ASTHMA

Bronchodilation by an inhaled VPAC₂ receptor agonist in patients with stable asthma

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Background: The synthetic vasoactive intestinal peptide (VIP) analogue Ro 25-1553 is a selective VIP-PACAP type 2 (VPAC₂) receptor agonist that causes a bronchodilatory effect in guinea pigs in vivo. The effect of Ro 25-1553 given by inhalation to patients with asthma was studied and compared with that of a long acting β_2 adrenoceptor agonist.

Methods: Twenty four patients with moderate stable asthma participated in a double blind, randomised, placebo controlled, crossover study. The primary variable was bronchodilatory effect (increase in forced expiratory volume in 1 second, FEV₁) after inhalation of Ro 25-1553 (100 μ g or 600 μ g) and formoterol (4.5 μ g), respectively. Putative side effects were characterised by monitoring sitting blood pressure, serum potassium, electrocardiography and echocardiography.

Results: Inhalation of 600 μ g Ro 25-1553 caused a rapid bronchodilatory effect (geometric mean increase in FEV₁ compared with placebo) within 3 minutes of 6% (95% CI 4 to 9), as did inhalation of formoterol (8% (95% CI 5 to 10)). The corresponding maximum bronchodilatory effect during 24 hours was similar for 600 μ g Ro 25-1553 (7% (95% CI 4 to 10)) and the reference bronchodilator formoterol (10% (95% CI 7 to 12)). However, for both doses of Ro 25-1553 the bronchodilatory effect was attenuated 5 hours after inhalation whereas formoterol still had a bronchodilatory effect 12 hours after inhalation. Neither Ro 25-1553 nor formoterol produced any clinically relevant side effects. No drug related difference in adverse events was observed.

Conclusion: Inhalation of a synthetic selective VPAC₂ receptor agonist constitutes a promising approach for bronchodilation in patients with asthma.

The endogenous neuropeptide vasoactive intestinal peptide (VIP) inhibits smooth muscle tone in guinea pig airways, both when administered in vitro and when given by inhalation in vivo.¹⁻⁵ As shown in guinea pig airways in vitro, this effect is mediated via VIP-PACAP (VPAC) receptors because VIP and pituitary adenylate cyclase activating peptide (PACAP) display a similar potency.^{6,7} The activation of these VPAC receptors is followed by a subsequent increase in intracellular cyclic AMP.⁸ Based on its effects in guinea pig airways, it has been suggested that VIP possesses potential as a bronchodilator and, indeed, it has been shown that VIP can inhibit histamine induced smooth muscle tone in human bronchi in vitro.⁹ It has also been shown that VIP can cause a bronchodilatory effect when it is administered intravenously in asthmatic subjects, but not in healthy volunteers.^{10,11} In spite of these findings, the bronchodilator potential of VIP has been questioned mainly for two reasons. Firstly, it has been shown that inhaled VIP is degraded by endogenous proteases such as neutral endopeptidase (NEP) present on bronchial epithelial cells as well as chymase and tryptase derived from mast cells.¹²⁻¹⁹ Secondly, intravenous administration of VIP has been shown to cause cardiovascular side effects in asthmatic and healthy subjects.^{10,11} To reduce systemic side effects a VIP-like bronchodilator should probably be given by inhalation, but must then be resistant to degradation by proteases.

A synthetic VIP analogue Ro 25-1553 has recently been developed by Bolin and coworkers.²⁰ This selective VPAC₂ receptor agonist is more potent than original VIP and the β_2 adrenoceptor agonist salbutamol in inhibiting histamine induced airway smooth muscle tone in guinea pig trachea and in human bronchi in vitro and when given by inhalation in guinea pigs in vivo.^{9,12,21,22} In vivo studies in guinea pigs have shown that the duration of the bronchodilatory effect tends to be longer for Ro 25-1553 than for salbutamol.⁹ In line with

this, Ro 25-1553 displays a duration of action similar to that of formoterol in the guinea pig trachea in vitro.²³

We have determined whether inhalation of the selective VPAC₂ receptor agonist Ro 25-1553 causes a bronchodilatory effect in patients with moderate stable asthma using the long acting β_2 adrenoceptor agonist formoterol as a reference bronchodilator.^{24,25}

