

Evolving concepts on the value of adenosine hyperresponsiveness in asthma and chronic obstructive pulmonary disease

R Polosa, S Rorke, S T Holgate

Thorax 2002;**57**:649–654

Adenosine is a purine nucleoside which mediates a variety of cellular responses relevant to asthma and COPD through interaction with specific receptors. Administration of adenosine by inhalation to patients with asthma and COPD is known to cause concentration related bronchoconstriction. Responses elicited by this purine derivative in asthma and COPD should not be considered as a mere reflection of non-specific airways hyperresponsiveness. Evaluation of airways responsiveness by adenosine induced bronchoconstriction may be valuable in differentiating asthma from COPD, monitoring of anti-inflammatory treatment in asthma, surveying disease progression, and assessing disease activity in relation to allergic airways inflammation.

porting this view.^{8–10} Elucidation of the fine mechanisms of adenosine induced bronchoconstriction has provided convincing evidence that responses elicited by this purine derivative in asthma and COPD are not a mere reflection of non-specific airways hyperresponsiveness but involve a selective interaction with activated inflammatory and structural cells.

This paper reviews the mechanism(s) by which adenosine mediates bronchoconstriction in asthma and COPD, the evidence in favour of the hypothesis that airway response to adenosine may better discriminate the inflammatory and immunological processes in asthma and COPD, and the possibility that adenosine responsiveness may represent a distinctive marker of disease severity and progression.

MECHANISM OF ADENOSINE INDUCED BRONCHOCONSTRICTION IN ASTHMA AND COPD

Despite the evidence that inhaled purine derivatives elicit dose related bronchoconstriction in patients with asthma and COPD,^{6,7} the action of adenosine on airway smooth muscle *in vitro* is conflicting, varying between species and, in the same species, varying with the type of preparation, the initial level of smooth muscle tone, and the concentration of the nucleoside used. In isolated guinea pig airway with high resting tone induced by carbachol, adenosine causes relaxation via an A₂ receptor mechanism,^{11,12} whereas constriction occurs when the preparation is maintained at intrinsic tone.¹³ In isolated human airway preparations the predominant effect of the nucleoside is contractile, although the effect is weak.¹⁴ However, bronchial preparations obtained from asthmatic subjects were more sensitive to the contractile effects of adenosine than those obtained from non-asthmatic controls,¹⁵ and when inhaled by asthmatics adenosine provoked bronchoconstriction that was not elicited in normal individuals.⁶

The adenosine nucleotides AMP and ADP are equipotent with the parent nucleoside.¹⁶ As neither has any effect on adenosine receptors,¹⁷ but both can be rapidly converted to adenosine by 5'-nucleotidase, it is likely that these nucleotides act *in vivo* after prior conversion to adenosine. Since AMP in particular is more soluble in aqueous solution, allowing higher concentrations of agonist to be delivered by aerosolisation, it has replaced adenosine as the most frequently used purine nucleoside bronchoprovoking agent.

Adenosine is a purine nucleoside which has the capacity to elicit a variety of cellular responses relevant to asthma and chronic obstructive pulmonary disease (COPD) through interaction with specific cell surface purinoreceptors as indicated by the ability of the adenosine uptake inhibitor, dipyridamole, to enhance adenosine induced effects.^{1–3} On the basis of molecular cloning and ligand affinity data, adenosine receptors are currently classified into four subtypes—A₁, A_{2A}, A_{2B}, and A₃—each with their unique patterns of tissue distribution and signal transduction.^{4,5}

In 1983 Cushley *et al*⁶ were the first to administer aerosolised adenosine to a group of asthmatic subjects. Whereas the nucleoside had no discernible effect on airway calibre in normal individuals, the asthmatics experienced concentration related bronchoconstriction with a maximum effect at 5 minutes and subsequent slow recovery that was complete by 45–60 minutes.⁶ Ten years later Oosterhoff *et al*⁷ reported hyperresponsiveness to adenosine administered by inhalation in 28 out of 30 patients with COPD. The severity of their response was significantly higher in the patients with COPD who smoked than in the non-smoking COPD patients, whereas no discernible difference in methacholine hyperresponsiveness was observed between the two groups.

Since these initial observations, a role for adenosine in asthma and COPD has been postulated and there have been several reviews which have set out in detail the key evidence sup-

See end of article for authors' affiliations

Correspondence to:
Dr R Polosa, Dipartimento
di Medicina Interna e
Specialistica, Ospedale
Tomaselli, Università di
Catania, Via Passo
Gravina 187, 95125
Catania, Italy;
rpolosa@hotmail.com

Since these observations were made, considerable effort has been directed at elucidating the mechanism by which adenosine mediates bronchoconstriction in asthma and COPD. Although no adenosine antagonists have acceptance for use in humans, alternative pharmacological approaches have suggested that it is unlikely that adenosine acts directly on smooth muscle cells *in vivo*, but indirectly through activation of purinoreceptors expressed on intermediary inflammatory cells such as mast cells or on afferent nerve endings.

