Calf blood flow and oxygen carriage after reversal of polycythaemia secondary to hypoxic lung disease PJW WALLIS, MCP APPS, AC NEWLAND, DW EMPEY From the Department of Thoracic Medicine, London Hospital, London ABSTRACT Reduction of packed cell volume has been recommended as a therapeutic procedure impatients with polycythaemia secondary to hypoxic lung disease. We have investigated the effects of

patients with polycythaemia secondary to hypoxic lung disease. We have investigated the effects of this policy on blood flow and oxygen carriage to the calf in 12 such patients. Packed cell volume was decreased from 0.61 to 0.51 (mean) by isovolaemic haemodilution on a cell separator, with significant reductions in blood viscosity at high and low shear rates. Resting calf blood flow was unchanged but peak flow during reactive hyperaemia increased by 17% and 21% one and sever days after the procedure. Oxygen carriage to the calf at rest was initially unchanged but had fallen by 20% at seven days. During reactive hyperaemia oxygen carriage was not impaired by the reduce tion in packed cell volume since the rise in blood flow offset any reduction in arterial oxygen. content. This study has shown that when blood flow is stressed during reactive hyperaemia oxygen carriage is not compromised by a therapeutic reduction in packed cell volume.

Although an appreciable proportion of patients with hypoxic lung disease develop secondary polycythaemia this response may not be advantageous. An increase in packed cell volume improves the oxygen carrying capacity of the blood but is also responsible for an exponential rise in blood viscosity.¹ Oxygen carriage to the tissues will be beneficial only if the increased arterial oxygen content is not offset by a reduction in blood flow due to the greater viscosity of polycythaemic blood. Experimental work has shown that polycythaemia decreases tissue blood flow and impairs tissue oxygenation, 2 3 indicating that an optimum packed cell volume may exist for oxygen carriage.

Patients with polycythaemia secondary to hypoxic lung disease derive useful increases in exercise tolerance after reduction of packed cell volume to approximately 0.50 by either venesection,⁵ Dextran 40 exchange transfusion,6 or more recently erythrapheresis (isovolaemic haemodilution on a cell separator)⁷—a response not attributable to a placebo The mechanisms underlying improvement are unclear, but may include greater mental alertness due to increased cerebral blood flow⁹ improvement in pulmonary dynamics. 10 11 The effects of reduction in packed cell

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volume on skeletal muscle blood flow and oxygen car riage are unknown but may also be relevant to the improvement of performance indices. We have measured calf blood flow at rest and during reactige hyperaemia in a group of patients with hypoxic lung disease before and after reversal of polycythaemia.

Methods

Twelve patients with hypoxic lung disease and segondary polycythaemia were studied (tables 1 and 2). All were free from oedema and were not suffering from pulmonary infection of from an acute exacerbation of their disease. Two patients were light smokers (five cigarettes daily); the remainder weß non-smokers. Calf blood flow was measured by venous occlusion plethysmography, a Whitney me cury in rubber strain gauge being used. 14 Flows were measured seven days and one day before and one day and seven days after reduction of packed cell volume All measurements were made with the patient lying semisupine after lying for 60 minutes with the leg raised to 45° to the horizontal. Room temperature was held constant (+ 0.5°C) during the course of the study. A light breakfast, without tea or coffee, was allowed at 0700-0800 hours and flows recorded between 1000 and 1200 hours. Methylxanthine preparations were discontinued 24 hours before study; diuretics and inhaled β_2 agonists were withheld on the morning of the investigation until after the blood flow

