Effect of different bronchodilators on airway smooth muscle responsiveness to contractile agents

B Gustafsson, C G A Persson

Abstract

"Functional antagonism" is often used to describe the general relaxant effect of beta₂ agonists and xanthines and their ability to protect the airways against bronchoconstrictor stimuli. This study in guinea pig isolated trachea addresses the question of whether the capacity of these drugs to protect against constrictor stimuli is related to smooth muscle relaxation. Three antimuscarinic drugs were also examined to determine whether antagonism of mediators other than muscarinic agonists might contribute to bronchodilatation by these antimuscarinic drugs. Terbutaline (1.1×10^{-7} , 2.2×10^{-7} M), theophylline (2.2×10^{-4} , 4.4×10^{-4} M), and enprofylline (5.2×10^{-5} , 1.0×10^{-4} M) relaxed the tracheal tension that remained after indomethacin treatment. They did not, however, alter the carbachol concentration-response curve significantly. In addition, neither theophylline (2.2×10^{-4} M) nor terbutaline (1.1×10^{-7} M) altered histamine induced contraction. Atropine sulphate, glycopyrrolate, and ipratropium bromide had EC₅₀ values of 10^{-2} – 10^{-8} M for relaxation of carbachol induced contractions, whereas concentrations of 10^{-6} – 10^{-3} M or greater were required to relax contractions induced by allergen and nine other non-muscarinic mediators. It is suggested that bronchodilatation by antimuscarinic drugs in vivo is due to inhibition of acetylcholine induced bronchoconstriction alone and that beta₂ agonists and xanthines have poor ability to protect airway smooth muscle against constrictor stimuli. Hence mechanisms other than bronchodilatation and "functional antagonism" should be considered to explain the protection against constrictor stimuli in asthma seen with beta₂ agonists and xanthines.

Three classes of drugs, beta₂ agonists, xanthines, and antimuscarinics, are known to produce acute bronchodilatation in asthmatic patients. Pharmacological antagonism of muscarinic contraction by antimuscarinic drugs has been amply documented in vitro and in vivo,^{1,2} but the specificity of action of these drugs against non-muscarinic contractile mediators has not been assessed to any extent. The present study has examined the specificity of muscarinic antagonism by atropine

sulphate, ipratropium bromide, and glycopyrrolate in tracheal preparations in vitro against 10 different contractile mediators and allergen. Xanthines and beta₂ receptor agonists are frequently described as functional mediator antagonists because they relax airway smooth muscle irrespective of the mediator or mediators that caused contraction.³ Physiological or functional antagonism has also been the favoured explanation for the protection offered by these drugs against provoked bronchoconstriction in asthmatic subjects,^{2,4-9} but this has not yet been shown experimentally. Possibly the pharmacology of relaxation is distinct from that of anticonstrictor effects. Indeed, the protective effects of beta₂ agonists and xanthines in asthma might conceivably result from an effect on cells other than smooth muscle.^{10,11} The present study has examined the effect of xanthines and terbutaline on carbachol and histamine induced contraction of guinea pig tracheal preparations, looking at concentrations that produce pronounced relaxant effects in vitro.¹²⁻¹⁴ These drug concentrations are above (about 5–10 times) the blood concentrations seen with systemic treatment (with xanthines and terbutaline), though a similar concentration of terbutaline may be reached in airway smooth muscle tissue after inhalation.

Methods

AIRWAY SMOOTH MUSCLE PREPARATIONS

Male guinea pigs, weighing 200–300 g, were killed by a blow to the head, and the trachea was dissected out. Two adjoining cartilage rings were cut out and an open tracheal ring was prepared by cutting through the cartilage opposite the muscle. This technique preserves the epithelial lining.¹³ The presence or absence of epithelium is of little or no consequence for the development of spontaneous tension in these preparations¹³ or for determining the relaxant potencies of beta₂ agonists and xanthines.¹³