It has long been recognised that mediator release from human mast cells contributes to the airflow limitation and accompanying symptoms of asthma.¹⁸ In active disease, immunohistochemical and altered structural analysis of submucosal and epithelial mast cells reveals that many of them are actively degranulating.^{18, 19} The role of the mast cell in the pathogenesis of COPD is more speculative. Increased levels of histamine have been found in the sputum of patients with obstructive bronchitis²⁰ and Postma *et al*²¹ have reported an increase in the urinary excretion of the N-methyl metabolite of histamine in the urine of patients with COPD. Lamb *et al*²² have also reported a greater number of mast cells in the respiratory epithelium in the distal airways of smokers than in non-smokers. Immunohistochemical analysis of bronchial mucosa obtained from patients with COPD reveals that larger numbers of mast cells are present in the bronchiolar epithelium than in the airways of smokers without airway obstruction.²³ Likewise, Pesci *et al*, who studied mast cell infiltration in bronchial biopsy specimens of subjects with chronic bronchitis, observed higher numbers of mast cells both in the epithelium and in the bronchial glands than in control subjects.²⁴ Relevant to this are the findings of de Boer *et al*²⁵ who have recently shown that the number of mast cells in the bronchiolar epithelium of COPD patients is strongly associated with the increased level of expression for epithelial transforming growth factor (TGF) β_1 , a well known chemotactic factor for mast cells.²⁶ Mediator secretion released from mast cells during the active “inflammatory” phase of COPD may therefore contribute to its airway pathophysiology. Indeed, mast cells may release chemotactic factors for neutrophils and secrete proteases—for example, tryptase, chymase, elastase—which are able to induce tissue injury,²⁷ airway smooth muscle hyperresponsiveness,²⁸ and airway mucus secretion.²⁹

Mast cells are likely to play a critical role in the bronchoconstrictor response to inhaled adenosine as indicated by *in vitro* studies in which adenosine markedly enhances the release of histamine and other preformed mediators from immunologically primed rodent mast cells.³⁰ The timing of adenosine addition seems quite critical to the effect produced, and pharmacological manipulations have suggested involvement of the A_{2B} receptor.³¹ A series of studies in human dispersed lung mast cells by Church *et al*³² and Peachell *et al*³³ have shown similar effects, including a small potentiation of leukotriene C₄ release in the latter study. Feoktistov and Biagioni have since shown that stimulation of the A_{2B} receptor in a human mast cell line *in vitro* produces cellular activation, and that phosphoinositide hydrolysis and intracellular calcium mobilisation are involved in this process.³⁴ Most of the above mentioned studies refer to mast cells obtained by either mechanical dispersion or enzymatic digestion of whole lung. In a recent study Forsythe *et al*³⁵ have produced evidence that adenosine can directly stimulate histamine release from human mast cells obtained by bronchoalveolar lavage.

There is also abundant evidence *in vivo* to indicate that the mast cell may be involved in the bronchoconstrictor response to inhaled adenosine, principally via release of granule derived preformed mediators. Premedication with the potent H₁ histamine receptor antagonists terfenadine and astemizole have been shown to inhibit the acute bronchoconstrictor response to inhaled AMP in asthmatic and COPD patients.^{36–39} These initial studies provided strong support for the concept that

mast cell derived mediators are implicated in the bronchoconstrictor response to inhaled adenosine in both asthma and COPD. More direct evidence that histamine released from airway mast cells is critical for adenosine induced responses has come from a study in which venous plasma histamine levels were measured after bronchial provocation with inhaled allergen and AMP in a group of atopic subjects. A small but significant increase in histamine levels was observed after AMP challenge.⁴⁰ Direct instillation of AMP into asthmatic bronchi⁴¹ or into the nose of patients with allergic rhinitis⁴² resulted in significant increases in the concentration of histamine and tryptase in their lavage fluid. However, in addition to histamine, a role for other mast cell derived mediators has to be considered. A role for prostanoids in the response to AMP is supported by the demonstration that potent cyclooxygenase inhibitors such as indomethacin and flurbiprofen attenuate the constrictor effect of the nucleotide.^{43, 44} In addition, lysine-aspirin administered by inhalation causes some attenuation of the AMP response.⁴⁵ More direct evidence for a role for newly generated mediators has come from the study by Polosa *et al*.⁴¹ In addition to the rise in histamine and tryptase levels in the bronchoalveolar lavage fluid, an even greater increase in concentrations of PGD₂ were found. Recently, premedication with ABT-761,⁴⁶ a potent 5-lipoxygenase inhibitor, and the selective cysteinyl leukotriene (Cys LT₁) receptor antagonist montelukast⁴⁷ has been shown to attenuate the acute bronchoconstrictor response to inhaled AMP, thus suggesting a role for spasmogenic leukotrienes.

Although activation of both cholinergic⁴⁸ and peptidergic neural pathways⁴⁹ may contribute to the contractile airway response to adenosine in asthma, the role of the neural pathway in adenosine induced bronchoconstriction in patients with COPD has not been fully addressed. In a recent study Reutgers *et al*⁵⁰ found no significant effect on AMP responsiveness after inhaled ipratropium bromide in patients with COPD, implying that vagal nerve activation does not play a role. This is at variance with the findings in asthmatic patients, where ipratropium bromide caused a significant increase in PC₂₀AMP.⁴⁸ It is possible that in asthma AMP stimulates mast cells to release histamine which causes an additive effect via vagal nerve stimulation. In COPD, histamine release may be smaller and inadequate to stimulate vagal nerve endings during AMP challenge.

CLINICAL VALUE OF AIRWAY RESPONSIVENESS TO ADENOSINE IN ASTHMA AND COPD

Airway or “bronchial” hyperresponsiveness (BHR) is best defined as airways that narrow too much to a provoking stimulus.⁵⁰ Although BHR is well established as a hallmark of asthma, the clinical and diagnostic relevance of airway responsiveness as currently defined is still unclear. BHR is neither sensitive nor specific for asthma,⁵¹ as it is also detected in approximately two thirds of smokers with COPD⁵² and in various other inflammatory airway diseases such as cystic fibrosis,⁵³ bronchiectasis,⁵⁴ and Sjogren's syndrome.^{55–56} Despite this lack of specificity and sensitivity, it remains an important physiological marker in diagnosing and determining asthma severity.