Table 1 An	thropometric and lung	function data	(percentages of	predicted norma	l values 12 in	parentheses)
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Patient No	Age (y)	Sex	Weight (kg)	Height (m)	Diagnosis	$FEV_1(l)$	FVC (l)	Paco ₂ (kPa)	Pao ₂ (kPa)
1	67	M	84	1.64	A	0.9 (38)	1.4 (40)	5.1	7.4
2	59	F	52	1.55	CAO	0.5 (26)	1.2 (51)	8.3	7.1
3	54	F	41	1.55	CAO	0.6 (28)	1.6 (64)	5.6	4.9
4	59	F	74	1.59	CAO	1.0 (49)	2.7 (108)	5.9	7.5
5	64	M	93	1.74	CAO	0.4 (14)	1.6 (40)	8.2	6.4
6	56	M	95	1.61	CAO	0.4 (15)	1.3 (37)	6.5	7.6
7	65	M	83	1.66	CAO + L	0.2 (8)	0.8 (22)	7.2	7.6
8	66	M	83	1.65	CAO	0.4 (16)	0.8 (23)	6.8	8.3
9	63	M	54	1.67	CAO	0.7 (26)	1.8 (49)	7.1	7.7
.0	68	M	72	1.64	CAO	0.5 (20)	1.7 (49)	6.3	7.5
Ĭ	56	M	75	1.75	CAO	0.3 (10)	1.2 (28)	10.1	4.8
2	66	M	65	1.78	CAO	0.7 (23)	2.0 (48)	7.1	6.3

A—asbestosis; CAO—chronic airflow obstruction; L—lobectomy for bronchial carcinoma; FVC—forced vital capacity; Paco₂, Pao₂—carbon dioxide and oxygen tensions.

Conversion: SI to traditional units—Blood gas tensions: 1 kPa = 7.5 mm Hg.

measurements. Other treatment, including the usual diuretic dose, was continued unaltered during the study. No oxygen was administered during the study period. Smokers were asked to refrain from smoking 24 hours before the investigation and had a carboxy-haemoglobin concentration of less than 1%.

After resting calf blood flow had been recorded peak calf flow during reactive hyperaemia was measured after four minutes of arterial occlusion at the thigh. All measurements were made with the foot isolated from the circulation by an ankle pressure cuff inflated to 250 mm Hg.

Arterial blood pressure was measured with a sphygmomanometer after the patients had lain semi-supine for 60 minutes. Venous blood was obtained without stasis and arterial blood sampled by radial artery puncture at the end of the investigation. Haemoglobin concentration (Hb) was measured in a Coulter Counter Model S plus IV. Packed cell volume was measured by centrifuging a capillary tube blood sample for 10 minutes at 1200 g and correcting for trapped plasma by labelling albumin with iodine 125.

Blood viscosity was determined in a Wells Brookfield cone and plate viscometer; carboxyhaemoglobin was measured spectrophotometrically; ¹⁵ and blood gas tensions were measured with a Radiometer ABLI analyser. Haemoglobin oxygen saturation (Sao₂) was calculated from arterial oxygen tension (Pao₂) by reference to the oxygen-haemoglobin dissociation curve ¹⁶ with corrections for shifts in the standard curve due to pH, temperature, and base excess differences. ¹⁷ Reduction of packed cell volume in similar patients has been shown not to affect the position or shape of the oxygen-haemoglobin dissociation curve (JA Wedzicha, unpublished observations). Arterial oxygen content (Cao₂,ml/dl) was calculated from the formula:

 $(1.39 \times Hb \times Sao_2) + (0.003 \times Pao_2)$, where Pao₂ is expressed in mm Hg.

Oxygen carriage to the calf was obtained from the product of calf blood flow and arterial oxygen content

Packed cell volume was lowered to about 0.50 by erythrapheresis using a Haemonetics V50 pheresis

Table 2 Haematological data before and after erythrapheresis (percentages of predicted normal values¹³ in parentheses)

