Effect of bradykinin on allergen induced increase in exhaled nitric oxide in asthma

F L M Ricciardolo, M C Timmers, J K Sont, G Folkerts, P J Sterk

Thorax 2003;58:840–845

See end of article for authors' affiliations

Correspondence to:
Dr F L M Ricciardolo,
Pulmonary Department, G
Gaslini Institute, Largo
Girolamo Gaslini 5,
16147 Genoa, Italy;
fabioricciardolo@
ospedale-gaslini.ge.it

Revised version received
24 October 2003
Accepted for publication
4 July 2003

Background: Exposure of patients with atopic asthma to allergens produces a long term increase in exhaled nitric oxide (FE_{NO}), probably reflecting inducible NO synthase (NOS) expression. In contrast, bradykinin (BK) rapidly reduces FE_{NO}. It is unknown whether BK suppresses increased FE_{NO} production after allergen exposure in asthma, and whether it modulates FE_{NO} via NOS inhibition.

Methods: Levels of FE_{NO} in response to aerosolised BK were studied before (day 3) and 48 hours after (day 10) randomised diluent (diluent/placebo/BK (Dil/P/BK)), allergen (allergen/placebo/BK (All/P/BK)), and allergen/L-NMMA/BK (All/L/BK) challenges (day 8) in 10 atopic, steroid naïve, mild asthmatic patients with dual responses to inhaled house dust mite extract. To determine whether BK modulates FE_{NO} via NOS inhibition, subjects performed pre- and post-allergen BK challenges after pretreatment with the NOS inhibitor L-NMMA in the All/L/BK period.

Results: Allergen induced a fall in FE_{NO} during the early asthmatic reaction (EAR) expressed as AUC₀₋₁ (ANOVA, $p=0.04$), which was followed by a rise in FE_{NO} during the late asthmatic reaction (LAR) expressed as AUC₁₋₄₈ (ANOVA, $p=0.008$). In the Dil/P/BK period, FE_{NO} levels after BK on pre- and post-diluent days were lower than FE_{NO} levels after placebo (difference 23.5 ppb (95% CI 6.2 to 40.9) and 22.5 ppb (95% CI 7.3 to 37.7), respectively; $p<0.05$). Despite the long lasting increase in FE_{NO} following allergen challenge in the LAR, BK suppressed FE_{NO} levels at 48 hours after allergen challenge in the All/P/BK period, lowering the increased FE_{NO} (difference from placebo 54.3 ppb (95% CI 23.8 to 84.8); $p=0.003$) to the baseline level on the pre-allergen day ($p=0.51$). FE_{NO} levels were lower after L-NMMA than after placebo on pre-allergen (difference 10.85 ppb (95% CI 1.3 to 20.4); $p=0.03$) and post-allergen (difference 36.2 ppb (95% CI 5.5 to 66.9); $p=0.03$) days in the All/L/BK and All/P/BK periods, respectively. L-NMMA did not significantly potentiate the pre- and post-allergen reduction in BK induced FE_{NO}.

Conclusions: Bradykinin suppresses the allergen induced increase in exhaled NO in asthma; this is not potentiated by L-NMMA. Bradykinin and L-NMMA may follow a common pathway in reducing increased NO production before and after experimental allergen exposure. Reinforcement of this endogenous protective mechanism should be considered as a therapeutic target in asthma.

Nitric oxide (NO) is generated from L-arginine by the enzyme NO synthase (NOS), of which constitutive (cNOS) and inducible (iNOS) isoforms have been described functionally.¹ Both are present in the airways.² cNOS (endothelial NOS (eNOS) and neuronal NOS (nNOS)) release small amounts of NO (in the picomolar range) within seconds in response to increases in cytosolic Ca²⁺. Several agonists, including bradykinin, activate cNOS upon receptor stimulation.³ In contrast, the expression of iNOS is induced by proinflammatory cytokines, a process that results in a protracted and Ca²⁺ independent release of high levels of NO (in the nanomolar range)⁴ which is inhibited by corticosteroids.⁵ NO formed by cNOS leads to cGMP dependent airway smooth muscle relaxation, whereas NO released by iNOS is associated with proinflammatory effects.⁷⁻⁸ Both enzyme isoforms are likely to play a role in asthma.⁷⁻⁸ NO is detectable in the exhaled air of healthy volunteers⁹ and is increased in patients with atopic asthma.¹⁰ It has recently been shown that the increased levels of NO in the exhaled air of asthmatic patients are related to airway eosinophilic inflammation¹¹ and to the increased expression of iNOS.⁶ Exhaled NO (FE_{NO}) is inhibited by treatment with inhaled corticosteroids⁵ and NOS inhibitors,¹² suggesting that it is sensitive to steroids and produced by the enzymatic NOS pathway.