The provoking stimuli can be classified into two categories: (1) those that act predominantly directly on airway smooth muscle such as histamine and methacholine; and (2) those that act indirectly through the release of inflammatory mediators or stimulation of neural pathways such as adenosine. Airway hyperresponsiveness is often linked to the degree of airway inflammation and this is reflected by the number and state of activation of various inflammatory cells.^{57–59} At present there is no single effective marker of the underlying inflammatory process in the lungs, but various surrogate markers have been used to reflect the severity of airway inflammation. The non-invasive technique of measuring BHR

using either a direct or indirect stimuli may provide us with more information about the inflammatory process and enable us to differentiate between different disease processes.

As illustrated previously, adenosine is an indirect stimulus that exerts its effect primarily on inflammatory cells, subsequently leading to smooth muscle contraction. This distinctive feature of adenosine suggests that AMP responsiveness may correlate better than other stimuli with airway inflammation, enabling superior diagnostic discrimination between asthma and COPD and allowing better monitoring of disease activity and progression. It should also be noted that AMP challenge testing would be useful in evaluating response to treatment.

Although there is not enough information to establish clear indications for AMP challenge testing and a standardised population based cut off PC_{20} AMP value needs to be delineated, airway responsiveness to AMP may be used to differentiate asthma from COPD when traditional diagnostic methods have not established a clear diagnosis. In addition, it appears that bronchoprovocation with AMP may be a more robust marker of disease activity in relation to allergic airway inflammation than other non-specific stimuli such as histamine or methacholine and has a greater probability than methacholine for the diagnosis of asthma.⁸ AMP challenge may also be used as a practical tool in determining bronchial hyperresponsiveness in epidemiological surveys as described by De Meer and colleagues using the short dosimeter protocol method of AMP challenge.⁶⁰

Atopy is the single most important determinant of enhanced adenosine induced responses *in vivo*. Phillips *et al*⁴⁰ have shown that atopic subjects are more responsive than non-atopic controls to inhaled adenosine than they are to methacholine, indicating that the airway response to these purines may be an index of mast cell priming. In this context, it is of interest that adenosine potentiates the release of inflammatory mediators when human mast cells are immunologically primed *in vitro*.³³⁻⁶¹ Increased adenosine responsiveness in the form of heightened histamine release has also been shown in sensitised mice compared with non-sensitised controls.⁶² Moreover, nasal challenge with AMP elicits rhinitic symptoms and a rapid increase in histamine levels in the lavage fluid with a greater increase occurring in atopic than in non-atopic individuals.³⁶ Adenosine induced bronchoconstriction in asthmatic and atopic subjects may therefore be used as an index of mast cell priming *in vivo*. The role of atopy in adenosine induced bronchoconstriction is also emphasised in a recent study by van Daele *et al*.⁶³ These authors compared histamine and AMP bronchial challenges in preschool children with recurrent wheeze to identify atopic mechanisms for their wheezing and found that all non-atopic children with wheeze had a negative adenosine provocation test. Adenosine bronchoprovocation testing is therefore more specific than histamine in establishing allergic factors in preschool children with wheeze.

There is mounting evidence that adenosine challenges could possibly be more exploited in differentiating asthma from COPD in subjects where the diagnosis is clinically uncertain. In adults, AMP and methacholine provocation both distinguish subjects with COPD from normal controls. However, only AMP could separate non-smoking COPD patients from asthmatic patients. In the COPD patients who smoked, AMP responsiveness was similar to that found in asthmatic patients, perhaps as a result of the additional inflammatory effect of cigarette smoking.⁷ In children, bronchoprovocation tests with inhaled AMP appears to be considerably more specific and sensitive than methacholine at discriminating asthma from paediatric chronic obstructive lung disorders such as cystic fibrosis, bronchiolitis, pulmonary ciliary dyskinesia, and bronchiectasis.⁶⁴ The mechanism underlying bronchial hyperresponsiveness may vary between asthma, COPD, and other diseases which also have a component of BHR. A

study investigating the bronchial responsiveness profile produced by AMP, methacholine, and cold air in subjects with asthma and Sjogren's syndrome suggested that more than one challenge may be required to detect different aspects of bronchial responsiveness. Atopic asthmatic subjects were significantly more responsive to AMP than non-atopic subjects and patients with Sjogren's syndrome.⁶⁵ From a practical standpoint we speculate that AMP challenge becomes useful only when the diagnosis of asthma or COPD is clinically uncertain. However, it is clear that more population based epidemiological studies are needed to determine how valuable is adenosine responsiveness in differentiating asthma from COPD.

The view that adenosine responsiveness may be used as a specific marker of disease activity with a closer relationship to allergic airway inflammation than histamine or methacholine has been addressed in a number of clinical studies. This feature could be exploited in the clinical setting to differentiate better asthma from COPD when traditional diagnostic methods have not established a clear diagnosis. In subjects with active allergic rhinitis we have recently shown that airways responsiveness to AMP, but not methacholine, is strongly correlated to sputum eosinophilia.⁶⁶ Exhaled nitric oxide (eNO) is increasingly being used as a marker of airway inflammation and, in a study by van den Toorn *et al*, a significant correlation could be established between eNO and responsiveness to AMP, but not between eNO and responsiveness to methacholine.⁶⁷ A recent study by van den Berge and colleagues also supported previous findings that PC_{20} AMP is a better marker of airway inflammation than PC_{20} methacholine.⁶⁸ One hundred and twenty atopic asthmatics underwent bronchial provocation testing with methacholine and AMP, as well as sputum induction, blood samples, and measurement of NO in exhaled air. PC_{20} AMP provided a better reflection of airway inflammation than PC_{20} methacholine since the percentage of sputum eosinophils explained 25% of the variance in PC_{20} AMP while it was not a significant independent predictor for PC_{20} methacholine. In non-smoking patients with COPD hyperresponsiveness to AMP was also related to increased percentages of eosinophils in induced sputum and increased numbers of mucosal CD8+ cells in bronchial biopsy specimens, thereby reflecting the close association between AMP hyperresponsiveness and airway inflammation in COPD.⁶⁹