METHODS

Patients with asthma

Patients were recruited from patient files within the three centres involved in the study. The inclusion criteria were male sex, outpatient care, age 18-60 years of age, and a history of stable asthma as defined by the American Thoracic Society.²⁶ Patients had to have a baseline FEV₁ of $\geq 60\%$ predicted and $<100\%$ predicted, and FEV₁ reversibility of $\geq 15\%$. Only patients who, apart from their asthma, were judged healthy by physical examination, vital signs, electrocardiography and clinical laboratory tests were included. All included patients were able to use the inhalation devices. The exclusion criteria for the study were oral glucocorticoid treatment, β adrenoceptor blocker treatment, or a respiratory tract infection during the 4 weeks before the study, scheduled inpatient surgery during the study, hypersensitivity to β adrenoceptor agonists or lactose, tobacco smoking during the 6 months before the study, abnormal echocardiogram, blood donation during the 3 months before the study, known alcohol or other drug abuse, previous randomisation in the study, or participation in another clinical study within 4 weeks of the current one. Inhaled long acting β_2 adrenoceptor agonists were not allowed during the study and had to be withdrawn at least 72 hours before visit 1.

The study was approved by the independent ethics committee at the Medical Faculties of Göteborg, Lund and Uppsala University, respectively, and was conducted in accordance with the Helsinki Declaration. All patients gave oral and written informed consent before participating in the study. Good clinical practice procedures were applied and the study was monitored by the sponsor (AstraZeneca R&D, Lund, Sweden).

Protocol

This was a placebo controlled study with a double blind, cross-over, double dummy design, comprising a total of six study visits. Visit 1 constituted a screening visit for informed consent and enrolment, as well as standard physical examination, lung function and reversibility testing. All patients underwent supervised practice with the inhalation devices (specified below) at visit 1. Provided that the dose was kept constant for at least 4 weeks before the study, inhaled and nasal glucocorticoids, disodium cromoglycate and antihistamines were allowed during the study. Inhalation of short acting β_2 adrenoceptor agonists and/or ipratropium bromide was allowed during the study, provided that dosing did not occur within 6 and 8 hours, respectively, before the administration of randomised treatment. Visit 2 constituted the first visit for randomised treatment and took place 1–14 days after visit 1. Visits 3–5 were also treatment visits and took place 3–14 days after the preceding visit. After receiving the randomised treatment in the morning at visits 2–5, the patient remained at the clinic for 12 hours and returned to the clinic for a 24 hour assessment during the following morning. Visit 6 was a follow up visit which took place 3–5 days later and included a physical examination.

Lung function

Forced expiratory volume in 1 second (FEV_1) was the primary variable of the study and all other variables were regarded as secondary. FEV_1 was assessed using a standard spirometer (Vitalograph Alpha, Vitalograph Ltd, Buckingham, UK) at visits 1–5. The reversibility test at visit 1 was conducted before and 15 minutes after one inhalation of terbutaline 0.5 mg (Bricanyl Turbuhaler, Astra Pharmaceutical Production, Sweden). For each FEV_1 recording during visits 2–5, the best of three FEV_1 measurements was used for evaluation. Throughout the study the FEV_1 had to be within $\pm 12\%$ of the baseline FEV_1 at visit 1. Patients were rescheduled up to two times if this criterion was not fulfilled. At visits 2–5 FEV_1 was assessed 15 minutes before and 3, 15, 30, 45, 60, 90 minutes and hourly for up to 12 hours after treatment. A final FEV_1 assessment was performed 24 hours after treatment.

Vital signs

The pulse rate, systolic and diastolic blood pressure assessments were performed at the same time points as FEV_1 (see above). In addition, sitting blood pressure was assessed before and 30, 60 and 90 minutes and hourly for up to 12 hours after treatment.

Electrocardiography (ECG)

The following ECG parameters were collected at visits 1 and 6 as well as before and 60 minutes, 12 and 24 hours after each randomised treatment at visits 2–5: conduction, extra systoles, heart rate, QRS duration, QT interval, PR interval, sinus rhythm, and ST-T changes. These parameters, plus QTc and QTc dispersion, were finally evaluated after the completion of the study.