In most sensitised asthmatics, allergen exposure induces an early (EAR) and late (LAR) asthmatic response. The EAR,

with early onset (10 minutes after exposure) and short duration (1–2 hours), is followed by the LAR (3–10 hours) which is characterised by bronchoconstriction associated with airway cellular inflammation.¹³ Interestingly, it has been shown that the LAR is associated with increased levels of NO in the exhaled air,¹⁴ suggesting that FE_{NO} may reflect allergic inflammation in asthmatic airways.

Bradykinin is an autacoid involved in the pathophysiology of asthma.¹⁵ Bradykinin induced hyperresponsiveness is increased by NOS inhibitors in asthma,¹⁶ suggesting that bradykinin releases Ca²⁺ dependent cNOS derived NO in the airways.¹⁶ Conversely, inhalation of bradykinin,¹⁷ as well as of prostaglandin E₂ (PGE₂),¹⁸ reduces FE_{NO} in stable asthmatics, which suggests that bradykinin has a role in modulating FE_{NO} in the absence of disease activity. However, it is unknown whether bradykinin can regulate increased FE_{NO} levels during late phase inflammation induced by allergen in asthma and whether it can modulate FE_{NO} via NOS inhibition.

We measured the effect of bradykinin inhalation on NO levels in the exhaled air of asthmatic patients, both at baseline and after experimental allergen exposure in the absence or presence of the NOS inhibitor L-NMMA. This study is part of a larger project for which the study design, methods, and results for other outcome parameters have already been published.¹⁶

Table 1 Characteristics of participants

	Age (years)	Atopic status*	FE _{NO} (ppb)†	FEV ₁ (% predicted) ‡	PC ₂₀ FEV ₁ histamine (mg/ml)
Mean	22.1	4.2	33.5	94.4	1.56
SD	2.6	0.9	23.7	7.2	
Range	19–26	3–6	14–95	81–104	0.29–4.23

SD = standard deviation; FE_{NO} = fractional exhaled concentration of nitric oxide; FEV₁ = forced expiratory volume in 1 s; PC₂₀FEV₁ histamine = provocative concentrations of histamine causing a 20% fall in FEV₁ in the screening period.
*Atopic status as determined by the number of wheal responses to 10 common allergen extracts (Vivodiagnost, ALK, Benelux).
†Values obtained at the first randomised visit.
‡Baseline values as percentage of predicted values in the screening period. Patients have also been described elsewhere.¹⁶

METHODS

Subjects

Ten non-smoking patients (five men) with mild intermittent asthma and house dust mite atopy,¹⁹ as described elsewhere,¹⁶ participated in the study (table 1). All subjects were symptom free at the time of the study and were not on regular medication. They had normal lung function and were hyperresponsive to inhaled histamine.²⁰ Inhaled short acting β_2 agonists were allowed on demand until 12 hours before testing. The subjects had documented EAR and LAR to inhaled house dust mite extract in the screening period.²⁰ The study was approved by the ethics committee of the Leiden University Medical Centre and all participants gave written informed consent.

Study design

The screening period for the selection criteria has been described previously.¹⁶ The study had a randomised, placebo controlled, crossover design and consisted of three periods (diluent/placebo/bradykinin (Dil/P/BK), allergen/placebo/bradykinin (All/P/BK), and allergen/L-NMMA/bradykinin (All/L/BK)) of five study days each, separated by a washout interval of 2–4 weeks. During each study period the participants pretreated with placebo (Dil/P/BK and All/P/BK periods) or L-NMMA (All/L/BK period) underwent a bradykinin challenge before (day 3) and 48 hours after (day 10) the diluent (Dil/P/BK period) or allergen challenge (All/P/BK and All/L/BK periods, day 8; fig 1). On day 9 the participants underwent spirometric tests and FE_{NO} monitoring (fig 1).

On day 1 of each study period a control PC₂₀ histamine measurement was performed.

FE_{NO} was measured (2 minutes after spirometry) at baseline, after placebo or L-NMMA, and at the end of bradykinin (after 5 minutes and every 10 minutes up to 1 hour) and allergen/diluent challenge (every 10 minutes in the first hour, every 30 minutes in the second hour, every 60 minutes up to 10 hours, and at 24 hours), when a fall in forced expiratory volume in 1 second (FEV₁) of $\geq 20\%$ was achieved or after three consecutive diluent inhalations.