A series of clinical studies have confirmed the potential usefulness of AMP in detecting inflammatory changes in adult and paediatric asthma. Various investigations have shown a pronounced improvement in AMP responsiveness compared with methacholine or histamine after allergen avoidance, suggesting reduced airway inflammation following avoidance of aeroallergens.⁷⁰⁻⁷² Doull *et al*⁷³ have shown that regular treatment of asthmatic children with the inhaled corticosteroid beclomethasone dipropionate results in a significant reduction in AMP but not methacholine or bradykinin responsiveness. This finding confirms earlier observations that regular treatment with inhaled budesonide resulted in greater attenuation of the airway response to AMP than to methacholine.⁷⁴⁻⁷⁶ In asthma the ability of this test to discriminate changes in airway reactivity with anti-inflammatory treatment better than histamine or methacholine has also been validated with inhaled ciclesonide, mometasone, and fluticasone propionate,⁷⁷⁻⁸⁰ as well as with oral prednisolone.⁸⁰ More recently, Ketchell *et al* have reported that sensitive prediction of the AMP response to inhaled corticosteroids is already apparent as early as 48 hours⁸¹ and have reported significant attenuation of airway responsiveness to AMP within 2 hours of a single dose of fluticasone propionate.⁸² In contrast, in patients with COPD adenosine appears to be as insensitive as methacholine in detecting changes in airway reactivity after treatment with high dose inhaled steroids.⁸³ This distinctive feature is of diagnostic interest as it may indicate an additional way by which adenosine challenge may be useful in discriminating asthma from "true" COPD.

The recent work by van den Berge *et al*⁸⁴ showing greater improvement in BHR for AMP than for methacholine after treatment with corticosteroids underscores the view that airway responsiveness to AMP may also be used as a sensitive marker to monitor the effects of steroid therapy in asthma. Perhaps a limitation of this study is that airway responsiveness was recorded at a single time point after only 2 weeks of treatment with corticosteroids. Only longitudinal studies can better define a role for AMP challenge testing in the assessment of anti-inflammatory therapy in asthma. We have recently examined the time course of change in sputum cellularity and in bronchial reactivity to inhaled AMP and methacholine after administration of inhaled budesonide (800 µg/day) in 10 asthmatic patients.⁸⁵ Treatment with budesonide significantly reduced the airway responsiveness to AMP as early as by the first week of treatment, whereas changes in methacholine airway responsiveness and in sputum cellularity could be observed only by the fourth week of treatment. These findings emphasise the superior sensitivity profile of AMP challenge testing in evaluating airways response to anti-inflammatory therapy. However, this should be discussed against the evidence that other non-invasive putative markers (such as exhaled NO) may be as sensitive as AMP challenge testing in monitoring glucocorticoid responsiveness in asthma.

There is substantial evidence that adenosine is a better marker of airway allergic inflammation than the direct stimuli histamine or methacholine, but whether it is closely related to disease severity needs to be further explored. Avital *et al*⁸⁶ compared exercise, methacholine, and AMP in 135 children and young adults. They concluded that the sensitivities of AMP and methacholine challenges in the detection of bronchial hyperreactivity were very similar, but that methacholine was better at discriminating between mild and moderate asthma than AMP. This finding was also confirmed in a retrospective analysis of 487 adult asthmatic patients.⁸⁷ Methacholine and AMP challenges were compared as screening tools and any relationships between BHR and disease severity markers identified. The results suggested that methacholine was a more appropriate screening tool for BHR than AMP in their population and was related to asthma severity. From these two studies adenosine does not seem to be a good indicator of disease severity but further clinical trials are needed to confirm this. However, since airway response to direct stimuli is more strictly related to the actual degree of airway constriction than inflammation, it is not surprising that AMP does not serve as a valuable tool for monitoring disease severity.

CONCLUDING REMARKS

The mechanism of airway hyperresponsiveness to adenosine/AMP has now been largely elucidated, although some questions remain. The available evidence clearly indicates that AMP challenge has a distinctive ability to probe immunological as well as non-specific responsiveness in asthma and COPD and, in this regard, can be expected to yield important and clinically relevant results in the future. Moreover, bronchoprovocation testing with adenosine offers substantial advantages (especially in term of sensitivity) over other non-invasive tests including induced sputum. The premise for this is that adenosine elicits bronchoconstriction by stimulating the release of bronchoconstrictor mediators from cells/nerves within the airway and thus may be sensitive to the underlying inflammatory state of the airway. However, BHR to direct stimuli such as methacholine remains an exceptionally sensitive diagnostic test and, as such, it serves well to exclude disease. By contrast, because of its superior specificity, BHR to inhaled AMP may be preferred to confirm a diagnosis of asthma.