Patient No	Haemoglobin (g/dl)		Packed cell volume		Blood viscosity (mPa s))	Red cell mass	Blood volume	Packed red
	Before	After	Before	After	23/s		230/s		(ml/kg) Before	(ml/kg) Before	cells removed
					Before	- After	Before	After			
1	17.0	15.4	0.56	0.50	10.5	8.5	5.9	4.6	32 (130)	64 (111)	7
2	16.0	14.6	0.58	0.48	14.2	8.3	7.7	4.8	57 (2 42)	106 (173)	10
3	18.9	14.2	0.61	0.45	13.2	6.8	7.6	4.3	41 (147)	79 (110)	12
4	18.4	15.4	0.55	0.48	10.3	5.9	6.2	3.8			7
5	20.6	16.1	0.72	0.59	18.9	9.9	9.9	5.1		_	17
6	18.2	15.4	0.59	0.49	9.2	5.7	5.6	3.9	28 (118)	55 (99)	8
7	18.3	15.6	0.59	0.49	9.2	6.0	5.6	3.6	_ (,	_	ž
8	17.4	15.8	0.55	0.50	10.0	6.3	5.9	4.2	30 (115)	56 (94)	6
9	17.8	16.3	0.63	0.53	12.0	7.0	7.7	4.8	68 (211)	121 (161)	13
10	20.2	16.4	0.61	0.49	13.0	7.4	7.2	4.7	_ (2.1.)	_ (,	15
11	18.9	16.0	0.61	0.52	9.5	5.8	5.9	3.6	49 (172)	89 (133)	ii
12	21.1	16.4	0.67	0.54	19.5	10.5	10.0	3.9	61 (197)	94 (130)	21
Mean	18.6	15.6	0.61	0.57	12.5	7.3	7.1	4.3	44 (163)	81 (125)	ĩi
SEM	0.4	0.2	0.01	0.01	1.0	0.5	0.5	0.2	4 (13)	7 (8)	i i

Conversion: SI to traditional units—Viscosity: 1 mPa s = 1 cps.

Table 3 Mean (SE) changes in body weight, blood pressure, pulmonary function, and laboratory conditions during the study

	Before	$After \\ (day + 1) n = 12$	p	$ \begin{array}{ll} After \\ (day + 7) & n = 9 \end{array} $	p	first
Body weight (kg) Mean blood pressure (mm Hg) FEV (l) FVC'(l) PacO ₂ (kPa) PaO ₂ (kPa) Room temperature (°C) Skin temperature (°C)	73 (5) 88 (4) 0.6 (0.1) 1.5 (0.2) 7.0 (0.4) 6.9 (0.3) 21.6 (0.2) 30.9 (0.3)	-0.5 (0.2) -4 (3) 0 (0.1) 0 (0.1) +0.1 (0.1) 0 (0.2) -0.1 (0.2) +0.3 (0.4)	<0.05 NS NS NS NS NS NS NS	-0.3 (0.4) -8 (4) 0 (0.1) 0 (0.1) +0.2 (0.3) +0.2 (0.2) -0.1 (0.2) +0.1 (0.4)	NS NS NS NS NS NS NS	published as 10

FVC—vital capacity; Paco₂, Pao₂—carbon dioxide and oxygen tensions. Conversion: SI traditional units—Blood gas tensions: 1 kPa = 7.5 mm Hg.

system, packed red cells (packed cell volume 0.80) being replaced with an equal volume of human plasma protein fraction BP. Red cell mass was measured by injection of ⁵¹Cr labelled autologous red blood cells. FEV₁ and forced vital capacity (FVC) were measured with a Vitalograph spirometer.

Informed written consent was obtained from each patient and the study was approved by the London Hospital ethics committee. Student's t test (paired, two tailed) was used to compare any differences between measurements made before and after erythrapheresis. Values are expressed as means with standard errors in parentheses. A p value of less than 0.05 was considered significant.

Results

Three patients were assessed only twice, before and one day after erythrapheresis, for the following reasons: diuretic dosage was inadvertently increased during the last week of the study in patient 1; patient 3 declined further investigation; and an infective exacerbation supervened during the final week of the study in patient 11. At other times during the study

the patients remained clinically stable with no change in pulmonary function, arterial blood gas tensions of systemic blood pressure. Laboratory conditions also remained constant, with no appreciable alteration room or calf skin temperature. A transient fall in body weight was observed one day after the reduction in packed cell volume (table 3).

