Figure 1 Histological features of (a) Merkel cell carcinoma (MCC; H&E staining, $\times 400$) and (b) small cell lung cancer (SCLC; H&E staining, $\times 400$). Both tumours are composed of highly mitotic, small basophilic tumour cells. (c) Electropherogram of a 1.5% agarose gel for MCPyV in eight patients with SCLC (three samples from patient 3) including a positive (+), and negative control (-) and an *HaeIII* size ladder (L). (d) Corresponding Southern blot. Samples 3b and 3c display specific PCR products, confirmed by Southern blot analysis.

completely. Rather we assume that a MCPyV prevalence of 7.5% represents the MCPyV prevalence in the general population. We found almost identical numbers for the MCPyV prevalence in non-MCC tumours of sun-exposed skin.⁷ More data will be needed to prove this assumption. Nevertheless, the morphological similarities of both tumours are striking and will keep the intriguing idea of a possibly (polyoma-) virus-associated pathogenesis in mind.

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REFERENCES

1. Bichakjian CK, Lowe L, Lao CD, *et al*. Merkel cell carcinoma: critical review with guidelines for multidisciplinary management. *Cancer* 2007;110:1–12.
2. Jemal A, Siegel R, Ward E, *et al*. Cancer statistics, 2008. *CA Cancer J Clin* 2008;58:71–96.
3. Rosti G, Bevilacqua G, Bidoli P, *et al*. Small cell lung cancer. *Ann Oncol* 2006;17(Suppl 2):ii5–10.
4. Junker K, Petersen I. [Small cell lung cancer: pathology and molecular pathology]. *Pathologe* 2009;30:131–40.
5. Feng H, Shuda M, Chang Y, *et al*. Clonal integration of a polyomavirus in human Merkel cell carcinoma. *Science* 2008;319:1096–100.
6. Kassem A, Schopflin A, Diaz C, *et al*. Frequent detection of Merkel cell polyomavirus in human Merkel cell carcinomas and identification of a unique deletion in the VP1 gene. *Cancer Res* 2008;68:5009–13.
7. Andres C, Belloni B, Puchta U, *et al*. Prevalence of MCPyV in Merkel cell carcinoma (MCC) and non-MCC tumors. *J Cutan Pathol* 2009;in press.
8. Wetzel CT, Hoefnagel JG, Bakkers JM, *et al*. Ultrastructural proof of polyomavirus in merkel cell carcinoma tumour cells and its absence in small cell carcinoma of the lung. *PLoS ONE* 2009;4:e4958.

Assessment of gender differences in health status with the Leicester Cough Questionnaire (LCQ)

Kelsall *et al* were surprised that there were no differences in Leicester Cough Questionnaire (LCQ)¹ scores in men and women with chronic cough and suggest that this was because the LCQ did not capture gender-specific differences in health status.² We disagree. We have previously reported significantly worse health status in a larger group of females with chronic cough (see table 1).^{3,4} Furthermore, Polley *et al* investigated the gender differences in health status in 147 patients with chronic cough using the LCQ and another health status measure (Cough specific quality of life questionnaire) and found a clinically significant impairment in health status in females with both questionnaires.⁵ So why did Kelsall and colleagues not detect gender differences in health status? The lack of a significant difference in day-time cough frequency between genders may have been one factor. Health status questionnaires tend to focus more on the adverse impact of cough during the day. Interestingly, female patients in our study had significantly longer duration of cough than males; this may have contributed to the gender differences (table 1). We have previously reported a poor to moderate relationship between cough symptoms, health status, cough reflex sensitivity and cough frequency.⁶ Cough frequency is just one of several factors likely to be important in determining the well-being of patients. This highlights the importance of using a combination of subjective and objective tools such as automated cough monitors to assess cough severity.⁷

Table 1 Gender differences in health status in patients with chronic cough assessed with the Leicester Cough Questionnaire (LCQ)

	Females	Males	p Value (t test)
Number (%)	112 (63)	66 (27)	
Age, years (SEM)	56 (1)	54 (2)	0.5
Cough duration, years (SEM)	5.5 (0.9)	3.3 (0.6)	0.04
LCQ total (range 3–21) (SEM)	13.5 (0.4)	14.9 (0.5)	0.01

Health status questionnaires are designed to quantify quality of life numerically using the least number of questions. They are not a substitute for taking a good history. During the validation of the LCQ, health-related issues particularly pertinent to females were evaluated.¹ Several items, including stress incontinence, were excluded because only a minority of patients reported them. There was also evidence that alterations in health status due to symptoms such as stress incontinence were adequately captured by items that relate particularly to the psychosocial impact. It is important to note that health status questionnaires should be designed for use in the wider population rather than targeted to a specific subset of patients.

In conclusion, the LCQ is a brief, well-validated and widely used health status questionnaire for patients with cough and can be used to detect gender differences.