Inhalation challenges

Freshly prepared N^G-monomethyl-L-arginine (L-NMMA; Clinalfa AG, Lufelfingen, Switzerland: 10 mg in 3 ml 0.9% saline) or placebo (3 ml 0.9% saline), bradykinin (Clinalfa AG, Lufelfingen, Switzerland: 0.0024–5.0 mg/ml), diluent, and allergen extract of *Dermatophagoides pteronyssinus* (SQ 503; Vivodiagnost, ALK, Benelux) were inhaled as described previously.¹⁶ FEV₁ measurements for bradykinin and allergen or diluent challenges were performed as previously reported.¹⁶

Measurements of exhaled NO

FE_{NO} measurements were performed, according to the present recommendation,²¹ using a Sievers NOA 270B chemiluminescence analyser (Sievers, Boulder, CO, USA) as previously described.²² Subjects performed a slow vital capacity manoeuvre with a constant expiratory flow of 100 ml/s using online visual monitoring. An expiratory

Figure 1 Study design. Dil = diluent; All = allergen; P = placebo; BK = bradykinin; L = N^G-monomethyl-L-arginine.

resistance of 5 cm H₂O was applied to prevent bias of the measurement with nasal NO. Plateau levels of NO were determined and expressed as parts per billion (ppb). Subjects inspired "NO free" air (<1 ppb) during measurements. Three successive recordings were made at 1 minute intervals and the mean was used in the analysis.

Statistical analysis

The difference in FE_{NO} after allergen challenge was expressed in absolute terms as the area under the time-response curve (AUC) from 0 to 48 hours (total AUC_{0-48/48} = mean FE_{NO}), from 0 to 1 hour (AUC₀₋₁) in the EAR, and from 1 to 48 hours (AUC₁₋₄₈) in the LAR. The effect of bradykinin or L-NMMA on FE_{NO} was calculated in terms of the difference between treatments or the difference from placebo with 95% confidence intervals for these differences.

Values in the text and figures are expressed as mean (SE) or 95% confidence intervals (95% CI). In order to measure the effect of allergen and diluent on FE_{NO} we compared AUC₀₋₁, AUC₁₋₄₈, and total AUC_{0-48/48} using one way analysis of variance (ANOVA). Repeated measures ANOVA was applied to test whether there were any differences in baseline FE_{NO} on different days and in FE_{NO} between treatments. Two tailed paired Student's *t* tests were applied to explore the differences. Repeated measures ANOVA of PC₂₀ histamine and FEV₁ values not described in the previous report¹⁶ were performed. In all cases a *p* value of <0.05 was considered significant.

Statistical analyses were performed using Statistica for Windows (StatSoft Inc; Tulsa, OK, USA). A power calculation was based on the standard deviation (SD) of the differences between the bradykinin induced reduction in FE_{NO} during placebo pretreatment in the Dil/P/BK (day 3) and All/P/BK (day 3) periods. Based on this SD (13.6% change in FE_{NO}), the current sample size allows us to detect a difference of 13.6% in FE_{NO} (1 SD) between repeated bradykinin challenges (power 0.8, α 0.05).

RESULTS

Airway responses

PC₂₀ histamine was similar in each study phase (*p* = 0.28). Airway responses to placebo of challenge with L-NMMA (0.9% saline) or L-NMMA (10 mg in 3 ml) plus diluent or allergen challenge have been reported previously.¹⁶ At 24 and 48 hours after allergen challenge FEV₁ was similar to baseline in both the All/P/BK (*p* = 0.57) and All/L/BK periods (*p* = 0.23). Bradykinin challenge before and after diluent or allergen provoked a similar percentage reduction in FEV₁ in the Dil/P/BK period (26 (2)% and 25 (2)%), All/P/BK period (27 (2)% and 28 (2)%), and All/L/BK period (26 (2)% and 28 (2)%), respectively (*p* = 0.44).

Exhaled NO

NO was detectable in the exhaled air of all asthmatic subjects. Baseline FE_{NO} values in the Dil/P/BK period (day 3: 37 (11) ppb; day 8: 38 (12) ppb), All/P/BK period (day 3: 35 (9) ppb; day 8: 33 (8) ppb), and All/L/BK period (day 3: 37 (10) ppb; day 8: 31 (8) ppb) were not different (MANOVA, *p* = 0.48).