- Inhaled adenosine causes concentration related bronchoconstriction in asthma and COPD.
- Airway hyperresponsiveness to adenosine may be valuable in differentiating asthma from COPD.
- The response to adenosine is likely to assess disease activity in relation to allergic airway inflammation.
- Serial measurements of airway hyperresponsiveness to adenosine may be useful in monitoring the anti-inflammatory effects of topical steroids.

Current GINA guidelines recommend careful monitoring of asthma symptoms and pulmonary function and recognise the need for “developing noninvasive test(s) of airway inflammation for use in diagnosis, monitoring the disorder’s activity, and evaluating treatments”. Based on the emerging evidence, adenosine bronchoprovocation testing can be put forward as being useful in differentiating allergic asthma from COPD and for monitoring airway inflammatory changes in adult and paediatric asthma. In particular, serial measurements of adenosine airway responsiveness may, in future, become of increased value in monitoring anti-inflammatory effects of asthma treatment. However, well planned and well conducted large clinical trials are needed to show that information gained from this test will lead to improved patient management.

Authors’ affiliations

R Polosa, Dipartimento di Medicina Interna e Specialistica, Ospedale Tomaselli, Università di Catania, 95125 Catania, Italy
S Rorke, S T Holgate, Adult Respiratory and Molecular Sciences Research, Southampton General Hospital, Southampton SO16 6YD, UK

REFERENCES

- 1 **Stafford A**. Potentiation of adenosine and the adenine nucleotides by dipyrindamole. *Br J Pharmacol* 1966;**28**:218–27.
- 2 **Cushley MJ**, Tallant N, Holgate ST. The effect of dipyrindamole on histamine- and adenosine-induced bronchoconstriction in normal and asthmatic subjects. *Eur J Respir Dis* 1985;**67**:185–92.
- 3 **Crimi N**, Palermo F, Oliveri R, *et al*. Enhancing effect of dipyrindamole inhalation on adenosine-induced bronchospasm in asthmatic patients. *Allergy* 1988;**43**:179–83.
- 4 **Linden J**. Cloned adenosine A3 receptors: pharmacological properties, species differences and receptor functions. *Trends Pharmacol Sci* 1994;**15**:298–306.
- 5 **Fredholm BB**, Abbracchio MP, Burnstock G, *et al*. Nomenclature and classification of purinoceptors. *Pharmacol Rev* 1994;**46**:143–56.
- 6 **Cushley MJ**, Tattersfield AE, Holgate ST. Inhaled adenosine and guanosine on airway resistance in normal and asthmatic subjects. *Br J Clinical Pharmacol* 1983;**15**:161–5.
- 7 **Oosterhoff Y**, de-Jong JW, Jansen MA, *et al*. Airway responsiveness to adenosine 5'-monophosphate in chronic obstructive pulmonary disease is determined by smoking. *Am Rev Respir Dis* 1993;**147**:553–8.
- 8 **Polosa R**, Holgate ST. Adenosine bronchoprovocation: a promising marker of allergic inflammation in asthma? *Thorax* 1997;**52**:919–23.
- 9 **Feoktistov I**, Polosa R, Holgate ST, *et al*. Adenosine A2B receptors: a novel therapeutic target in asthma? *Trends Pharmacol Sci* 1998;**19**:148–53.
- 10 **Fozard JR**, Hannon JP. Adenosine receptor ligands: potential as therapeutic agents in asthma and COPD. *Pulm Pharmacol Ther* 1999;**12**:111–4.
- 11 **Coleman RA**. Effects of some purine derivatives on the guinea pig trachea and their interaction with drugs that blocks adenosine uptake. *Br J Pharmacol* 1976;**57**:51–7.
- 12 **Brown CM**, Collis MG. Evidence for an A2/Ra adenosine receptor in the guinea-pig trachea. *Br J Pharmacol* 1982;**76**:381–7.
- 13 **Advenier C**, Bidet D, Floch Saint'Aubin A, *et al*. Contribution of prostaglandins and thromboxanes to the adenosine and ATP-induced contraction of guinea-pig isolated trachea. *Br J Pharmacol* 1982;**77**:39–44.
- 14 **Finney MJ**, Karlsson JA, Persson CG. Effects of bronchoconstrictors and bronchodilators on a novel human small airway preparation. *Br J Pharmacol* 1985;**85**:29–36.
- 15 **Björck T**, Gustafsson LE, Dahlen SE. Isolated bronchi from asthmatics are hyperresponsive to adenosine, which apparently acts indirectly by liberation of leukotrienes and histamine. *Am Rev Respir Dis* 1992;**145**:1087–91.
- 16 **Mann JS**, Holgate ST, Renwick AG, *et al*. Airway effects of purine nucleosides and nucleotides and release with bronchial provocation in asthma. *J Appl Physiol* 1986;**61**:1667–76.
- 17 **Wolff J**, Londos C, Cooper DM. Adenosine receptors and the regulation of adenylate cyclase. *Adv Cyclic Nucleotide Res* 1981;**14**:199–214.