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REFERENCES

1. **Birring SS**, Prudon B, Carr AJ, *et al.* Development of a symptom specific health status measure for patients with chronic cough: Leicester Cough Questionnaire (LCQ). *Thorax* 2003;**58**:339–43.
2. **Kelsall A**, Decalmer S, McGuinness K, *et al.* Sex differences and predictors of objective cough frequency in chronic cough. *Thorax* 2009;**64**:393–8.
3. **Birring SS**, Patel RB, Prudon B, *et al.* Quality of life in chronic cough. *Am J Respir Crit Care Med* 2003;**167**:A135.
4. **Raj AA**, Pavord DI, Birring SS. Clinical cough IV: what is the minimal important difference for the Leicester Cough Questionnaire? *Handb Exp Pharmacol* 2009;(187):311–20.
5. **Polley L**, Yaman N, Heaney L, *et al.* Impact of cough across different chronic respiratory diseases: comparison of two cough-specific health-related quality of life questionnaires. *Chest* 2008;**134**:295–302.
6. **Birring SS**, Matos S, Patel RB, *et al.* Cough frequency, cough sensitivity and health status in patients with chronic cough. *Respir Med* 2006;**100**:1105–9.

7. **Birring SS**, Fleming T, Matos S, *et al.* The Leicester Cough Monitor: preliminary validation of an automated cough detection system in chronic cough. *Eur Respir J* 2008;**31**:1013–8.

Airway epithelial cells as guardians of immune homeostasis?

We read with interest the paper by Wang *et al* and accompanying editorial by Smyth showing that healthy murine airway epithelial cells (AECs) are potent inhibitors of dendritic cell (DC)-induced T cell activation.^{1,2} AECs infected with respiratory syncytial virus (RSV) lost this regulatory function, allowing activation of T cell responses and airway inflammation.¹ These in vitro observations match with the high concentrations of pro-inflammatory mediators and cells found clinically in the bronchoalveolar lavage fluid of infants with RSV bronchiolitis.²

The paper by Wang *et al* adds to a growing body of evidence that AECs are involved in maintaining airway immune homeostasis.³ Mayer *et al* previously showed that primary murine and immortalised human AECs induce an anti-inflammatory microenvironment inhibiting DC maturation and reducing T cell proliferation through constitutive secretion of transforming growth factor- β .³ Wang *et al* comment that further studies in human primary AECs are required to validate the findings in a clinical setting. Smyth also highlights the importance of research to investigate the function of AECs in health.²

Primary AECs cultured from protocol bronchoscopic brushings taken from clinically stable lung allograft recipients free from chronic allograft dysfunction represent a useful model to study AEC function in a healthy, steady state, albeit alloimmune environment. In a recent paper we have shown that epithelial cell-conditioned medium from stable lung allografts drives the production of macrophage-like cells from monocytes rather than DCs.⁴ It is unclear whether this effect only occurs in the airway of lung transplant recipients or if it reflects a general role for AECs in the homeostasis of DC populations in the lung. Nonetheless, our findings provide complementary human evidence to the murine observations of Wang *et al* and indicate that, in a steady state, AECs may be important in local immune homeostasis and promote an anti-inflammatory and pro-phagocytic airway milieu.

An emerging hypothesis that encompasses these observations is therefore that, in the healthy state, AECs regulate local immune homeostasis in the epithelium and promote anti-inflammatory conditions in the airway. In response to epithelial damage such as RSV infection, danger signals are released into the microenvironment by AECs which drive the production and maturation of professional antigen-presenting DCs, promoting T cell activation and airway inflammation.⁵

To explore this hypothesis more fully we suggest that future studies should include primary human tissue in both health and disease, and that this strategy can complement and extend animal studies.

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REFERENCES

1. **Wang H**, Su Z, Schwarze J. Healthy but not RSV-infected lung epithelial cells profoundly inhibit T cell activation. *Thorax* 2009;**64**:283–90.
2. **Smyth RL**. The airway epithelium in health and disease: "calm on the surface, paddling furiously underneath". *Thorax* 2009;**64**:277–8.
3. **Mayer AK**, Bartz H, Fey F, *et al.* Airway epithelial cells modify immune responses by inducing an anti-inflammatory microenvironment. *Eur J Immunol* 2008;**38**:1689–99.
4. **Ward C**, Eger K, Diboll J, *et al.* Bronchial epithelial cells cultured from clinically stable lung allograft patients promote the development of macrophages from monocytes rather than dendritic cells. *Thorax* 2009;**64**:430–5.
5. **Matzinger P**. Friendly and dangerous signals: is the tissue in control? *Nat Immunol* 2007;**8**:11–3.

Are patients on treatment for pulmonary TB who stop expectorating sputum genuinely culture negative?

In patients receiving treatment for pulmonary tuberculosis (TB), change in sputum culture from positive to negative is the principal outcome measure of a therapeutic response in both clinical practice^{1,2} and drug trials.³ Patients will often stop producing sputum early in the course of treatment.⁴ We have tested the assumption that "no sputum" means that the patient is "culture negative", as this has never been confirmed experimentally.

We prospectively followed 16 patients with newly diagnosed pulmonary TB. Sputum samples were collected at diagnosis, during weeks 1 and 2, at months 1, 2 and 4, and on completing treatment. Those patients who were not producing sputum spontaneously had specimens collected by induction. This was performed in a purpose-built negative pressure isolation chamber (Elwyn E Roberts Isolators, Shropshire, UK) where patients inhaled 3% hypertonic saline via an ultrasonic nebuliser for 20 min. Samples were homogenised with Sputasol (Oxoid, Basingstoke, UK). A dilution series