There was no change in FE_{NO} up to 48 hours after diluent challenge (day 8), whereas allergen challenge on day 8 reduced FE_{NO} in the first hour after the end of the challenge in the All/P/BK and All/L/BK periods (fig 2), as shown by the significant differences between AUC₀₋₁ for FE_{NO} in the All/P/BK period (19.9 ppb/h (95% CI 9.2 to 30.7)), the All/L/BK period (17.9 ppb/h (95% CI 8.5 to 27.4)), and the Dil/P/BK period (38.1 ppb/h (95% CI 9.6 to 66.6); *p* = 0.04). FE_{NO} was increased in the LAR after allergen challenge in the All/P/BK and All/L/BK periods (fig 2). AUC₁₋₄₈ for FE_{NO} on day 8 was

Figure 2 Time course of the change in exhaled nitric oxide levels after diluent (circles) and allergen (allergen/placebo/bradykinin: squares; allergen/L-NMMA/bradykinin: rhombus) challenges in 10 atopic patients with mild asthma. Mean (SE) values are shown.

significantly different in the All/P/BK (3282 ppb/h (95% CI 1693 to 4870)), All/L/BK (2579 ppb/h (95% CI 1467 to 3690)), and Dil/P/BK periods (1781 ppb/h (95% CI 471 to 3091); *p* = 0.008). The differences in total AUC_{0-48/48} for FE_{NO}, interpreted as the mean FE_{NO}, on day 8 in the All/P/BK (68.8 ppb (95% CI 35 to 102)), All/L/BK (54.1 ppb (95% CI 31 to 77)), and Dil/P/BK periods (37.8 ppb (95% CI 10 to 65)) were significant (*p* = 0.009). The differences in all the AUCs for FE_{NO} between diluent and the two allergen challenges are shown in table 2.

In the Dil/P/BK period FE_{NO} values were significantly lower after bradykinin than after placebo (fig 3) on day 3 (difference 23.5 ppb (95% CI 6.2 to 40.9); *p* = 0.01) and day 10 (difference 22.5 ppb (95% CI 7.3 to 37.7); *p* = 0.008). Both FE_{NO} values after bradykinin in the Dil/P/BK period were not significantly different from the corresponding FE_{NO} value on the pre-allergen day in the All/P/BK period (*p* = 0.15). Forty eight hours after allergen challenge in the All/P/BK period bradykinin completely suppressed the increased FE_{NO} levels (fig 3), as shown by the reduction in the allergen induced increase in FE_{NO} (difference from placebo: 54.3 ppb (95% CI 23.8 to 84.8); *p* = 0.003) to the baseline level of the pre-allergen day (*p* = 0.51). The difference between the reductions in FE_{NO} induced by bradykinin from the respective placebo 48 hours after allergen in the All/P/BK period and after diluent in the Dil/P/BK period was statistically significant (31.8 ppb (95% CI 7.1 to 56.5); *p* = 0.017).

In the Dil/P/BK and All/P/BK periods, neither placebo nor L-NMMA (0.9% saline) changed baseline FE_{NO} on any study days (fig 3). In the All/L/BK period the differences between the level of FE_{NO} after L-NMMA and placebo before and after allergen were significant (All/L/BK: 10.85 ppb (95% CI 1.3 to 20.4), *p* = 0.029; All/P/BK: 36.2 ppb (95% CI 5.5 to 66.9), *p* = 0.026).

After pretreatment with L-NMMA, bradykinin significantly reduced FE_{NO} on pre-allergen (difference from L-NMMA: 14.1 ppb (95% CI 6.8 to 21.4); *p* = 0.002) and post-allergen (difference from L-NMMA: 29.7 ppb (95% CI 16 to 43.5); *p* = 0.0008) days (fig 3). The differences between the level of FE_{NO} after placebo plus bradykinin in the All/P/BK period and after L-NMMA plus bradykinin in the All/L/BK period on pre- and post-allergen days (1.96 ppb (95% CI -2.1 to 6), *p* = 0.3; and 9.4 ppb (95% CI -0.5 to 19.5), *p* = 0.06, respectively) were not statistically significant. The difference between the baseline values of FE_{NO} (48 hours after allergen) on the post-allergen day (day 10) in the All/P/BK and All/L/BK periods was 15.8 ppb (95% CI -5 to 36.4); *p* = 0.11),

Table 2 Mean (95% CI) differences in AUC₀₋₁, AUC₁₋₄₈, and total AUC/48 for FE_{NO} between diluent (Dil/P/BK) and allergen (All/P/BK and All/L/BK) challenges

