment period (p = 0.16, 0.87, 0.70 and 0.23 respectively, data not shown).

**Size selectivity of the respiratory membrane: relative coefficient of excretion (RCE)***

Overall, during the 1 year treatment period the RCE was significantly lower in the fluticasone/salmeterol group than in the fluticasone group (p = 0.001, fig 4).

**Effect of adding salmeterol on allergen induced changes in bronchial inflammation**

There were no significant differences between the treatment groups in the change in FEV₁ after the allergen challenges, which were preceded by the inhalation of salbutamol (p = 0.81).

**Sputum eosinophils and ECP**

Sputum eosinophils (fig 5) and ECP concentrations increased significantly after the pre-randomisation allergen challenge.
as well as after the end of treatment allergen challenge (table 4).

There was a trend for a smaller increase in the percentage of sputum eosinophils in the fluticasone/salmeterol group after the end of treatment allergen challenge (p = 0.09), but there were no significant differences between the two treatment groups after the end of treatment allergen challenge in the changes in numbers of sputum eosinophils and ECP concentrations (table 4).

Sputum neutrophils, IL-8, MPO and serum IL-5
The number of sputum neutrophils, sputum IL-8, sputum MPO, and serum IL-5 increased after the pre-randomisation allergen challenge (p < 0.001, 0.001, 0.005 and < 0.001, respectively) as well as after the end of treatment allergen challenge (p = 0.02, 0.06, 0.02, and 0.02, respectively, data not shown). There were no significant differences between the two treatment groups in the changes in these parameters after the end of treatment allergen challenge (p = 0.92, 0.85, 0.93 and 0.89, respectively, data not shown).

Size selectivity of the respiratory membrane: relative coefficient of excretion (RCEs)
There was no significant difference between the two treatment groups in the changes in the RCEs after the end of treatment allergen challenge (p = 0.15, table 4). Likewise, there were no significant differences between the two treatment groups in the changes in sputum eosinophils after the end of treatment allergen challenge (p = 0.33).

DISCUSSION
This is the first randomised clinical trial to investigate over a 1 year treatment period whether the improved clinical outcomes resulting from adding salmeterol to fluticasone are accompanied by an additional effect on bronchial inflammation. In agreement with earlier studies, peak expiratory flows, symptom scores, rescue medication usage, and bronchial hyperresponsiveness were significantly improved in the fluticasone/salmeterol group relative to the fluticasone group. Similar levels of sputum eosinophils, sputum ECP, sputum IL-8, sputum MPO, and serum IL-5 were found in both groups throughout the treatment period as well as after the bronchial allergen challenges. However, we did find significantly reduced ratios of sputum α2-macroglobulin and albumin in the fluticasone/salmeterol group relative to the fluticasone group. This secondary outcome, the relative coefficient of excretion (RCEs), was chosen as a marker of size selectivity of plasma protein permeation across the respiratory membrane.

The lack of a sustained effect of adding salmeterol on cellular bronchial inflammation, as found in induced sputum, is in agreement with a recent study by Overbeek et al23 who failed to show an additional anti-inflammatory effect in bronchial biopsy specimens of adding formoterol to low doses of budesonide during 16 weeks of treatment. Earlier in vitro24 25 and short term in vivo studies1 26 did report an initial anti-inflammatory effect which may be in line with our observation of a transient decrease in the number of sputum eosinophils 1 month after randomisation in the fluticasone/salmeterol group. This difference in sputum eosinophils at

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**Table 4** Allergen induced increase in inflammatory parameters

