Inhalated corticosteroids (ICS) are regarded as the first line of anti-inflammatory treatment for asthma because of their lower systemic adverse effects compared with oral corticosteroids. Nonetheless, the possibility of unwanted systemic effects following long term use of ICS is still a concern, especially among pediatricians. The side effects seem to be mainly dose dependent. Low doses of fluticasone propionate (200 µg/day) or budesonide (400 µg/day) are usually considered safe, although adrenal suppression has been reported even at these doses. The optimal test to detect adrenal suppression is still a matter of debate. Assessment of adrenal function by measuring morning serum cortisol levels and urinary free cortisol (UFC) excretion provides information on the basal endogenous adrenocortical activity and not on the ability of the axis to react to stress. Furthermore, UFC is a valuable tool in conditions resulting from high cortisol levels, but it is not useful when hypocorticism is suspected. This is related to the lack of sensitivity of the method at low cortisol levels and because low cortisol excretion is often found in healthy individuals. Despite these limitations, morning basal cortisol concentrations and UFC excretion are frequently used by respiratory physicians to evaluate the adrenal suppressive effect of ICS. Furthermore, it has been observed that state of the art tests—which are adrenocorticotropic hormone (ACTH) stimulation test and measurement of the 24 hour integrated serial plasma cortisol concentration—have been inappropriately used. It has recently been shown in small number of adult asthmatics that the corticotrophin releasing hormone (CRH) test, which stimulates pituitary ACTH secretion and subsequently cortisol secretion from the adrenal glands, provides a more accurate assessment of the function of the entire hypothalamic-pituitary-adrenal (HPA) axis than the ACTH test. The CRH test is also a sensitive tool for evaluating the ability of the HPA axis to react to stress.

The aim of this study was to evaluate the systemic effect of ICS treatment, as detected by morning serum cortisol levels, 24 hour UFC excretion, and the CRH test, to compare the relationship between basal and systemic activity of the adrenal glands in asthmatic children.

METHODS

Patients:
Thirty two prepubertal children suffering from frequent episodic or persistent asthma who required maintenance treatment with ICS were studied. Sixteen received inhaled fluticasone propionate (FP) by a metered dose inhaler (MDI) and spacer (Aereochamber), 100 µg twice daily (50 µg per actuation), and 16 received budesonide (BUD), 200 µg twice daily by Turbhaler (200 µg per actuation). Two subjects in the FP group failed to present at the scheduled visits and did not complete the study. Data are therefore presented on 30 patients (14 treated with FP and 16 treated with BUD) of mean (SD) age 6.7 (1.8) years, a height standard deviation score of 0.45 (0.77), body mass index (BMI) of SDS (0.98), and body surface area 1.05 (0.35). The children were supervised in the correct use of the devices according to the manufacturers’ recommendations. Each dose was given at 08.00 and 20.00 hours. No child had been treated with oral or nasal corticosteroids in the previous 6 months and none had received inhaled corticosteroids. None of the children suffered from enuresis.

Study design
The HPA axis was evaluated in all children at baseline and 12 weeks after treatment with ICS. After an overnight fast and...
after the first dose of ICS, a CRH test (1 µg/kg IV as bolus) was performed at 09.00 hours to assess serum cortisol and ACTH levels (samples drawn at the following times: –15, 0, 15, 30, 45, 60, 90, 120 minutes). Twenty four hour urine samples were collected at home on the previous day for UFC assessment. All samples were frozen at –20°C until assayed.

Cortisol was assayed with Immulite 2000 cortisol kit (DPC, Los Angeles, CA, USA) with a sensitivity of 0.02 µg/ml with an intra-assay coefficient of variation (CV) of 7.2%. ACTH was assayed with the Immulite 2000 ACTH kit (DPC, Los Angeles, CA, USA) which has a sensitivity of 5 pg/ml with intra- and inter-assay CVs of 6.7% and 8.2%, respectively. UFC excretion was measured with Immunotech cortisol kit (Mar- seille, France) which has a sensitivity of 0.36 µg/ml with intra- and inter-assay CVs of 3.5% and 7.5%, respectively.

**Statistical analysis**

The aim of this study was to evaluate the behaviour of the basal and reserve activity of the HPA axis after treatment with ICS and not to compare the effect of different corticosteroids. We therefore evaluated all the patients together to study the relationship between basal morning serum cortisol levels, the peak cortisol level and the area under the curve (AUC) during the CRH test was calculated according to the trapezoidal rule. A p value of <0.05 was considered statistically significant. Data were reported as mean values and standard deviations (SD). All statistical calculations were performed using SAS statistical software (SAS, Cary, NC, USA) for personal computers.