- 18 **Redington AE**, Polosa R, Walls AF, *et al*. Role of mast cells and basophils in asthma. *Chem Immunol* 1995;**62**:22–59.
- 19 **Church MK**, Levi-Schaffer F. The human mast cell. *J Allergy Clin Immunol* 1997;**99**:155–60.
- 20 **Turnbull LW**, Turnbull LS, Crofton J, *et al*. Variations in chemical mediators of hypersensitivity in the sputum of chronic bronchitis: correlation with peak expiratory flow. *Lancet* 1978;ii:184–6.
- 21 **Postma DS**, Keyzer JJ, Koeter GH, *et al*. Influence of the parasympathetic and sympathetic nervous system on nocturnal bronchial obstruction. *Clin Sci* 1985;**69**:251–8.
- 22 **Lamb D**, Lumsden A. Intra-epithelial mast cells in human airway epithelium: evidence for smoking-induced changes in their frequency. *Thorax* 1982;**37**:334–42.
- 23 **Grashoff WF**, Sont JK, Sterk PJ, *et al*. Chronic obstructive pulmonary disease: role of bronchiolar mast cells and macrophages. *Am J Pathol* 1997;**151**:1785–90.
- 24 **Pesci A**, Rossi GA, Bertorelli G, *et al*. Mast cells in the airway lumen and bronchial mucosa of patients with chronic bronchitis. *Am J Respir Crit Care Med* 1994;**149**:1311–6.
- 25 **de Boer WI**, van Schadewijk A, Sont JK, *et al*. Transforming growth factor β_1 and recruitment of macrophages and mast cells in airways in COPD. *Am J Respir Crit Care Med* 1998;**158**:1951–7.
- 26 **Gruber BL**, Marchese MJ, Kew RR. Transforming growth factor- β_1 mediates mast cell chemotaxis. *J Immunol* 1994;**152**:5860–7.
- 27 **Caughey GH**. Roles of mast cell tryptase and chymase in airway function. *Am J Physiol* 1989;**257**:L39–46.
- 28 **Sekizawa K**, Caughey GH, Lazarus SC, *et al*. Mast cell tryptase causes airway smooth muscle hyperresponsiveness. *J Clin Invest* 1989;**83**:175–9.
- 29 **Sommerhof CP**, Caughey GH, Finkbeiner WE, *et al*. Mast cell chymase: a potent secretagogue for airway gland serous cells. *J Immunol* 1989;**142**:2450–6.
- 30 **Marquardt DL**, Walker LL, Wasserman SI. Adenosine receptors on mouse bone marrow-derived mast cells: functional significance and regulation by aminophylline. *J Immunol* 1984;**133**:932–7.
- 31 **Marquardt DL**, Walker LL, Heinemann S. Cloning of two adenosine receptor subtypes from mouse bone marrow-derived mast cells. *J Immunol* 1994;**152**:4508–15.
- 32 **Church MK**, Pao GJ, Holgate ST. Characterization of histamine secretion from mechanically dispersed human mast cells: effects of anti-IgE, calcium ionophore A23187, compound 48/80, and basic polypeptides. *J Immunol* 1982;**129**:2116–21.
- 33 **Peachell PT**, Columbo M, Kagey-Sobotka A, *et al*. Adenosine potentiates mediator release from human lung mast cells. *Am Rev Respir Dis* 1988;**138**:1143–51.
- 34 **Feoktistov I**, Biaggioni I. Adenosine A2b receptors evoke interleukin-8 secretion in human mast cells. An enprofylline-sensitive mechanism with implications for asthma. *J Clin Invest* 1995;**96**:1979–86.
- 35 **Forsythe P**, McGarvey LP, Heaney LG, *et al*. Adenosine induces histamine release from human bronchoalveolar lavage mast cells. *1999*;**96**:349–55.
- 36 **Rutgers SR**, Koeter GH, Van Der Mark TW, *et al*. Protective effect of oral terfenadine and not inhaled ipratropium on adenosine 5'-monophosphate-induced bronchoconstriction in patients with COPD. *Clin Exp Allergy* 1999;**29**:1287–92.
- 37 **Rafferty P**, Beasley R, Holgate ST. The contribution of histamine to immediate bronchoconstriction provoked by inhaled allergen and adenosine 5' monophosphate in atopic asthma. *Am Rev Respir Dis* 1987;**136**:369–73.
- 38 **Phillips GD**, Rafferty P, Beasley R, *et al*. Effect of oral terfenadine on the bronchoconstrictor response to inhaled histamine and adenosine 5'-monophosphate in non-atopic asthma. *Thorax* 1987;**42**:939–45.
- 39 **Phillips GD**, Polosa R, Holgate ST. The effect of histamine-H1 receptor antagonism with terfenadine on concentration-related AMP-induced bronchoconstriction in asthma. *Clin Exp Allergy* 1989;**19**:405–9.
- 40 **Phillips GD**, Ng WH, Church MK, *et al*. The response of plasma histamine to bronchoprovocation with methacholine, adenosine 5'-monophosphate, and allergen in atopic nonasthmatic subjects. *Am Rev Respir Dis* 1990;**141**:9–13.
- 41 **Polosa R**, Ng WH, Crimi N, *et al*. Release of mast-cell-derived mediators after endobronchial adenosine challenge in asthma. *Am J Respir Crit Care Med* 1995;**151**:624–9.
- 42 **Polosa R**, Pagano C, Prosperini G, *et al*. Histamine release upon adenosine 5'-monophosphate (AMP) nasal provocation in allergic subjects. *Thorax* 1999;**54**:230–3.
- 43 **Crimi N**, Palermo F, Polosa R, *et al*. Effect of indomethacin on adenosine-induced bronchoconstriction. *J Allergy Clin Immunol* 1989;**83**:921–5.
- 44 **Phillips GD**, Holgate ST. The effect of oral terfenadine alone and in combination with flurbiprofen on the bronchoconstrictor response to inhaled adenosine 5'-monophosphate in nonatopic asthma. *Am Rev Respir Dis* 1989;**139**:463–9.
- 45 **Crimi N**, Polosa R, Magri S, *et al*. Inhaled lysine acetylsalicylate (L-ASA) attenuates the bronchoconstrictor response to adenosine 5'-monophosphate (AMP) in asthmatic subjects. *Eur Respir J* 1995;**8**:905–12.
- 46 **Van Schoor J**, Joos GF, Kips JC, *et al*. The effect of ABT-761, a novel 5-lipoxygenase inhibitor, on exercise- and adenosine-induced bronchoconstriction in asthmatic subjects. *Am J Respir Crit Care Med* 1997;**155**:875–80.
- 47 **Rorke S**, Jennison S, Jeffs JA, *et al*. The role of cysteinyl leukotrienes in adenosine 5'-monophosphate-induced bronchoconstriction in asthma. *Am J Respir Crit Care Med* 2001;**163**:A429.