Echocardiography

Two dimensional echocardiography was carried out in accordance with standard clinical procedures by a skilled investigator for the evaluation of pericardial fluid (normal or abnormal) at visits 1 and 6 as well as before and 24 hours after

each randomised treatment at visits 2–5. This was done because a preliminary safety study on primates (*Cynomolgus*), conducted by other investigators, indicated an increased amount of pericardial fluid in some subjects after inhalation of Ro 25-1553 (unpublished data).

Clinical laboratory tests

Blood samples for control of serum potassium levels were collected at visits 1 and 6, as well as before and 12 hours after each randomised treatment at visits 2–5. Blood haemoglobin, haematocrit, red blood cells, platelets, white blood cells, differential blood cell count, serum levels of calcium, ASAT, ALAT, ALP, GT, bilirubin (total), albumin, sodium, creatinine, urea, glucose, chloride, and C reactive protein, and urinary protein, haemoglobin, erythrocytes, and glucose were also measured at these time points. All these samples were analysed using methods of standard clinical practice (Quintiles AB, Uppsala).

Adverse events

The adverse event profile was evaluated by means of open standardised questions at visits 2–6 and, when needed, by additional physical examination.

Study drugs

The study drugs were delivered at visits 2–5. The nebuliser solution of the synthetic VIP analogue Ro 25-1553 (trifluoroacetate salt) was manufactured by Pharmaceutical and Analytical R&D, AstraZeneca R&D, Lund, Sweden and was delivered for inhalation as an aerosol (100 μ g or 600 μ g delivered dose free peptide) using a nebuliser (HaloLite, Medic-Aid Ltd, UK). The corresponding placebo aerosol (2 ml of 9 mg/ml saline) was delivered using exactly the same conditions. The reference bronchodilator formoterol fumarate dihydrate was manufactured by Astra Pharmaceutical Production, Sweden and was delivered as inhalation powder (4.5 μ g delivered dose) using a commercially available inhaler (Oxis Turbuhaler). The corresponding placebo powder (lactose) was delivered using exactly the same conditions.

Statistical analysis

The primary variable of this study was maximum change in FEV_1 after each treatment. In addition, FEV_1 at 3 minutes and 12 hours after treatment was analysed. The effect of each treatment on FEV_1 was compared using a multiplicative (log transformation of the dependent variable and covariate) analysis of variance model with patient, period, and treatment as fixed factors and the pretreatment FEV_1 as a covariate. From this model, geometric mean treatment differences and corresponding 95% confidence intervals (CI) were calculated. A significance level of 5% was used—that is, differences were considered statistically significant when the 95% CI did not include a 0% increase.

The sample size was based on previous single dose studies with β_2 agonists where the within patient coefficient of variation in 12 hour mean FEV_1 had usually been around 6%. Assuming a similar variability in this study, 24 completed patients would give 80% power to detect a true increase of 5%. This assumed a significance level of 5% and a two sided alternative hypothesis.

RESULTS

Patients

Twenty four men (22 white and two of mixed race) with asthma were initially included in the study. However, one patient discontinued the study at day 10 because of exacerbation of asthma during the washout period after receiving only placebo (randomised treatment) and was not included in the analyses. Another patient received the incorrect treatment by mistake at two visits. Data from these two visits were excluded from analyses. The mean (SD) characteristics of the 23

Figure 1 Forced expiratory volume in 1 second (FEV₁, % of baseline before treatment) in patients with stable asthma after inhalation of the VPAC₂ receptor agonist Ro 25-1553 (open triangles, 100 µg; closed triangles, 600 µg), the β_2 adrenoceptor agonist formoterol 4.5 µg (closed circles) and placebo (open circles). Data are presented as geometric mean. The maximum between-patient coefficient of variation (CV) was 10.6%; n=22–23.

patients included in the analyses at visit 1 were: FEV₁ 3.38 (0.47) l/s corresponding to 78 (7)% of predicted; reversibility 18.7 (3.9)% of FEV₁; age 32 (9) years; weight 83 (11) kg; height 182 (8) cm; body mass index 25.1 (3.2) kg/m²; duration of asthma 18.8 (12.0) years. Fourteen of the 23 patients were receiving regular treatment with inhaled glucocorticoids.

Lung function

The mean (SD) baseline FEV₁ was 3.40 (0.56) l/s for 100 µg Ro 25-1553, 3.39 (0.51) l/s for 600 µg Ro 25-1553, 3.40 (0.55) l/s for formoterol (4.5 µg), and 3.37 (0.47) l/s for placebo; there were therefore no substantial differences in this parameter before each treatment.