The open tracheal ring was mounted in a jacketed, temperature controlled (37°C), 25 ml organ bath containing Krebs's solution of the following composition (mM): NaCl 118.0, KCl 4.6, CaCl₂ 2.5, MgSO₄ 1.15, NaHCO₃ 24.9, KH₂PO₄ 1.15, and glucose 5.5 with a pH of 7.4 and aerated with 5% carbon dioxide in oxygen. Change in muscle tension was measured isometrically by a strain gauge transducer (Grass FT03) and recorded on a Grass polygraph (model 7D). The initial mounting tension was adjusted to around 0.5 g. Prepara-

Department of
Pharmacology,
AB Draco, Box 34,
S-221 00 Lund
B Gustafsson
C G A Persson

Department of
Clinical
Pharmacology,
University Hospital,
S-221 00 Lund, Sweden
C G A Persson

Reprint requests to:
Dr Persson

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tions were allowed to stabilise for at least 30 minutes with repeated washings. Drugs were added by injection into the organ baths.

MEASUREMENT OF DRUG EFFECTS

Theophylline, enprofylline, and terbutaline

These experiments were carried out in the presence of indomethacin (1×10^{-6} M) to reduce spontaneous tension so that near maximal to slightly supramaximal relaxant effects could be produced by the β_2 agonist and xanthines (the results are similar in the absence of indomethacin; data not shown).

A cumulative concentration-response relationship for carbachol was obtained by increasing the concentration (10^{-8} – 10^{-3} M) in the bath stepwise until no further response occurred with two consecutive concentrations. The trachea was then washed. Two carbachol concentration-response studies were performed with each preparation to confirm repeatability. It was then treated with one of the following: theophylline 2.2×10^{-4} or 4.4×10^{-4} M, enprofylline 5.2×10^{-5} or 1.0×10^{-4} M, or terbutaline 1.1×10^{-7} or 2.2×10^{-7} M (these concentrations are known to produce maximal or near maximal relaxant effects in intrinsic tension tracheal ring preparations^{12,13} and appreciable relaxation of carbachol contracted preparations¹⁴). The effect of the relaxant drug was allowed to develop fully for 10–15 minutes, before a third carbachol curve was obtained in the presence of the drug. A cumulative concentration-response relationship for histamine in the presence of indomethacin (1×10^{-6} M) was obtained in the same way. A third histamine curve was obtained in the presence of theophylline 2.2×10^{-4} M or terbutaline 1.1×10^{-7} M.

Atropine sulphate, ipratropium bromide, and glycopyrrolate

Cumulative concentration-response relationships were obtained for atropine sulphate (10^{-8} – 10^{-3} M), ipratropium bromide (10^{-8} – 10^{-4} M), and glycopyrrolate (10^{-8} – 10^{-3} M) in preparations precontracted with one of the following mediators: carbachol (6×10^{-7} M), histamine (1×10^{-6} M), 5-hydroxytryptamine (2×10^{-7} M), prostaglandin $F_{2\alpha}$ (1.8×10^{-6} M), substance P (3.7×10^{-6} M), leukotriene C_4 (1×10^{-8} M), leukotriene D_4 (1×10^{-8} M), bradykinin (1×10^{-6} M), lys-bradykinin (1×10^{-6} M), met-lys-bradykinin (1×10^{-6} M). Some tracheal preparations from guinea pigs sensitised to ovalbumin (by intraperitoneal injection of $1 \mu\text{g} + 100 \text{ mg}$ aluminium hydroxide in 0.5 ml saline 11 weeks before the experiment) were contracted with ovalbumin (1.5×10^{-9} M). We selected a concentration of constrictor drug that caused a tracheal contraction equal to that obtained with the EC_{50} dose (the concentration producing 50% of maximal response) of carbachol. Carbachol 6×10^{-7} M was initially added to each trachea to standardise the contraction.