- 48 **Polosa R**, Phillips GD, Rajakulasingam K, *et al*. The effect of inhaled ipratropium bromide alone and in combination with oral terfenadine on bronchoconstriction provoked by adenosine 5'-monophosphate and histamine in asthma. *J Allergy Clin Immunol* 1991;**87**:939–47.
- 49 **Polosa R**, Rajakulasingam K, Church MK, *et al*. Repeated inhalation of bradykinin attenuates adenosine 5'-monophosphate (AMP) induced bronchoconstriction in asthmatic airways. *Eur Respir J* 1992;**5**:700–6.
- 50 **Woolcock AJ**, King G. Is there a specific phenotype for asthma? *Clin Exp Allergy* 1995;**25**(suppl 2):3–7; discussion 17–8.
- 51 **Sears MR**, Jones DT, Holdaway MD, *et al*. Prevalence of bronchial reactivity to inhaled methacholine in New Zealand children. *Thorax* 1986;**41**:283–9.
- 52 **Tashkin DP**, Altose MD, Bleecker ER, *et al*. The lung health study: airway responsiveness to inhaled methacholine in smokers with mild to moderate airflow limitation. The Lung Health Study Research Group. *Am Rev Respir Dis* 1992;**145**:301–10.
- 53 **van Asperen P**, Mellis CM, South RT, *et al*. Bronchial reactivity in cystic fibrosis with normal pulmonary function. *Am J Dis Child* 1981;**135**:815–9.
- 54 **Pang J**, Chan HS, Sung JY. Prevalence of asthma, atopy, and bronchial hyperreactivity in bronchiectasis: a controlled study. *Thorax* 1989;**44**:948–51.
- 55 **Gudbjornsson B**, Hedenstrom H, Stalenheim G, *et al*. Bronchial hyperresponsiveness to methacholine in patients with primary Sjogren's syndrome. *Ann Rheum Dis* 1991;**50**:36–40.
- 56 **Potena A**, La Corte R, Fabbri LM, *et al*. Increased bronchial responsiveness in primary and secondary Sjogren's syndrome. *Eur Respir J* 1990;**3**:548–53.
- 57 **Wardlaw AJ**, Dunnette S, Gleich GJ, *et al*. Eosinophils and mast cells in bronchoalveolar lavage in subjects with mild asthma. Relationship to bronchial hyperreactivity. *Am Rev Respir Dis* 1988;**137**:62–9.
- 58 **Robinson DS**, Bentley AM, Hartnell A, *et al*. Activated memory T helper cells in bronchoalveolar lavage fluid from patients with atopic asthma: relation to asthma symptoms, lung function, and bronchial responsiveness. *Thorax* 1993;**48**:26–32.
- 59 **Casale TB**, Wood D, Richerson HB, *et al*. Elevated bronchoalveolar lavage fluid histamine levels in allergic asthmatics are associated with methacholine bronchial hyperresponsiveness. *J Clin Invest* 1987;**79**:1197–203.
- 60 **De Meer G**, Heederik DJ, Brunekreef B, *et al*. Repeatability of bronchial hyperresponsiveness to adenosine-5'-monophosphate (AMP) by a short dosimeter protocol. *Thorax* 2001;**56**:362–5.
- 61 **Hughes PJ**, Holgate ST, Church MK. Adenosine inhibits and potentiates IgE-dependent histamine release from human lung mast cells by an A2-purinoceptor mediated mechanism. *Biochem Pharmacol* 1984;**33**:3847–52.
- 62 **Hoffman HM**, Marquardt DL. The effect of adenosine on histamine release from allergen-sensitized mouse lung tissue. *J Allergy Clin Immunol* 1997;**99**(Part 2):S89.
- 63 **van Daele SG**, de Baets F, Vinaimont F, *et al*. Adenosine and histamine challenges in preschool children with recurrent wheeze. *Am J Respir Crit Care Med* 2001;**163**:A568.
- 64 **Avital A**, Springer C, Bar Yishay E, *et al*. Adenosine, methacholine, and exercise challenges in children with asthma or paediatric chronic obstructive pulmonary disease. *Thorax* 1995;**50**:511–6.
- 65 **Ludviksdottir D**, Janson C, Bjornsson E, *et al*. Different airway responsiveness profiles in atopic asthma, nonatopic asthma, and Sjogren's syndrome. BHR Study Group. *Bronchial hyperresponsiveness. Allergy* 2000;**55**:259–65.
- 66 **Polosa R**, Ciamarra I, Mangano G, *et al*. Bronchial hyperresponsiveness and airway inflammation markers in nonasthmatics with allergic rhinitis. *Eur Respir J* 2000;**15**:30–5.
- 67 **van den Toorn LM**, Prins JB, Overbeek SE, *et al*. Adolescents in clinical remission of atopic asthma have elevated exhaled nitric oxide levels and bronchial hyperresponsiveness. *Am J Respir Crit Care Med* 2000;**162**:953–7.
- 68 **van den Berge M**, Meijer RJ, Kerstjens HA, *et al*. PC20 adenosine 5'-monophosphate is more closely associated with airway inflammation in asthma than PC20 methacholine. *Am J Respir Crit Care Med* 2001;**163**:1546–50.
- 69 **Rutgers SR**, Timens W, Tzanakis N, *et al*. Airway inflammation and hyperresponsiveness to adenosine 5'-monophosphate in chronic obstructive pulmonary disease. *Clin Exp Allergy* 2000;**30**:657–62.
- 70 **van Velzen E**, van den Bos JW, Benckhuijsen JA, *et al*. Effect of allergen avoidance at high altitude on direct and indirect bronchial hyperresponsiveness and markers of inflammation in children with allergic asthma. *Thorax* 1996;**51**:582–4.
- 71 **Benckhuijsen J**, van den Bos JW, van Velzen E, *et al*. Differences in the effect of allergen avoidance on bronchial hyperresponsiveness as measured by methacholine, adenosine 5'-monophosphate, and exercise in asthmatic children. *Pediatr Pulmonol* 1996;**22**:147–53.
- 72 **Grootendorst DC**, Dahlen SE, Van Den Bos JW, *et al*. Benefits of high altitude allergen avoidance in atopic adolescents with moderate to severe asthma, over and above treatment with high dose inhaled steroids. *Clin Exp Allergy* 2001;**31**:400–8.
- 73 **Doull IJ**, Sandall D, Smith S, *et al*. Differential inhibitory effect of regular inhaled corticosteroid on airway responsiveness to adenosine 5' monophosphate, methacholine, and bradykinin in symptomatic children with recurrent wheeze. *Pediatr Pulmonol* 1997;**23**:404–11.
- 74 **Polosa R**, Rajakulasingam K, Prosperini G, *et al*. Budesonide attenuates bronchial reactivity to AMP to a greater extent than to methacholine in mild asthma. *Thorax* 1993;**48**:415.