The study was approved by the medical ethics committee of our hospital and parents gave written and verbal informed consent for their children.

**RESULTS**

No significant change in basal serum cortisol levels (mean change –18; 95% CI –41 to 5; p=0.118) or delta (peak minus basal) levels (mean change –13; 95% CI –38 to 12; p=0.308) was seen after treatment, but a significant reduction in the peak cortisol level (mean change –31; 95% CI –55 to –7; p=0.013) and AUC (mean change –175; 95% CI –288 to –63; p=0.003; table 1) was seen after the CRH test.

<table>
<thead>
<tr>
<th>Variable</th>
<th>Before</th>
<th>After</th>
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<th>p value*</th>
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<tr>
<td>Cortisol (ng/ml)</td>
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<td>Basal</td>
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<td>65</td>
<td>137</td>
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<td>Peak</td>
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<tr>
<td>ACTH (pg/ml)</td>
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<tr>
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<tr>
<td>Delta</td>
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<tr>
<td>AUC</td>
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<td>7.1</td>
<td>15.0</td>
<td>0.1</td>
</tr>
</tbody>
</table>

UFC=urinary free cortisol.

Values are mean (SD).

*Paired t test.

After the 12 week treatment period four patients, two treated with FP and two with BUD, who had a normal response before treatment had an abnormally low peak serum cortisol level (168 and 170 ng/ml) in the patients treated with FP, 165 and 161 ng/ml in the patients treated with BUD, normal value >180 ng/ml; fig 1). In these four children, however, the basal serum cortisol level was normal (68 and 120 ng/ml in FP treated patients, 97.8 and 113 ng/ml in BUD treated patients, normal value >50 ng/ml).

Basal (mean change –14, 95% CI –27 to –0.4, p=0.045), peak (mean change –26, 95% CI –43 to –9, p=0.004), and AUC (mean change –77, 95% CI –124 to –29, p=0.003) ACTH levels were significantly lower after treatment, while delta ACTH was similar before and after treatment (mean change –12, 95% CI –31 to 7, p=0.199; table 1). In three of the four patients with a subnormal peak cortisol response after ICS treatment, the peak ACTH levels were reduced even if they were within the range of our study population, whereas in one patient the ACTH levels showed no change after treatment.

No significant reduction was observed in 24 hour UFC excretion (µg/24 hours) after treatment (14.9 (7.1) v 15.0 (11.6), normal range 2–27 µg/24 hours; mean change 0.1, 95% CI –5.2 to 5.4, p=0.967; table 1). In three of the four patients with a subnormal peak cortisol response there was also a reduction in 24 hour UFC excretion which was, however, within the observed range of our study population. Nonetheless, one patient showed no changes after treatment.
No correlation was found between UFC and any of the parameters of cortisol excretion following the CRH test, either before or after treatment with ICS.

DISCUSSION
Assessment of HPA axis function appears to be the most sensitive and easily measured parameter of the potential systemic side effects of ICS treatment. The clinical significance of slight alterations in the HPA axis is uncertain and, in most patients, the presence of suppressed endogenous cortisol is not clinically relevant in itself. However, it can be used as a marker of systemic activity of steroid treatment as well as a surrogate marker for potential adverse effects in other tissues. Based on this, the aim of our study was to assess the function of the HPA axis in a group of corticosteroid naïve asthmatic children before and after treatment with ICS.

Two broad types of tests can be used to detect HPA axis function—namely, measurement of basal adrenocortical activity and the dynamic stimulation test to evaluate adrenocortical reserve. In our study the dynamic evaluation of the HPA axis seems to be more sensitive than basal serum cortisol levels and 24 hour UFC excretion in detecting the systemic effect of corticosteroid treatment. Following the CRH test, four of our children (two treated with FP and two with BUD) had an inadequate response which could not have been predicted by normal morning serum cortisol levels (fig 1). The CRH test enables suppression of pituitary ACTH secretion to be distinguished from the more direct action on the adrenal response to ACTH. The CRH test stimulates pituitary ACTH secretion and this, in turn, produces a cortisol signal from the adrenal gland. It has been suggested that this physiological stimulation by including pituitary assessment may be more sensitive at detecting systemic steroid induced effects than basal adrenal testing alone, particularly with low dose ICS treatment.