SOURCE AND PREPARATION OF DRUGS

Indomethacin, carbamylcholine chloride (carbachol), ovalbumin grade III, 5-hydroxytryptamine (serotonin), bradykinin, lys-

bradykinin, met-lys-bradykinin, and atropine sulphate were obtained from Sigma Chemicals, USA. Terbutaline sulphate, theophylline, and enprofylline were obtained from Draco, Lund, Sweden; leukotriene C_4 and leukotriene D_4 from Miles Scientific, USA; histamine dihydrochloride from ACO, Sweden; prostaglandin $F_{2\alpha}$ (Amoglandin) from Kabi Vitrum, Sweden; substance P from Peninsula Laboratories, England; ipratropium bromide (Atrovent) from Boehringer Ingelheim, Germany; and glycopyrrolate (AHR-504) from A H Robins, England.

Indomethacin was dissolved in 5% NaHCO_3 before dilution with 0.9% NaCl. Leukotriene C_4 and D_4 were dissolved in 70% ethanol before dilution with phosphate buffer (pH 6.9). All other drugs were diluted with saline.

ANALYSIS

The maximal changes in tension induced by the xanthines, terbutaline, carbachol, and histamine were determined. Relaxation was expressed as a percentage of baseline tension and the response to each concentration of carbachol and histamine as a percentage of the maximal contraction seen in each run with either agonist. Linear regression analysis was performed on three or four points on the linear part of the concentration-response curves, and the EC_{50} value calculated. EC_{50} values were log transformed for analysis and geometric mean values are given with SEM in parentheses. A test for the parallelism of the curves was performed on the linear part of the concentration-response curves.

The relaxant response induced by each concentration of the antimuscarinic drug was expressed as a percentage of the maximal relaxation of each mediator induced contraction. Maximal relaxation was determined by adding increasing concentrations of the drug until no further relaxation occurred with two consecutive concentrations. Calculation of EC_{50} values was as above.

Values in the text represent mean (SEM) values and n refers to the number of animals (that is, preparations) studied. Analysis of variance was carried out and differences between means were examined by Student's t test for paired or unpaired observations. A p value below 0.05 was taken as significant.

Results

EFFECT OF INDOMETHACIN

Indomethacin significantly reduced spontaneous basal tension in the tracheal ring preparations before carbachol ($n = 36$) and histamine ($n = 30$) (initial tension: 10.9 (0.5) and 14.3 (0.4) mN; after indomethacin: 9.3 (0.4) and 11.4 (0.5) mN ($p < 0.001$)). The EC_{50} value for the carbachol concentration-response study in the absence of indomethacin (5.3×10^{-7} M) was less than in its presence (2.4×10^{-6} M; $p < 0.001$), but the maximally induced tension was not altered (data not shown; $p = 0.76$, $n = 72$).

Table 1 Concentrations of carbachol producing tracheal contractions equal to 50% of the maximal carbachol response (EC_{50}) after pretreatment with different drugs

	Geometric mean (SEM)	n
Control 1	2.0×10^{-6} (1.2)	57
Control 2	2.4×10^{-6} (1.1)	57
Control 3	3.1×10^{-6} (1.2)	5
Drug and concentration (M)		
Terbutaline		
1.1×10^{-7}	2.4×10^{-6} (1.2)	12
2.2×10^{-7}	5.1×10^{-6} (1.3)	6
Enprofylline		
5.2×10^{-5}	2.3×10^{-6} (1.1)	12
1.0×10^{-4}	2.6×10^{-6} (1.2)	6
Theophylline		
2.2×10^{-4}	4.1×10^{-6} (1.1)	12
4.4×10^{-4}	4.9×10^{-6} (1.1)	6

EFFECTS OF XANTHINES AND TERBUTALINE ON CARBACHOL CONTRACTION

Following indomethacin further relaxation was recorded after the addition of the xanthines and terbutaline. The percentage relaxation produced by terbutaline (1.1×10^{-7} and 2.2×10^{-7} M) was 4.9 (1.4) ($n = 12$) and 11.3 (2.8) ($n = 6$) respectively, that produced by enprofylline (5.2×10^{-5} and 1.0×10^{-4} M) 20.2 (3.3) ($n = 12$) and 15.2 (4.2) ($n = 6$), and that produced by theophylline (2.2×10^{-4} and 4.4×10^{-4} M) 19.3 (2.5) ($n = 12$) and 24.1 (6.0) ($n = 6$).