During the 24 hours of lung function monitoring after treatment, the geometric mean maximum increase in FEV₁ was 16.2% for 100 µg Ro 25-1553, 18.2% for 600 µg Ro 25-1553, 21.5% for formoterol (4.5 µg), and 10.5% for placebo (as a percentage of baseline before treatment, n=22–23; see fig 1 for maximum CV). Compared with placebo, these differences were statistically significant (geometric mean (95% CI) difference: 5.1% (95% CI 2.8 to 7.6) for 100 µg Ro 25-1553, 7.0% (95% CI 4.5 to 9.5) for 600 µg Ro 25-1553, and 10.0% (95% CI 7.4 to 12.5) for formoterol 4.5 µg. There was no statistically significant difference between 600 µg and 100 µg Ro 25-1553 (geometric mean difference 1.7% (95% CI –0.6 to

4.1)). Formoterol 4.5 µg, however, produced a statistically significantly greater increase than 600 µg Ro 25-1553 (geometric mean difference 2.8% (95% CI 0.4 to 5.3)).

The time course of change in FEV₁ during the first 6 hours after each treatment is shown in fig 1. Three minutes after inhalation Ro 25-1553 (100 µg and 600 µg) as well as formoterol (4.5 µg) had increased FEV₁ more than placebo. At this time point the geometric mean difference (as a percentage of baseline before treatment) in FEV₁ was 4.4% (95% CI 1.9 to 7.0) for 100 µg Ro 25-1553, 6.5% (95% CI 3.8 to 9.2) for 600 µg Ro 25-1553, and 7.8% (95% CI 5.1 to 10.5) for formoterol (4.5 µg) compared with placebo. Five hours after treatment the increasing effect on FEV₁ of each dose of Ro 25-1553 was attenuated compared with placebo (fig 1). In contrast, the increasing effect of formoterol on FEV₁ remained at 12 hours after treatment (9.7% v 0.3% for placebo); this difference was statistically significant (geometric mean difference 9.4% (95% CI 4.6 to 14.3)). Neither dose of Ro 25-1553 produced any increasing effect on FEV₁ at 24 hours after treatment (100 µg Ro 25-1553 1.1%; 600 µg Ro 25-1553 –4.1 %; 4.5 µg formoterol 5.1%) compared with placebo (2.3%).

Vital signs

Overall, the maximum effect on systolic and diastolic blood pressure was small for all treatments (table 1). Similarly, the effect on pulse rate and sitting blood pressure of each respective treatment was small (data not shown).

Electrocardiography

The maximum effect of each treatment on heart rate and QTc was modest (table 1). Similarly, we were unable to detect any pronounced effect on either conduction, QRS duration, QT interval, PR interval, sinus rhythm, or ST-T changes (data not shown).

Echocardiography

The amount of pericardial fluid was within normal limits for all subjects before and 24 hours after each inhaled treatment (data not shown).

Clinical laboratory tests

The maximum effect of each treatment on serum potassium levels was small (table 1). Only very small changes or no change was seen in the other clinical laboratory tests during each treatment (data not shown).

Adverse events

There was no treatment related difference in reported adverse events (data not shown). No severe adverse events were reported throughout the course of the study.

Table 1 Mean (SD) changes in systolic blood pressure, diastolic blood pressure, heart rate, QTc, and serum potassium levels 12 hours after inhalation of placebo, 100 µg Ro 25-1553 (Ro 100), 600 µg Ro 25-1553 (Ro 600), or 4.5 µg formoterol (Fo 4.5); n=22–23

	Placebo	Ro 100	Ro 600	Fo 4.5
Systolic blood pressure (mm Hg)	16.2 (11.0)	15.9 (9.4)	14.4 (8.8)	18.7 (12.1)
Diastolic blood pressure (mm Hg)	–7.5 (4.6)	–7.3 (6.2)	–11.6 (4.9)	–7.1 (5.2)
Heart rate (beats/min)	–0.4 (6.5)	0.2 (6.6)	2.9 (7.2)	0.9 (6.0)
QTc (ms)	2.7 (18.1)	5.2 (21.2)	5.3 (14.7)	5.8 (21.7)
Serum potassium (mmol/l)	0.00 (0.33)	0.03 (0.30)	–0.07 (0.34)	–0.01 (0.34)

Values sampled as followed: blood pressure sitting (repeatedly during 24 hours, see Methods), heart rate and QTc (from ECG conducted at 1, 12 and 24 hours), serum potassium (at 12 hours).