<table>
<thead>
<tr>
<th></th>
<th>Pre-randomisation allergen challenge</th>
<th>End of treatment allergen challenge</th>
<th>p value FP vs FP/S</th>
</tr>
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<tbody>
<tr>
<td></td>
<td>(n = 54)</td>
<td>Fluticasone (FP) (n = 23)</td>
<td>Fluticasone/salmeterol (FP/S) (n = 27)</td>
</tr>
<tr>
<td>Sputum eosinophils (n)</td>
<td>3.7 (1.9 to 7.1)</td>
<td>4.4 (2.4 to 8.1)</td>
<td>2.6 (1.0 to 6.8)</td>
</tr>
<tr>
<td>Sputum eosinophils (%)</td>
<td>2.4 (1.5 to 3.8)</td>
<td>3.4 (1.9 to 6.1)</td>
<td>1.8 (0.9 to 3.8)</td>
</tr>
<tr>
<td>Sputum ECP</td>
<td>2.0 (1.2 to 3.4)</td>
<td>2.1 (1.0 to 4.4)</td>
<td>1.4 (0.7 to 2.5)</td>
</tr>
<tr>
<td>RCEs</td>
<td>1.4 (1.0 to 1.8)</td>
<td>0.8 (0.5 to 1.3)</td>
<td>1.3 (0.9 to 1.8)</td>
</tr>
<tr>
<td>Sputum α2-macroglobulin</td>
<td>1.3 (1.0 to 1.7)</td>
<td>1.2 (0.7 to 2.0)</td>
<td>1.0 (0.7 to 1.5)</td>
</tr>
<tr>
<td>Sputum albumin</td>
<td>0.9 (0.6 to 1.3)</td>
<td>1.5 (0.9 to 2.5)</td>
<td>0.8 (0.5 to 1.3)</td>
</tr>
</tbody>
</table>

Data shown as geometric mean (95% confidence interval) values.

The increases were calculated from the levels 24 hours after relative to the levels 24 hours before the bronchial allergen challenges.

ECP: eosinophil cationic protein; RCEs: relative coefficient of excretion, representing the ratio of α2-macroglobulin to albumin in induced sputum.
1 month after the pre-randomisation allergen challenge may be explained by a slower recovery from the allergen induced increase in bronchial inflammation in the fluticasone group. Bronchial allergen challenge increases levels of inflammatory markers, which may increase the sensitivity to detect differences in levels of inflammatory markers between the treatment groups. On the other hand, it should be noted that the medium to high daily dose of fluticasone resulted in relatively low baseline levels of the inflammatory parameters. This may have obscured a modest long term anti-inflammatory effect on cellular bronchial inflammation of adding salmeterol. The study was powered to detect a 30% difference in primary outcomes (sputum eosinophils and ECP) between the groups, since such a difference was considered to be clinically relevant. Therefore, adding salmeterol to an inhaled corticosteroid under the conditions chosen neither causes a clinically relevant deleterious or masking long term effect on cellular bronchial inflammation nor significantly improves it. It should be emphasised that this finding may be unique to the administration of the combination product of salmeterol and fluticasone as patients may be tempted to leave off the latter when using both medications separately. It was reported previously that reducing the dose of inhaled corticosteroids in patients on salmeterol can mask increasing inflammation and delay awareness of worsening asthma. Moreover, we found a significant decrease in the RCEs. This finding is in keeping with an earlier study. Moreover, specific serum albumin binding proteins have been identified. The role of albumin in the airway secretions is at present unclear. There is evidence that albumin may bind various mediators such as leukotrienes and may therefore render potentially active luminal agents less effective. Albumin may also act as a luminal antioxidant, preventing the formation of oxygen free radicals. In this respect, increased levels of albumin in sputum may have physiological advantages.

In summary, we have shown that improved clinical outcomes resulting from adding a long acting β2 agonist to maintenance treatment with inhaled corticosteroids are accompanied by similar levels of markers of chronic as well as allergen induced cellular bronchial inflammation. However, size selectivity of plasma protein permeation across the respiratory membrane appeared to be significantly improved by adding salmeterol to fluticasone, and this may contribute to the improved clinical outcomes seen.

**ACKNOWLEDGEMENTS**

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An extended version of the Methods section is available in the online data supplement at http://www.thoraxjnl.com/supplemental.

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Effects of terbutaline and budesonide on sputum cells and bronchial hyperresponsiveness in asthma. Am J Respir Crit Care Med 2000;161:1459–64.


Effects of terbutaline and budesonide on sputum cells and bronchial hyperresponsiveness in asthma. Am J Respir Crit Care Med 2000;161:1459–64.