The carbachol concentration-response studies showed good repeatability (table 1). The EC_{50} values for carbachol obtained in the presence of terbutaline or the xanthines did not differ significantly from the control values (second control run of carbachol: $p > 0.05$ —table 1). Neither theophylline, enprofylline, or

CONTRACTION(%)

Figure 2 Concentration-response curves, showing arithmetic means with SEM (vertical lines), for histamine obtained from tracheal ring preparations in the presence of indomethacin and theophylline (THEO) ($n = 12$), terbutaline (TERB) ($n = 12$), and no drug (CONTROL) ($n = 12$). The top graph shows the mean curves for the first and second control histamine studies ($n = 30$) plus the third ($n = 6$). Response is expressed as percentage of maximal contraction obtained with histamine.

CONTRACTION(%)

Figure 1 Concentration-response curves for carbachol, showing arithmetic means with SEM (vertical lines), obtained from tracheal ring preparations in the presence of indomethacin and theophylline (THEO), enprofylline (ENPRO), terbutaline (TERB), and no drugs (CONTROL). Response is expressed as percentage of maximal contraction obtained with carbachol.

Table 2 Concentrations of histamine producing tracheal contractions equal to 50% of the maximal histamine response (EC_{50}) after pretreatment with different drugs

	Geometric mean (SEM)	n
Control 1	5.4×10^{-7} (1.2)	30
Control 2	1.2×10^{-6} (1.1)	30
Control 3	1.3×10^{-6} (1.3)	6
Drug and concentration value (M)		
Terbutaline		
1.1×10^{-7}	1.2×10^{-6} (1.3)	12
Theophylline		
2.2×10^{-6}	1.6×10^{-6} (1.2)	12

terbutaline produced any shift of the carbachol concentration-response line (fig 1).

The presence of xanthines or terbutaline did not reduce the maximal tension induced by carbachol. Hence the difference between the lowest (starting tension) and the highest (maximal) tension for the carbachol contraction was larger when carbachol was added to tracheal rings that had been relaxed by a bronchodilator than when it was added to the control preparations ($p < 0.001$, $n = 54$). Consequently the EC_{50} values of carbachol given in table 1 are not directly comparable. Indeed, an unchanged EC_{50} value suggests that the carbachol induced increase in tension is slightly greater in the presence of a bronchodilator than that observed with the same concentration of carbachol in the absence of the bronchodilator.

EFFECTS OF THEOPHYLLINE AND TERBUTALINE ON HISTAMINE CONTRACTION

Both terbutaline 1.1×10^{-7} M and theophylline 2.2×10^{-6} M caused relaxation of basal tension (by 6.7% (2.3%) and 23.3% (6.7%), $n = 12$). The initial histamine concentration-response line was further to the left than the second ($p < 0.001$) and the third ($p < 0.05$) lines; the latter two lines did not differ (fig 2, table 2). The second line was used as the control for subsequent comparisons. Neither theophylline nor terbutaline produced any significant shift of the histamine concentration-response line (fig 2). The mean EC_{50} values for the histamine concentration-response study in the presence of theophylline 2.2×10^{-6} M and terbutaline 1.1×10^{-7} M did not differ ($p > 0.05$) from the second control response (table 2).

The maximal tension induced by histamine was not reduced by theophylline or terbutaline ($p > 0.05$, analysis of variance). The difference between starting tension and maximal histamine induced contraction was therefore larger when histamine was added to tracheal rings that had been relaxed by terbutaline or theophylline than when it was added to control preparations ($p < 0.001$).