	Difference in AUC ₀₋₁ (ppb/h)	Difference in AUC ₁₋₄₈ (ppb/h)	Difference in total AUC/48 (ppb)
Diluent (Dil/P/BK period) v allergen (All/P/BK period)	18.1 (-1.3 to 37.5); p=0.06	-1500 (-2469 to -532.7); p=0.006	-31 (-51 to -11); p=0.007
Diluent (Dil/P/BK period) v allergen (All/L/BK period)	20.2 (-4.8 to 45); p=0.10	-798 (-1908 to 312); p=0.13	-16.2 (-40 to 7); p=0.15
Allergen (All/P/BK period) v allergen (All/L/BK period)	2.05 (-5 to 9.1); p=0.53	702 (-55 to 1460); p=0.07	14.7 (-1 to 30); p=0.07

which explains the slight difference between FE_{NO} after placebo plus bradykinin in the All/P/BK period and after L-NMMA plus bradykinin in the All/L/BK period on the post-allergen day.

On all bradykinin treatment days the recovery time (up to 60 minutes) was characterised by a complete spontaneous recovery in FEV₁ values to baseline levels, whereas FE_{NO} levels remained significantly lower than baseline ($p < 0.05$, fig 4).

DISCUSSION

This study shows, for the first time, that the allergen induced increase in FE_{NO} in atopic asthma is suppressed by bradykinin, and that the NOS inhibitor L-NMMA significantly reduces pre- and post-allergen FE_{NO} levels and does not potentiate the bradykinin induced reduction in FE_{NO}. Taken together, our results indicate that bradykinin, via NOS inhibition, modulates the prolonged increase in FE_{NO} induced by allergen exposure. Our findings suggest that bradykinin inhibits excessive NO production resulting from an overexpression of the NOS pathway in asthmatic airways during an exacerbation.

The bradykinin induced fall in FE_{NO} levels in stable asthmatics has recently been reported by Kharitonov *et al.*¹⁷. We have shown that bradykinin suppresses increased levels of FE_{NO} 48 hours after allergen challenge, which suggests that it has a role in modulating exacerbations of allergic inflammation. Furthermore, the finding that FE_{NO} levels were still significantly reduced 60 minutes after the end of the pre-allergen and post-allergen bradykinin challenge, when FEV₁ had completely recovered, suggests that the rapid suppression of FE_{NO} by bradykinin has a relatively long duration and is not dependent on airway calibre. One possible explanation for the action of bradykinin is vasodilation¹⁵ with potentially enhanced NO trapping by haemoglobin²³ and/or altered diffusion of NO to the airway lumen due to mucosal swelling and gland secretion¹⁵ with subsequent airway obstruction; however, the short term effect of vasodilation and the reduction in FE_{NO} even after recovery of the obstruction limit the validity of this hypothesis.

In this study we also examined whether bradykinin negatively modulates FE_{NO} through inhibition of NOS. L-NMMA has previously been found to reduce FE_{NO} levels significantly in stable asthma,¹² so we studied the effect of L-NMMA on the bradykinin induced reduction in FE_{NO}. The lack of significant potentiation by L-NMMA on the bradykinin induced reduction in FE_{NO} suggests that bradykinin and L-NMMA share a common pathway in inhibiting FE_{NO} production in the airways of patients with asthma. It has recently been found that prostaglandins also reduce FE_{NO} in asthmatic subjects.¹⁸ Bradykinin stimulates PGE₂ release from airway epithelial cells, either by the constitutive isoform of cyclo-oxygenase enzyme (COX-1) or by the inducible isoform (COX-2),²⁴ and PGE₂ negatively modulates the induction of iNOS expression at the transcriptional level.²⁵

It is therefore possible that the reduction in FE_{NO} induced by bradykinin is a result of the inhibition of iNOS expression through the release of PGE₂. However, the rapid reduction in FE_{NO} observed after bradykinin inhalation does not favour such a delayed mechanism, but suggests a possible direct or indirect influence of bradykinin on NOS at the post-transcriptional level. It has previously been shown that high levels of NO can downregulate NOS pathways by negative feedback,²⁶ and also that augmented production of FE_{NO} in the lower respiratory tract of eNOS deficient mice is associated with pulmonary iNOS overexpression, indicating a counter-regulation between the two NOS isoforms.²⁷ Furthermore, a recent study in human mesangial cells showed a negative post-transcriptional regulation of iNOS by the NO/cGMP pathway.²⁸ On the basis of the present results we believe that bradykinin is not only an activator of the Ca²⁺ dependent cNOS/cGMP pathway which modulates airway hyperresponsiveness,²⁹ but also may downregulate excessive NO release, probably derived by iNOS, through negative feedback in allergic asthma.