Blood viscosity was raised in every patient before erythrapheresis and the reduction in mean packed cell volume from 0.61 (0.01) to 0.51 (0.01) lowered blood viscosity by 42% at shear rate 23/s and 39% at shear rate 230/s (-5.2 (0.6) mPas and -2.8 (0.4) mPas respectively; p < 0.001) (table 2).

CALF BLOOD FLOW (table 4)

Resting calf blood flow was unchanged one and seven days after erythrapheresis (-0.01 (0.19) and -0.09 (0.24) ml 100 ml⁻¹ min⁻¹). Peak calf blood flow during reactive hyperaemia could not be measured in patient 3 owing to discomfort from the thigh cuff. The the remaining subjects peak calf flow increased by 17% one day after erythrapheresis and 21% seven days after (+2.37 (0.56), p < 0.01; and +2.99 (1.00) ml 100 ml⁻¹ min⁻¹, p < 0.02). There was, however,

Table 4 Calf blood flow before and one and seven days after reduction in packed cell volume

Patient No	Resting flow	(ml 100 ml ⁻¹ min ⁻¹)		Peak hyperaemic flow (100 ml ⁻¹ min ⁻¹)			
	Before	Before After		Before	After		
		$\overline{Day + 1}$	Day + 7		$\overline{Day + 1}$	Day + 7	
1	2.4	0	_	7.7	+1.9	•	
2	3.5	-1.4	-1.1	13.6	-1.6	+3.4	
3	1.6	-1.1	_	_	_	+3.4 C	
4	1.9	0	+0.3	14.6	+1.2		
5	1.0	+0.2	+0.2	15.8	+4.8	+3.5	
6	4.0	+0.6	+0.9	15.7	+4.9		
ž	1.3	+0.3	+0.7	11.2	+2.1	+3.4 □	
Ŕ	2.5	+1.0	-0.4	14.3	+2.3	+6.8	
å	2.5	-0.3	-1.1	18.1	+1.2	-3.6	
10	1.8	+0.3	+0.2	19.3	+ 2.6		
11	2.0	+0.4	+ 0.2	14.6	+4.2		
12	1.3	-0.1	-0.5	10.4	+2.5	- +2.1	
12	1.3	-0.1	-0.5	10.4	+ 2.3	(U	
Mean	2.15	-0.01	-0.09	14.12	+2.37	+2.99	
SEM	0.26	0.19	0.24	1.01	0.56	1.00	
	0.20	NS	NS	1.01	< 0.01	< 0.02	
p value		140	140		₹0.01	< 0.02	

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Table 5 Oxygen carriage to the calf before and one and seven days after reduction in packed cell volume

Patient No	Resting oxyg			Peak hyperaemic oxygen carriage (ml 100 ml ⁻¹ min ⁻¹)			
	Before	After		Before	After		
		$\overline{Day + 1}$	Day + 7		$\overline{Day + 1}$	Day + 7	
1	0.49	-0.04	_	1.59	+0.21		
2	0.66	-0.31	-0.21	2.58	-0.60	+0.60	
3	0.31	-0.24	_				
4	0.43	-0.10	-0.04	3.28	-0.58	+0.21	
5	0.23	-0.05	-0.08	3.62	-0.14	+0.11	
6	0.91	-0.04	-0.03	3.59	+0.32	-0.58	
7	0.29	+0.03	+0.10	2.46	+0.19	+0.38	
8	0.55	+0.16	-0.12	3.13	+0.22	+1.14	
9	0.55	-0.10	-0.26	3.95	-0.03	-0.97	
10	0.45	-0.02	-0.04	4.79	-0.34	+0.22	
11	0.32	0	_	2.34	+0.19		
12	0.31	-0.08	-0.15	2.49	-0.02	+0.07	
Mean	0.458	-0.066	-0.092	3.075	-0.053	+0.131	
SEM	0.056	0.035	0.036	0.270	0.099	0.205	
p value		NS	< 0.05	_	NS	NS	

no correlation between the change in peak hyperaemic flow and the reduction in blood viscosity (23/s r = -0.018, 230/s r = -0.092; n = 11) or the fall in arterial oxygen content (r = -0.458; n = 11).