EFFECTS OF ATROPINE SULPHATE, IPRATROPIUM BROMIDE, AND GLYCOPYRROLATE

The tracheal ring preparations had a mean tension of 9.0 (0.2) mN before administration of drug. Addition of contractile mediators increased the tension to at least 13.7 (0.5) mN (bradykinin 1×10^{-6} M, $n = 34$) and at most 19.6 (1.3) mN (leukotriene D_4 1×10^{-8} M, $n = 23$). Allergen (ovalbumin) 1.5×10^{-9} M increased the tension to a mean value of 21.2 (1.4) mN ($n = 24$).

Atropine sulphate, ipratropium bromide, and glycopyrrolate had no effect on the spontaneous tension of untreated preparations given at concentrations below 1×10^{-4} M (concentrations above 1×10^{-4} M produced slight contractions). All three drugs caused pronounced relaxation in preparations precontracted with carbachol. The EC_{50} values for carbachol contracted trachea were more than 100 times lower than those obtained with the next most effective mediator (table 3).

Discussion

The concentration-response characteristics, including the maximally induced tension, of a muscarinic agonist, carbachol, and histamine were not affected, or were only marginally and inconsistently affected, by the presence of theophylline, enprofylline, or terbutaline. The main effect of relaxation induced by these bronchodilators was to allow carbachol and histamine to cause contraction over a wider tension range. These data suggest that xanthines and β_2 receptor agonists may not exert a prophylactic effect at the level of airway smooth muscle. These drugs may be functional antagonists when they relax contracted preparations³ but, in the concentrations studied, appear to be without functional antagonism against the initiation and development of airway contraction.

Supramaximal relaxant concentrations of β_2 agonists in the airways may protect airway smooth muscle, but whether such concentrations are obtained after inhalation of β_2 agonists remains to be shown. The concentration of these drugs is probably much larger on the mucosal surface and in the lamina propria than around the smooth muscle as this is deeper in the airway wall. Before reaching the muscle in vivo an inhaled drug has to pass an abundant and extensively perfused microvascular network located just beneath the epithelial lining.

Adenosine was not included in this study because it may not contract the preparations we used.¹³ Adenosine, however, contracts human airway smooth muscle. Theophylline acts on adenosine receptors as a specific antagonist.^{10,15}

Table 3 Concentrations of atropine sulphate, glycopyrrolate, and ipratropium bromide producing tracheal contractions equal to 50% of the maximal response (EC_{50}) (numbers of preparations in square brackets)

Mediator	EC_{50} (M, geometric mean (SEM))		
	Atropine	Glycopyrrolate	Ipratropium
Carbachol	3.8×10^{-9} (0.5) [6]	3.8×10^{-9} (0.3) [6]	1.4×10^{-9} (0.4) [6]
Histamine	8.5×10^{-7} (2.5) [8]	5.0×10^{-6} (1.2) [8]	7.2×10^{-5} (6.0) [4]
Serotonin	7.9×10^{-7} (4.0) [7]	5.0×10^{-5} (2.2) [7]	3.6×10^{-4} (3.1) [6]
Prostaglandin $F_{2\alpha}$	— [6]	$> 1 \times 10^{-3}$ [6]	3.5×10^{-2} (0.6) [8]
Substance P	— [6]	$> 1 \times 10^{-3}$ [5]	1.1×10^{-2} (1.1) [5]
Leukotriene C_4	— [8]	$> 1 \times 10^{-4}$ [7]	2.9×10^{-3} (0.4) [5]
Leukotriene D_4	— [8]	$> 1 \times 10^{-3}$ [8]	3.1×10^{-2} (0.6) [5]
Ovalbumin	— [6]	$> 1 \times 10^{-3}$ [12]	5.2×10^{-3} (1.9) [5]
Bradykinin	— [12]	$> 1 \times 10^{-3}$ [10]	3.2×10^{-3} (1.0) [9]
Lys-bradykinin	— [6]	$> 1 \times 10^{-4}$ [6]	3.6×10^{-3} (0.9) [5]
Met-lys-bradykinin	— [6]	$> 1 \times 10^{-3}$ [6]	2.5×10^{-3} (1.6) [3]