- 75 **O'Connor BJ**, Ridge SM, Barnes PJ, *et al.* Greater effect of inhaled budesonide on adenosine 5'-monophosphate-induced than on sodium-metabisulfite-induced bronchoconstriction in asthma. *Am Rev Respir Dis* 1992;**146**:560-4.
- 76 **Wilson AM**, Lipworth BJ. Dose-response evaluation of the therapeutic index for inhaled budesonide in patients with mild-to-moderate asthma. *Am J Med* 2000;**108**:269-75.
- 77 **Taylor DA**, Jensen MW, Kanabar V, *et al.* A dose-dependent effect of the novel inhaled corticosteroid ciclesonide on airway responsiveness to adenosine-5'-monophosphate in asthmatic patients. *Am J Respir Crit Care Med* 1999;**160**:237-43.
- 78 **Holgate ST**, Arshad H, Stryczak P, *et al.* Mometasone furoate antagonizes AMP-induced bronchoconstriction in patients with mild asthma. *J Allergy Clin Immunology* 2000;**105**:906-11.
- 79 **Weersink EJ**, Douma RR, Postma DS, *et al.* Fluticasone propionate, salmeterol xinafoate, and their combination in the treatment of nocturnal asthma. *Am J Respir Crit Care Med* 1997;**155**:1241-6.
- 80 **Meijer RJ**, Kerstjens HA, Arends LR, *et al.* Effects of inhaled fluticasone and oral prednisolone on clinical and inflammatory parameters in patients with asthma. *Thorax* 1999;**54**:894-9.
- 81 **Ketchell RI**, Jensen MW, Loh LC, *et al.* High dose fluticasone propionate rapidly attenuates airway responsiveness to adenosine 5'-monophosphate in mild asthma. *Eur Respir J* 1999;**14**(suppl 30):467s.
- 82 **Ketchell RI**, Jensen MW, Lumley P, *et al.* Rapid effect of a single dose of inhaled fluticasone propionate on airway responsiveness to AMP in mild asthma. *Am J Respir Crit Care Med* 2001;**163**:A420.
- 83 **Rutgers SR**, Koeter GH, van der Mark TW, *et al.* Short-term treatment with budesonide does not improve hyperresponsiveness to adenosine 5'-monophosphate in COPD. *Am J Respir Crit Care Med* 1998;**157**:880-6.
- 84 **van den Berge M**, Kerstjens HA, Meijer RJ, *et al.* Corticosteroid-induced improvement in the PC20 of adenosine monophosphate is more closely associated with reduction in airway inflammation than improvement in the PC20 of methacholine. *Am J Respir Crit Care Med* 2001;**164**:1127-32.
- 85 **Prosperini G**, Rey J-P, Rajakulasingam K, *et al.* Effect of budesonide on AMP and methacholine airway responsiveness and sputum cellularity: a time-course study. *Eur Respir J* 2001;**18**(suppl 33):163s.
- 86 **Avital A**, Godfrey S, Springer C. Exercise, methacholine, and adenosine 5'-monophosphate challenges in children with asthma: relation to severity of the disease. *Pediatr Pulmonol* 2000;**30**:207-14.
- 87 **Fowler SJ**, Dempsey OJ, Sims EJ, *et al.* Screening for bronchial hyperresponsiveness using methacholine and adenosine monophosphate. Relationship to asthma severity and beta(2)-receptor genotype. *Am J Respir Crit Care Med* 2000;**162**:1318-22.



Have your say

eLetters

If you wish to comment on any article published in *Thorax* you can send an eLetter using the eLetters link at the beginning of each article. Your response will be posted on *Thorax* online within a few days of receipt (subject to editorial screening).

www.thoraxjnl.com