In our study the lower peak ACTH secretion after treatment suggests that the suppression is mainly at the HPA level. Even though there were trends for both cortisol and ACTH reduction after treatment in all the parameters measured (table 1), only some of these changes reached statistical significance, reflecting the variability of the measurements and the changes over time, in addition to the effects of ICS treatment. More patients would improve the statistical power and the changes over time, in addition to the effects of ICS treatment.

The 24 hour UFC excretion did not change after treatment in our study population. This was also the case in the four children with a reduced peak cortisol response; although it was reduced, 24 hour UFC excretion remained within the normal range. Furthermore, we did not find any correlation between 24 hour UFC excretion and adrenal dynamic response. This was in agreement with the findings in a study in adults. The on the other hand, other studies in adult patients have shown that suppression of 24 hour UFC excretion by ICS correlates with blunting of the stimulated cortisol response to CRH or to a physiological low dose of ACTH. These different results can be explained either by inter-individual sensitivity to the systemic effect or by different doses of ICS and their dose-response curve for adrenal suppression. Furthermore, Fink and colleagues measured UFC in 13 adults subjects after ICS administration (FP or BUD) and concluded that UFC estimation may be an unreliable surrogate marker of adrenal suppression.

Some patients are particularly sensitive to the systemic effects of ICS, which can be occasionally clinically relevant. In our patients there was no clinical evidence of adrenal suppression; however, the CRH test proved to be particularly sensitive to detecting the systemic activity of ICS, especially with respect to the ACTH response. The CRH test is expensive but it is not associated with potential side effects observed with other dynamic tests, such as metyrapone and insulin-induced hypoglycaemia. Metyrapone induced diziness and nausea, as well as hypoglycaemia, and cannot be used in paediatric studies. The ACTH test might be an alternative, but it only gives an indirect evaluation of the HPA axis and it has already been shown in adult patients to be less accurate than the CRH test. Furthermore, both the standard (250 µg) and low dose (1 µg) ACTH tests lack the sensitivity and specificity to diagnose ACTH deficiency compared with the metyrapone test in adults. Moreover, the ACTH test is contraindicated in allergic and asthmatic subjects because of occasional reports of severe anaphylaxis.

In conclusion, our results suggest that a thorough evaluation of adrenal function cannot be obtained simply by measuring basal serum cortisol concentrations and/or 24 hour UFC excretion. In studies evaluating the systemic effects of ICS, dynamic evaluation of the HPA axis is more appropriate and CRH should provide an alternative research tool in asthmatic children.

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REFERENCES
3 American Academy of Allergy, Asthma and Immunology. Pediatric asthma: promoting best practice. Guides for managing asthma in children. Milwaukee, WI: American Academy of Allergy, Asthma and Immunology, 1999: 66-8
7 Lipworth BJ. Systemic adverse effects of inhaled corticosteroid therapy: a systemic review and meta-analysis. Arch Intern Med 1999; 159:941-55
8 Lipworth BJ, Clark DJ, McFarlane M. Adrenocortical activity with repeated twice daily dosing of fluticasone propionate and budesonide given via a large volume spacer to asthmatic school children. Thorax 1997; 52:686-9
No cure for the common cold


Previous studies of Echinacea have reported success varying from 10% to 50% in the treatment of colds. A few randomised trials have reported only a small effect in preventing colds.

In this randomised, double blind, placebo controlled study the authors report on the effect of 10 days of treatment with dried, encapsulated, whole plant Echinacea starting within 36 hours of onset of symptoms in 142 students reporting common colds (69 Echinacea, 73 placebo). Their primary objective was to measure the severity and duration of reported upper respiratory tract infections. They concluded that there was no statistical difference between the two groups. They do, however, believe this should not be the last word on the use of Echinacea in the treatment of colds because (1) they used whole plant mixture rather than extracts as reported in previous trials, (2) they studied a healthy population of students in whom the clinical effect would be small, and (3) the trial was of modest size and would have missed an effect of 5–10%. Furthermore, they did not confirm infection by serological testing or inflammatory markers and simply based it on reported symptoms. This raises doubt about the exact aetiology of the illness.

This is a well designed study which clearly shows that whole plant Echinacea has no significant clinical effect on the course and severity of the common cold, although it is unlikely that this will end discussion on this subject.

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