Adding Salmeterol to an Inhaled Corticosteroid: Long-term Effects on Bronchial Inflammation in Asthma

Julia G. Koopmans, René Lutter, Henk M. Jansen and Jaring S. van der Zee

Online Data Supplement
Extended Material and Method

Patients

Patients with documented, mild to moderate persistent, allergic asthma (GINA II and III) [1] were enrolled (Table 1). The following inclusion criteria were used: 1. sensitization to house dust mite (*Dermatophagoides pteronyssinus*) and/or cat dander and/or grass pollen, as determined by Radio-Allergo-Sorbent-Test (RAST) and skin prick test; 2. age between 18 and 60 years; 3. FEV$_1$ $\geq$ 70 % of the predicted value after maximal bronchodilation; 4. bronchial hyperresponsiveness to histamine, PC$_{20}$histamine $\leq$ 8.0 mg/ml at the end of the run-in period; 5. clinically stable disease, no exacerbations of asthma within 3 months prior to inclusion requiring oral steroids and/or antibiotics; 6. no changes to regular asthma medication during 4 weeks before entry; 7. able to correctly inhale via a Diskus inhaler; 8. able to perform reproducible lung function tests.

Exclusion criteria were as follows: 1. comorbidity likely to interfere with the study; 2. lower respiratory tract infection during 4 weeks before entry; 3. use of theophylline, sodium cromoglycate, nedocromil sodium or antileukotrienes during the study or antibiotics 4 weeks prior to the study; 4. current smoking, regularly smoking within 6 months before entry or a smoking history of more than 10 pack years; 5. pregnant or lactating females; 6. unable to follow the therapy instructions; 7. participation in another clinical trial within 4 weeks prior to the study. The participating patients were recruited via advertisement and via the outpatient department of Pulmonology at the Academic Medical Center (AMC). The Medical Ethical Committee approved the study and all subjects gave written informed consent.

Study design

A schedule of the study design is shown in Figure 1. The study had a double-blind, randomized, two-armed parallel design (Figure 1). After a 2-week steroid wash-out period, a 4-week run-in period with fluticasone propionate 250 $\mu$g twice daily and a baseline bronchial allergen challenge, eligible patients were randomized to receive 1 year of twice daily treatment with either fluticasone propionate (250 $\mu$g) or twice daily treatment with fluticasone in a combination inhaler with salmeterol (250/50 $\mu$g). Patients were provided with rescue salbutamol 200 $\mu$g (GlaxoSmithKline, Zeist, The Netherlands) for relieve of symptoms during the study. All drugs were administered via a dry powder inhaler (Diskus). At the start of the wash-out period a full medical history, physical examination and FEV$_1$ were performed. Baseline values were measured at the end of the run-in period. Other visits were scheduled after the wash-out period and during the randomized treatment period after 1, 6, 9 (no PC$_{20}$histamine), 11 (no inflammatory parameters) and 12 months of randomization. In
addition, a bronchial allergen challenge was performed one day before randomization and at the end of the randomized treatment period. In order to measure differences in allergen-induced changes between both groups outcomes were determined 24 hours before and 24 hours after bronchial allergen challenges, which were performed at the end of the run-in period (pre-randomization challenge) and at the end of the randomized treatment period (end-of-treatment challenge) (Figure 1). A daily dairy card was completed during two weeks preceding every visit, an additional dairy card was completed after 3 months of randomization. The cards contained peak expiratory flows, rescue-short-acting β2-agonist usage and morning and evening symptoms scores (score 0-4 and 0-5 respectively). Before every visit patients abstained from rescue salbutamol (except before the end-wash-out visit) for 8 hours and from the study medication for 12 hours. Before the visit at 11 months of randomization patients abstained from the study medication for 36 hours, to exclude the influence of bronchodilatory effects of salmeterol on the level of PC20histamine. Asthma exacerbations were treated either by increasing the dose of fluticasone (defined as a mild exacerbation) or by oral glucocorticosteroids (moderate exacerbation), as judged by the investigator.