DISCUSSION

Using the synthetic VIP analogue Ro 25-1553, this study is the first to show that inhalation of a selective VPAC₂ receptor agonist can cause a clinically relevant bronchodilatory effect in patients with moderate stable asthma.

Ro 25-1553 was selected as a potential bronchodilator because of its advantageous combination of an aromatic amino acid sequence in the N-terminal domain, C-terminal helix, and its increased resistance to proteolytic degradation at key amino acid sites.^{12 20 27 28} In contrast to original VIP, the Ro 25-1553 molecule with its lactam ring inhibits airway smooth muscle tone not only in naive but also in inflamed airways. This has previously been shown by its effect on allergen induced smooth muscle tone in sensitised guinea pig airways in vivo, more recently in vitro in human bronchi from patients undergoing surgery for lung cancer (of whom the majority were tobacco smokers), and is now shown in patients with stable asthma.^{20 28 29} It is likely that the clearcut bronchodilatory effect in the airways of patients with asthma is facilitated by the ability of Ro 25-1553 to inhibit airway smooth muscle contraction caused by several pathogenetically relevant stimuli, as has been demonstrated in vitro and in vivo including histamine, leukotriene D₄, and carbachol.⁹ The fact that the effect of Ro 25-1553 has been virtually unaffected by the β adrenoceptor blocker propranolol in tracheal smooth muscle from guinea pigs in vitro indicates a mechanism that is independent of the β_2 adrenoceptor.⁹

When assessing the pharmacotherapeutic potential of Ro 25-1553, it should be emphasised that our study shows a longer duration of action for the reference bronchodilator formoterol than for Ro 25-1553 in the airways of patients with moderate stable asthma. This illustrates the limitation of the guinea pig airway model in which Ro 25-1553 was found to have a relatively long duration of action.^{9 23} Because of its limited duration of action, Ro 25-1553 will probably not be established as a bronchodilator for clinical use in asthma. Rather, the current results should prompt the development of new selective VPAC₂ receptor agonists with an increased duration of action. Such synthetic analogues of VIP may provide a useful complement to β_2 adrenoceptor agonists in the treatment of asthma.

Our study shows that inhalation of Ro 25-1553 at doses causing a bronchodilatory effect does not lead to any clinically substantial side effects on the cardiovascular system. No pronounced effects were seen on either blood pressure, heart rate, QTc, or pericardial fluid. Furthermore, no significant effects were seen on serum potassium and other clinical laboratory tests. There were no adverse events related to the inhalation of Ro 25-1553.

In addition to its bronchodilatory effect, Ro 25-1553 may exert anti-inflammatory effects. Thus, it has been shown that Ro 25-1553, but not original VIP, inhibits allergen induced thromboxane A₂ release in sensitised lungs from guinea pigs perfused in vitro as well as allergen induced eosinophil recruitment in sensitised guinea pig airways in vivo.³⁰ Ro 25-1553 also inhibits endotoxin induced release of the pro-inflammatory cytokines tumour necrosis factor (TNF)- α and interleukin (IL)-12 in human whole blood and in monocytes in vitro, presumably through activation of the VPAC₂ receptor.³¹ It remains to be determined whether the bronchodilatory effect of inhaled Ro 25-1553 is paralleled by the anti-inflammatory effects in the airways of patients with asthma.

In conclusion, inhalation of the VPAC₂ receptor agonist Ro 25-1553 in a single dose causes a rapid bronchodilatory effect in patients with moderate stable asthma without severe cardiovascular side effects. However, because of the limited duration of its bronchodilatory effect, longer acting VPAC₂ receptor agonists should be developed and the bronchodilatory and anti-inflammatory effects evaluated in patients with asthma.