In line with this hypothesis, we have previously shown that iNOS (but not eNOS or nNOS) immunostaining is higher after allergen exposure in the epithelium of bronchial biopsy sections of subjects with asthma,¹⁶ which suggests two major findings on the basis of the present data: (1) iNOS may be responsible for the increase in FE_{NO} after allergen; (2) bradykinin should affect iNOS activity by reducing FE_{NO} in asthmatic patients and, in particular, by suppressing the increase in FE_{NO} after exposure to allergen.

In the present study, allergen inhalation led to two opposing changes in FE_{NO} levels in atopic mild asthmatics. Exhaled NO tended to decrease during the EAR and exhibited a prolonged increase during and after the LAR. These findings are consistent with previous data showing the effect of allergen on FE_{NO} levels in atopic asthmatic patients with LAR.¹⁴ In the early phase we found a reduction in FE_{NO} levels compared with baseline, which confirms previous observations.³⁰ In previous studies acute airways obstruction has been found to have a partial effect on FE_{NO} levels.³¹⁻³² It is therefore possible that bronchoconstriction could, at least partially, lead to a fall in FE_{NO} levels during the early phase of the bronchial allergic reaction, possibly as a result of limiting NO diffusion or increased airflow velocity in constricted airways. We also postulate an indirect effect of allergen on FE_{NO} levels during the early phase reaction via the release of proinflammatory mediators such as kinins or prostaglandins.¹⁷⁻¹⁸

Exhaled NO appeared to increase 9–48 hours after allergen inhalation, confirming a previous report.¹⁴ In the latter study the authors observed a decline in FE_{NO} by 27 hours. The longer duration of increased FE_{NO} in our study could be a reflection of a possibly higher level of allergic inflammation induced by allergen exposure. Several lines of evidence suggest that FE_{NO} reflects iNOS expression in asthma,⁵⁻⁶ which recognises that FE_{NO} is a non-invasive marker of

Figure 3 Effect of bradykinin challenge on exhaled nitric oxide in (A) the diluent/placebo/bradykinin period, (B) the allergen/placebo/bradykinin period, and (C) the allergen/L-NMMA/bradykinin period. Exhaled nitric oxide levels were measured before pre-treatment (open columns), after pretreatment (with placebo or L-NMMA: shaded columns), and 5 minutes after bradykinin challenge (filled columns) on days 3 and 10 of each study period. On day 8 diluent (diluent/placebo/bradykinin) and allergen (allergen/placebo/bradykinin and allergen/L-NMMA/bradykinin) challenges were performed. Mean (SE) values are shown. * $p < 0.05$ v respective placebo or L-NMMA.

airway inflammation in asthma.³³ The delayed and long term increase in FE_{NO} during the LAR could be explained by the action of proinflammatory cytokines released from inflammatory cells recruited to the airways after allergen exposure, and of reactive oxygen species with subsequent upregulation of iNOS expression.^{34–35} Furthermore, an additional explanation could be derived from the recent evidence that airway acidification converts nitrite (NO_2^-) to NO gas in quantities consistent with those observed in expired air during an asthma exacerbation,³⁶ or after the LAR to allergen.

Figure 4 Time course of the change in exhaled nitric oxide (rhombus) and forced expiratory volume in one second (FEV_1 ; squares) during the recovery period after diluent in (A) the diluent/placebo/bradykinin period and after allergen in (B) the allergen/placebo/bradykinin period and (C) the allergen/L-NMMA/bradykinin period on the day of bradykinin challenge (day 10) in 10 atopic patients with mild asthma. Mean (SE) values are shown.

To avoid differences in FE_{NO} as a result of smoking or medication use, we selected a homogeneous group of non-smoking, atopic, mild intermittent asthmatic subjects who were not using any steroid medication before or during the experiments. It has recently been shown that repeated forced vital capacity (FVC) manoeuvres significantly reduce FE_{NO} levels in healthy and asthmatic subjects by 13% and 10%, respectively.³⁷ Even though our FE_{NO} measurements were performed 2 minutes after an FVC manoeuvre, we do not consider that it had any effect because no effect on the FE_{NO} measurement was seen following diluent.