*Lung function and allergy tests*

The FEV1 was measured at each visit with a dry rolling seal spirometer (Sensor Medics BV, The Netherlands) according to standardized guidelines [2]. Values are expressed as the percentage of the predictive value. Bronchial hyperresponsiveness to histamine (PC20histamine) was determined by a 2-minute tidal breathing method [2]. PC20histamine was not performed if FEV1 was \( \leq 60\% \) of the predicted value or if salbutamol could not be abstained during 8 hours before the measurement. Total and specific Immunoglobulin-E directed against house dust mite were determined, as reported previously [3].

Bronchial allergen challenge

A standardized *Dermatophagoides pteronyssinus*, cat dander or grass pollen extract (containing 50,000 biological units (BU), ALK Abelló, Nieuwegein, The Netherlands) was used for the bronchial allergen challenges. Dilutions of the allergen extract, containing 5000 BU/ml were kept at – 20 °C in aliquots of 0.5 ml. A dilution, containing 200 BU/ml, was made freshly from the stock immediately before use, in phosphate buffered saline, 0.03% human serum albumin, 0.5% phenol (ALK Abelló, Nieuwegein, The Netherlands), of which a 0.5 ml sample was nebulized during the bronchial allergen challenge. A reservoir aerosol delivery system was used according to the method described by Sterk and colleagues [2] with
modifications [4]. Briefly, a collapsible reservoir of approximately 30 liter, made of static field dissipative material (RCAS 1206, Richmond Redlands, CA, USA) and filled with dry air, was connected to a nebulizer (Mallinckrodt Diagnostica, Petten, The Netherlands) producing aerosols from a 0.5 ml sample of diluted HDM extract. Previous studies demonstrated a 70% recovery of nebulized allergen. The amount of allergen was not titrated on the decline in FEV$_1$ during the early asthmatic reaction. For patients convenience all bronchial allergen challenges were preceded 15 minutes before the challenge by the inhalation salbutamol hemisulphate, 500 mg/ml solution, which was nebulized, via a Micro-Mist nebulizer, (Hudson, Ternecula, California, USA) driven by dry-compressed air, at a flow of 5 L/min during 1.5 min. The dose is equivalent to approximately 400 µg salbutamol via a dry powder inhaler, considering the loss of drug by delivering during both inspiration and expiration [5]. The amount of allergen administered to the patients (70 % of 100 biological units, (for Dermatophagoides pteronyssinus equivalent to 42 ng major allergen Der p1) was based on the results of an earlier study which was done in asthma patients with a similar range of severity [6]. However, the patients in that study abstained from anti-inflammatory therapy 6 weeks prior to allergen challenge [6]. The median cumulative dose of allergen that resulted in a decline in FEV$_1$ of 20 % from baseline was 107 biological units. Baseline FEV$_1$ (median of three measurements) was measured with a dry rolling seal spirometer (Sensor Medics BV, The Netherlands) as well as with a portable spirometer (Micromedical diarycard, Sensor Medics BV, The Netherlands). After allergen challenge FEV$_1$ was determined at 10 and 30 minutes with the dry rolling seal and subsequently hourly from 4 to 8 hours after the allergen inhalation with a portable spirometer (best of two measurements). Change in FEV$_1$ after allergen challenge was expressed as percentage from baseline, as measured with the corresponding spirometer.

Sputum induction
Sputum induction was performed by inhalation of aerosols of hypertonic saline (NaCl) during quiet tidal breathing. An Aerodyne Omega Ultrasonic nebulizer (Kendall, Neustadt/ Donau, Germany) was used with plastic tubes of 30-cm length. This nebulizer generates aerosols with a mass median aerodiameter of 4.5 µm. Before inhalation of the nebulized saline was started, a baseline FEV$_1$ was recorded. For safety reasons, all subjects received salbutamol (nebulized dose of 400 µg). Each subject inhaled increasing concentrations of nebulized, sterile saline during 21 minutes, namely 3, 4 and 5 % sodium chloride for 7 minutes each. Spirometry was repeated between each concentration step; only if there was no change in FEV$_1$ or if the fall in FEV$_1$ was less than 10 % the induction procedure was proceeded to the next inhalation; if the
fall in FEV\textsubscript{1} was between 10 and 20% the same concentration was inhaled for another 7 minutes. The induction procedure was discontinued if FEV\textsubscript{1} was fallen by more than 20%, or when patients experienced any discomfort like chest tightness or dyspnea caused by the induction procedure. Before expectorating the sputum subjects were asked to blow their nose and clear their throats and subsequently carefully rinse their mouths with water and swallow the water. Sputum samples were collected in plastic containers on ice. Processing was started within 15 minutes.