OXYGEN CARRIAGE TO THE CALF (table 5)

Arterial oxygen content decreased by 17% one day and 15% seven days after erythrapheresis (-3.61 (0.55) and -3.17 (0.61) ml/dl; p < 0.001). Resting oxygen carriage to the calf was unchanged one day after reduction in packed cell volume (-0.066 (0.035) ml $100 \,\mathrm{ml}^{-1} \,\mathrm{min}^{-1}$) but decreased by 20% seven days after the procedure (-0.092 (0.036) ml $100 \,\mathrm{ml}^{-1} \,\mathrm{min}^{-1}$; p < 0.05). Because there was an increase in peak calf blood flow during reactive hyperaemia, oxygen carriage to the calf during peak flow was maintained one day and seven days after erythrapheresis (-0.053 (0.099) and -0.131 (0.205) ml $100 \,\mathrm{ml}^{-1} \,\mathrm{min}^{-1}$). No ischaemic symptoms occurred in any of the patients at rest or during exercise after the procedure.

Discussion

This study shows that reducing blood viscosity by lowering packed cell volume does not impair oxygen carriage to the calf during reactive hyperaemia. At rest, however, a reduction in oxygen carriage is observed one week after the procedure. These findings conflict with the results of acute studies in animals, where reducing blood viscosity by reversing polycythaemia produces an improvement in blood flow that more than compensates for the reduction in arterial oxygen content.²³ These animal studies, however, are unlikely to represent fully the chronic condition that exists in patients with polycythaemia secondary to hypoxic lung disease, in whom compen-

satory mechanisms operate to maintain cardiac output, ¹⁸ and therefore oxygen transport, within normal limits.

Our findings are also at variance with those of Milligan et al, ¹⁹ who found a reduction in haemoglobin transport to the calf at rest and during reactive hyperaemia after venesection in patients with primary polycythaemia and polycythaemia secondary to cyanotic heart disease. This discrepancy might result from the use of venesection by Milligan et al, a technique that produces an immediate fall in both blood volume and cardiac output.²⁰ By replacing packed red cells with an equal volume of plasma protein fraction, erythrapheresis produces a more gradual reduction in blood volume, thus avoiding any sudden fall in cardiac output immediately after the procedure.²¹

Of interest is our finding that, despite the considerable reduction in blood viscosity at both high and low shear rates, no increase in resting blood flow to the calf occurred and only a relatively small increase in peak calf blood flow was observed during reactive hyperaemia. Packed cell volume is certainly the most important determinant of blood viscosity²² but its effect in vivo is not as marked as in vitro^{23 24} owing to the reduced viscosity of blood in small vessels.²⁵ This may account for the present observations since measurements of blood viscosity were made in vitro.

Our record of calf blood flow using the Whitney strain gauge plethysmograph of course provides no information about the nature of blood flow within the microcirculation of the calf. After reduction of packed cell volume the improved rheological properties of the blood might enhance skeletal muscle microperfusion by increasing capillary flow and by recruitment of previously non-perfused capillaries as well as by reducing both plasma skimming and arteriovenous shunting of red cells. This could increase

oxygen availability within calf muscle even though the amount of oxygen carried to the calf does not rise. Alternatively, by increasing blood flow to the calf while maintaining oxygen carriage, reduction of packed cell volume might facilitate the removal of lactic acid, thereby enabling exercise tolerance to increase.

Although we and others have recommended that blood viscosity should be decreased by reversing polycythaemia in patients with hypoxic lung disease, it is important to ensure that any reduction in the oxygen carrying capacity of the blood does not impair tissue oxygenation. In this study the fall in oxygen carriage to the calf at rest, one week after erythrapheresis, did not prove deleterious. Furthermore, we have shown that when blood flow to the calf is stressed during reactive hyperaemia, as it is during exercise, oxygen carriage is not compromised by a reduction in packed cell volume to about 0.50.

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