Enprofylline, in line with its general lack of adenosine antagonism, does not shift the contractile effect of adenosine in human bronchi.¹⁰ The particular ability of theophylline to exert a protective effect through adenosine antagonism may not be important. The similar clinical efficacy of enprofylline and theophylline suggests that adenosine antagonism is of little consequence in asthmatic airways.¹⁵

Interestingly, neither carbachol nor histamine concentration-response curves were shifted by the bronchodilators even though these two agents have different biochemical mechanisms of action. Muscarinic agents inhibit adenylate cyclase¹⁶ and may thus produce contractions that are particularly difficult to reverse with drugs such as β_2 agonists, which are believed to act through cyclic AMP production. Both carbachol and histamine produce concentration related increases in inositol phosphates in airway smooth muscle and bronchoconstriction by these agents may depend on hydrolysis of phosphoinositides.^{17,18} β_2 receptor agonists have been found to inhibit the inositol phosphate response to histamine¹⁷ but to leave the response to methacholine unaffected.¹⁸ The concentrations of histamine used in those biochemical studies^{17,18} (0.1 and 1 mM) were, however, much larger than those producing a maximum contractile effect in this study, and the biochemical studies were carried out in other species. Hence the present lack of interaction between terbutaline and histamine cannot easily be related to the phosphoinositide hypothesis. We cannot exclude the possibility that contractile mediators other than histamine and carbachol¹⁹ may be more sensitive to β_2 receptor stimulants and xanthines.

We selected a muscarinic agent and histamine because they are widely used for challenges to test airway responsiveness in asthma.²⁰ Single dose studies with xanthines and β_2 agonists in asthmatic subjects have shown variable bronchodilatation and protection against bronchoconstriction.^{2,4-9} No correlation has been found between bronchodilatation and protection in individual subjects. This lack of correlation is compatible with the present observations. The protective effect of xanthines and β_2 agonists may be due to actions in tissues other than airway smooth muscle. This view is supported by the finding of Britton *et al*²¹ that acute bronchodilatation (produced by an inhaled antimuscarinic drug) in itself was not associated with any protection against histamine provocation, whereas the same degree of bronchodilatation induced by an inhaled β_2 agonist caused substantial protection.²¹ The mechanism of protection with β_2 agonists and xanthines has not been explained, but both drugs have several anti-inflammatory effects that may contribute.^{10,11,22}

The tension of guinea pig tracheal preparations in vitro has not been fully explained^{3,12} but is reduced by indomethacin, a drug that inhibits the generation of arachidonate metabolites of the cyclo-oxygenase pathway.^{13,23} Indomethacin reduced spontaneous tension. The further relaxant effects of xan-

thines and terbutaline appeared to be maximal for theophylline and enprofylline and near maximal for the partial agonist terbutaline. These data are consistent with previous observations on the potency of β_2 agonists and xanthines in vitro.^{3,12-14} The EC_{50} of carbachol was increased by indomethacin. Indomethacin has been associated with reduced sensitivity to several contractile agents in guinea pig trachea^{23,24} and, less consistently, in human bronchi in vitro²⁵ and in vivo.²⁶

Ipratropium bromide, glycopyrrolate, and atropine sulphate did not interact significantly with any mediator other than the muscarinic agonist. The three antimuscarinic drugs caused substantial antagonism of carbachol with a selectivity ratio by comparison with other mediators of at least 10^2 . With ipratropium bromide the ratio was more than 10^3 . The small interactions that occurred with other mediators are probably of little interest as such concentrations are unlikely to be seen even with large inhaled doses. These data support the view that antimuscarinic drugs antagonise only acetylcholine induced bronchoconstriction in asthma.¹

The subcellular mechanisms leading to the initiation of contraction differ from those concerned in the maintenance of an established contraction.²⁷ As xanthines and β_2 agonists reduce established contractions and antimuscarinic drugs inhibit one mediator only, a search for drugs that inhibit the initiation and development of airway smooth muscle contractions may be warranted.

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