Sputum processing

The volume of the whole sputum sample was determined and an equal volume of dithiotreitol (10 mM DTT in 135 mM Tris buffer, pH 8.0) was added. The samples were then mixed gently at 4 °C for 15 min. In case a sample was still not homogenized after this procedure, DNAse 1:1000 was added and the sample was mixed at 4 °C for another 15 min. The homogenized sputum was centrifuged at 1640 rpm at 4 °C for 10 min. The supernatants were aspirated and frozen at −20 °C for later analysis. The cell pellets were resuspended in 1 ml 2% Human Serum Albumin (HSA) in phosphate buffered saline (PBS). Total cell number was determined by counting manually in a Bürker counting chamber. Cells were cytocentrifuged for 2 minutes at 550 rpm in Shandon Cytocentrifuge and stained with Romanovsky (Diff-Quick) and Jenner-Giemsa. One investigator, blinded for the subject’s history, counted 200 non-squamous cells on each sputum slide; squamous and non-squamous epithelial cells, macrophages, lymphocytes, neutrophils and eosinophils were identified. Differential cell counts were expressed as number per gram sputum and as percentage of cells, excluding squamous epithelial cells. In case the percentage of eosinophils was less than 10% but more than 1%, a total of 500 non-squamous cells was counted; 1000 non-squamous cells were counted in case the eosinophil percentage was less than 1%. Sputum samples containing more than 80% squamous cells on differential cell counting were excluded from cell differential analysis.

**Protein assays**

Levels of ECP were determined with an ELISA [7]. The detection limit was 15 pg/ml. Assay reagents (Rabbit-anti-human ECP antiserum and anti-ECP antibody-biotin) were kindly donated by dr. A. Zuurbier (CLB Sanquin, Amsterdam, The Netherlands). A series of standard dilutions of ECP was obtained from Pharmacia & Upjohn (Uppsala, Sweden). Major basic protein (MPO) [8] and interleukin (IL)-8 [9] were measured with an ELISA as described earlier. Size selectivity of plasma protein leakage across the respiratory membrane, into the
airway lumen was analyzed by measuring the relative coefficient of excretion (RCE₂), which is defined as the ratio of α2-macroglobulin and albumin in valid sputum samples [10,11]. The levels of albumin were measured by an immunoturbidimetric assay with a Cobas Bio analyzer (Roche Diagnostics, Inc.). Antiserum for albumin was obtained from Dako (code A001, Glostrup, Denmark). As a standard we used N protein standard serum for nephelometry (Behring, Marburg, Germany). The levels of A2M were measured with an ELISA [11].

**Statistical analysis**

SAS (SAS Institute Inc., Cary, NC, USA version 8.2) was used for statistical analyses. The study had 80% power to detect a 50% difference in change from baseline between the groups with a sample size of 54 subjects. Changes over the run-in period were determined using Wilcoxon signed ranks test, or in case of normally distributed data with t-test. Differences within and between the treatment-groups were determined using a mixed model ANOVA, adjusted for differences at baseline (end of run-in). Differences in allergen-induced changes were determined using ANCOVA and adjusted for baseline allergen-induced changes. Differences between the treatment groups in changes from baseline in PC₂₀histamine at the visit at 11 months of randomization were analyzed by multiple linear regression analysis, including baseline PC₂₀histamine as co-variable. All p-values are two-tailed and p-levels of less than 0.05 were considered significant.
Reference List