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REFERENCES

- 1 Said SI, Mutt V. Relationship of spasmogenic and smooth muscle relaxant peptides from normal lung to other vasoactive compounds. *Nature* 1977;**265**:84-6.
- 2 Dey RD, Shannon WA, Said SI. Localization of VIP-immunoreactive nerves in airways and pulmonary vessels of dogs, cats, and human subjects. *Cell Tissue Res* 1981;**220**:231-8.
- 3 Laitinen A, Partanen M, Hervonen A, et al. VIP like immunoreactive nerves in human respiratory tract: Light and microscopy study. *Histochemistry* 1985;**82**:313-9.
- 4 Bhogal R, Sheldrick RLG, Coleman RA, et al. The effects of PACAP and VIP on guinea pig tracheal smooth muscle in vitro. *Peptides* 1994;**15**:1237-41.
- 5 Lindén A, Cardell LO, Yoshihara S, et al. PACAP 1-38 as an inhaled bronchodilator in guinea pigs in vivo. *Peptides* 1998;**19**:93-8.
- 6 Conroy DM, St-Pierre S, Sirois P. Relaxant effects of pituitary adenylate cyclase-activating polypeptide (PACAP) on epithelium-intact and -denuded guinea-pig trachea: a comparison with vasoactive intestinal peptide. *Neuropeptides* 1995;**29**:121-7.
- 7 Foda HD, Sharaf HH, Absoud A, et al. Pituitary adenylate-cyclase-activating peptide (PACAP), a VIP-like peptide, has prolonged airway smooth muscle relaxant activity. *Peptides* 1995;**16**:1057-61.
- 8 Kanemura T, Tamaoki J, Chiyotani A, et al. Role of Na⁺-K⁺-ATPase in airway smooth muscle relaxation by vasoactive intestinal peptide and pituitary adenylate cyclase activating peptide. *Res Commun Chem Pathol Pharmacol* 1993;**79**:11-22.
- 9 O'Donnell M, Garipis RJ, Rinaldi N, et al. Ro 25-1553: a novel, long-acting vasoactive intestinal peptide agonist. Part I: In vitro and in vivo bronchodilator studies. *J Pharmacol Exp Ther* 1994;**270**:1282-8.
- 10 Morice A, Sever PS, Unwin RJ. Vasoactive intestinal peptide causes bronchodilation and protects against histamine-induced bronchoconstriction in asthmatic subjects. *Lancet* 1983;**26**:1225-6.
- 11 Palmer J, Cuss F, Warren J, et al. Effect of infused vasoactive intestinal peptide on airway function in normal subjects. *Thorax* 1986;**41**:663-6.
- 12 Bolin DR, Cottrell JM, Mickalewsky J, et al. Degradation of vasoactive intestinal peptide in bronchoalveolar lavage fluid. *Biomed Res* 1992;**13**:25-30.
- 13 Franconi GM, Graf PD, Lazarus SC, et al. Mast cell tryptase and chymase reverse airway smooth muscle relaxation induced by vasoactive intestinal peptide in the ferret. *J Pharmacol Exp Ther* 1989;**248**:947-51.
- 14 Hachisu M, Hiranuma T, Tani S, et al. Enzymatic degradation of helodermin and vasoactive intestinal polypeptide. *J Pharmacobiodyn* 1991;**14**:121-31.
- 15 Lilly CM, Martins MA, Drazen JM. Peptidase modulation of vasoactive intestinal peptide pulmonary relaxation in tracheal superfused guinea pig lungs. *J Clin Invest* 1993;**91**:235-43.
- 16 Lilly CM, Kobzik L, Hall AE, et al. Effects of chronic airway inflammation on the activity and enzymatic inactivation of neuropeptides in guinea pig lungs. *J Clin Invest* 1994;**93**:2667-74.
- 17 Sakai N, Tamaoki J, Kobayashi K, et al. Vasoactive intestinal peptide stimulates ciliary motility in rabbit tracheal epithelium: modulation by neutral endopeptidase. *Reg Peptides* 1991;**34**:33-41.
- 18 Tam EK, Franconi GM, Nadel JA, et al. Protease inhibitors potentiate smooth muscle relaxation induced by vasoactive intestinal peptide in isolated human bronchi. *Am J Respir Cell Mol Biol* 1990;**2**:449-52.
- 19 Tam EK, Caughey GH. Degradation of airway neuropeptides by human lung tryptase. *Am J Respir Cell Mol Biol* 1990;**3**:27-32.
- 20 Bolin DR, Michalewsky J, Wasserman MA, et al. Design and development of a vasoactive intestinal peptide analog as a novel therapeutic for bronchial asthma. *Biopolymers (Peptide Science)* 1995;**37**:57-66.
- 21 Gourlet P, Vertongen P, Vandermeers A, et al. The long-acting vasoactive intestinal polypeptide agonist Ro 25-1553 is highly selective of the VIP₂ receptor subclass. *Peptides* 1997;**18**:403-8.
- 22 Juarranz MG, Rampelbergh JV, Gourlet P, et al. Different vasoactive intestinal polypeptide receptor domains are involved in the selective recognition of two VPAC₂ selective ligands. *Mol Pharmacol* 1999;**56**:1280-7.