NO released by iNOS is likely to exert proinflammatory activities such as induction of eosinophil chemotaxis and activation of the Th2 driven immune response,^{8–11} as observed during the late phase response to inhaled allergen. The bradykinin induced suppression of raised FE_{NO} levels during the late phase suggests that bradykinin can modulate allergic inflammation exacerbations in the airways by the inhibition of proinflammatory NOS. On the basis of our study, a clinically relevant scenario could be outlined in patients with asthma where an acute inflammatory mediator (bradykinin) could have a modulatory role in limiting the effect of a detrimental long lasting inflammatory pathway (iNOS). The

reinforcement of such endogenous protective mechanisms should be considered as a potential therapeutic target in asthma.

ACKNOWLEDGEMENTS

This study was supported by a Research Fellowship of the European Respiratory Society (ERS).

Authors' affiliations

F L M Ricciardolo, M C Timmers, P J Sterk, Department of Pulmonology, Leiden University Medical Center, The Netherlands

F L M Ricciardolo, Pulmonary Department, G Gaslini Institute, Genoa, Italy

J K Sont, Department of Medical Decision Making, Leiden University Medical Center, The Netherlands

G Folkerts, Department of Pharmacology and Pathophysiology, Faculty of Pharmacy, Utrecht University, The Netherlands