- 23 **Källström B-L**, Waldeck B. Bronchodilating properties of the VIP receptor agonist Ro 25 1553 compared to those of formoterol on the guinea-pig isolated trachea. *Eur J Pharmacol* 2001;**430**:335–40.
- 24 **Wong BJ**, Dolovich J, Ramsdale H, *et al*. Formoterol compared with beclomethasone and placebo on allergen-induced asthmatic responses. *Am Rev Respir Dis* 1992;**146**:1156–60.
- 25 **Selroos O**. Oxis® (formoterol) Turbuhaler®: its pharmacological and clinical properties. *Eur Respir Rev* 1998;**8**:216–20.
- 26 **American Thoracic Society (ATS)**. Standards for the diagnosis and care of patients with chronic obstructive pulmonary disease (COPD) and asthma. *Am Rev Respir Dis* 1987;**136**:225–44.
- 27 **Bolin DR**, Cottrell JM, O'Donnell M, *et al*. N-terminal analogs of vasoactive intestinal peptide: identification of a binding pharmacophore. In: Rivier JE, Marshall GR, eds. *Peptides: chemistry and biology*. Proceedings of the Eleventh American Peptide Symposium. Leiden, Netherlands: ESCOM, 1989: 208–10.
- 28 **O'Donnell M**, Garippa RJ, O'Neill N, *et al*. Structure-activity studies of vasoactive intestinal peptide (VIP). *J Biol Chem* 1991;**266**:6389–92.
- 29 **O'Donnell M**, Garippa RJ, Rinaldi N, *et al*. Ro 25-1553: a novel, long-acting vasoactive intestinal peptide agonist. Part II: Effect on in vitro and in vivo models of pulmonary anaphylaxis. *J Pharmacol Exp Ther* 1994;**270**:1289–94.
- 30 **Schmidt DT**, Ruhlman E, Waldeck B, *et al*. The effect of the vasoactive intestinal polypeptide agonist Ro 25 1553 on induced tone in isolated human airways and pulmonary artery. *Naunyn Schmiederbergs Arch Pharmacol* 2001;**364**:314–20.
- 31 **Dewitt D**, Gourlet P, Amraoui Z, *et al*. The vasoactive intestinal peptide analogue Ro 25-1553 inhibits the production of TNF and IL-12 by LPS-activated monocytes. *Immunol Lett* 1998;**60**:57–60.

LUNG ALERT

Identifying adventure travellers at risk of developing exacerbations of asthma

▲ Golan Y, Onn A, Villa Y, *et al*. Asthma in adventure travelers: a prospective study evaluating the occurrence and risk factors for acute exacerbations. *Arch Intern Med* 2002;**162**:2421–6

This Israeli study followed up 180 mild to moderate asthmatics (mean age 23.9 years, 50% female) who were travelling to developing countries for, on average, 4 months. History, examination and spirometric tests were performed before and after exercise.

Asthma attacks during travel were reported by 88 subjects; 32 had their worst ever attack and 11 had life threatening exacerbations. Frequent (≥ 3 /week) use of bronchodilators before travel (RR 3.35, $p < 0.001$), emergency department treatment in the previous year, pre-exercise wheezing, and significant wheezing after exercise were all identified as risk factors for worsening of asthma. Intensive trekking was also a risk factor (RR 2.04, $p = 0.04$). It was noted that most subjects failed to foresee the possibility of worsening of their asthma during travel and there was significant underuse of inhaled corticosteroids in all groups.

Further studies are needed to assess the usefulness of optimising asthma control before travel, avoidance of intensive trekking, and the use of crisis management plans to minimise the frequency of asthma attacks in adventure travellers.

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