REFERENCES

- Moncada S, Palmer RMJ, Higgs EA. Nitric oxide: physiology, pathophysiology, and pharmacology. *Pharmacol Rev* 1996;**43**:109–42.
- Gaston B, Lobzik L, Stamler JS. Distribution of nitric oxide synthase in the lung. In: Zapol WM, Bloch KD, eds. *Nitric oxide and the lung*. New York: Marcel Dekker, 1997:75–86.
- Nathan C. Nitric oxide as a secretory product of mammalian cells. *FASEB J* 1992;**6**:3051–64.
- Morris SM, Billiar TR. New insights into the regulation of inducible nitric oxide synthesis. *Am J Physiol* 1994;**266**:E829–39.
- Kharitonov SA, Yates DH, Barnes PJ. Inhaled glucocorticoids decrease nitric oxide in exhaled air of asthmatic patients. *Am J Respir Crit Care Med* 1996;**153**:454–7.
- Saleh D, Ernst P, Lim S, et al. Increased formation of the potent oxidant peroxynitrite in the airways of asthmatic patients is associated with induction of nitric oxide synthase: effect of inhaled glucocorticoid. *FASEB J* 1998;**12**:929–37.
- Nijkamp FP, Folkerts G. Nitric oxide and bronchial reactivity. *Clin Exp Allergy* 1994;**24**:905–14.
- Barnes PJ, Liew FY. Nitric oxide and asthmatic inflammation. *Immunol Today* 1995;**16**:128–30.
- Gustafsson LE, Leone AM, Persson MG, et al. Endogenous nitric oxide is present in exhaled air of rabbits, guinea pigs and humans. *Biochem Biophys Res Commun* 1991;**181**:852–7.
- Gratziau Ch, Lignos M, Dassiou M, et al. Influence of atopy on exhaled nitric oxide in patients with stable asthma and rhinitis. *Eur Respir J* 1999;**14**:897–901.
- Jatakanon A, Lim S, Kharitonov SA, et al. Correlation between exhaled nitric oxide, sputum eosinophils, and methacholine responsiveness in patients with mild asthma. *Thorax* 1998;**53**:91–5.
- Yates DH, Kharitonov SA, Robbins RA, et al. Effect of a nitric oxide synthase inhibitor and a glucocorticosteroid on exhaled nitric oxide. *Am J Respir Crit Care Med* 1995;**152**:892–6.
- Weersink EJM, Postma DS, Aalbers R, et al. Early and late asthmatic reaction after allergen challenge. *Respir Med* 1994;**88**:103–14.
- Kharitonov SA, O'Connor BJ, Evans DJ, et al. Allergen-induced late asthmatic reactions are associated with elevation of exhaled nitric oxide. *Am J Respir Crit Care Med* 1995;**151**:1894–9.
- Barnes PJ. Bradykinin. In: Barnes PJ, Grunstein MM, Leff AR, Woolcock AJ, eds. *Asthma*. New York: Lippincott-Raven, 1997:577–84.
- Ricciardolo FLM, Timmers MC, Geppetti P, et al. Allergen-induced impairment of bronchoprotective nitric oxide synthesis in asthma. *J Allergy Clin Immunol* 2001;**108**:198–204.
- Kharitonov SA, Sapienza MM, Chung KF, et al. Prostaglandins mediate bradykinin-induced reduction of exhaled nitric oxide in asthma. *Eur Respir J* 1999;**14**:1023–7.
- Kharitonov SA, Sapienza MA, Barnes PJ, et al. Prostaglandins E₂ and F_{2α} reduce exhaled nitric oxide in normal and asthmatic subjects irrespective of airway caliber changes. *Am J Respir Crit Care Med* 1998;**158**:1374–8.
- National Heart, Lung and Blood Institute. Expert panel report 2. Guidelines for the diagnosis and management of asthma, NIH Publication No 97-4051. Bethesda, MD: National Institutes of Health, 1997.
- Sterk PJ, Fabbri LM, Quanjer PH, et al. Airway responsiveness: standardized challenge testing with pharmacological, physical and sensitizing stimuli in adults. *Eur Respir J* 1993;**6**(Suppl 16):53–83.
- American Thoracic Society. Recommendations for standardized procedures for the online and offline measurement of exhaled lower respiratory nitric oxide and nasal nitric oxide in adults and children. *Am J Respir Crit Care Med* 1999;**160**:2104–17.
- De Gouw HWFM, Marshall-Partridge SJ, van der Veen H, et al. Role of nitric oxide in the airway response to exercise in healthy and asthmatic subjects. *J Appl Physiol* 2001;**90**:586–92.
- Gow AJ, Stamler JS. Reactions between nitric oxide and haemoglobin under physiological conditions. *Nature* 1998;**391**:169–73.
- Saunders MA, Belvisi MG, Cirino G, et al. Mechanisms of prostaglandin E₂ release by intact cells expressing cyclooxygenase-2: evidence for a "two-component" model. *J Pharmacol Exp Ther* 1999;**288**:1101–6.
- Tetsuka T, Daphna-Iken D, Srivastava SK, et al. Cross-talk between cyclooxygenase and nitric oxide pathways: prostaglandin E₂ negatively modulates induction of nitric oxide synthase by interleukin 1. *Proc Natl Acad Sci USA* 1994;**91**:12168–72.
- Grisavage JM, Hobbs AJ, Ignarro LJ. Negative modulation of nitric oxide synthase by nitric oxide and nitroso compounds. *Adv Pharmacol* 1995;**34**:215–34.
- Cook SP, Vollenweider P, Menard B, et al. Increased eNO and pulmonary iNOS expression in eNOS null mice. *Eur Respir J* 2003;**21**:770–3.
- Perez-Sala D, Cernuda-Morallon E, Diaz-Cazorla M, et al. Posttranscriptional regulation of human iNOS by the NO/cGMP pathway. *Am J Physiol* 2001;**280**:F466–73.
- Figini M, Ricciardolo FLM, Javdan P, et al. Evidence that epithelium-derived relaxing factor released by bradykinin in the guinea pig trachea is nitric oxide. *Am J Respir Crit Care Med* 1996;**153**:918–23.
- Dweik RA, Comhair SAA, Gaston B, et al. NO chemical events in the human airway during the immediate and late antigen-induced asthmatic response. *Proc Natl Acad Sci USA* 2001;**98**:2622–7.
- De Gouw HWFM, Hendriks J, Wolman AM, et al. Exhaled nitric oxide (NO) is reduced shortly after bronchoconstriction to direct and indirect stimuli in asthma. *Am J Respir Crit Care Med* 1998;**158**:315–9.
- Ho LP, Wood FT, Robso A, et al. The current single exhalation method of measuring exhaled nitric oxide is affected by airway calibre. *Eur Respir J* 2000;**15**:1009–13.
- Gustafsson LE. Exhaled nitric oxide as a marker in asthma. *Eur Respir J* 1998;**11**(Suppl 26):49S–52S.
- Hamid Q, Springall DR, Riveros-Moreno V, et al. Induction of nitric oxide synthase in asthma. *Lancet* 1993;**342**:1510–3.
- Adcock IM, Brown CR, Kwon OJ, et al. Oxidative stress induces NF-κB DNA binding and inducible NOS mRNA in human epithelial cells. *Biochem Biophys Res Commun* 1994;**199**:1518–24.
- Hunt JF, Fang K, Malik R, et al. Endogenous airway acidification. Implications for asthma pathophysiology. *Am J Respir Crit Care Med* 2000;**161**:694–9.
- Silkoff PE, Wakita S, Chatkin J, et al. Exhaled nitric oxide after β₂-agonist inhalation and spirometry in asthma. *Am J Respir Crit Care Med* 1999;**159**:940–4.