Histamine dose-response curves in asthma: reproducibility and sensitivity of different indices to assess response

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ABSTRACT In 18 clinically stable asthmatic patients histamine inhalation challenges were performed with a Wright's nebuliser and tidal volume breathing for two minutes on two to four occasions for each subject at a maximum interval of two weeks. The response was measured in terms of specific lung conductance (sGL) by the subtraction technique, maximum partial and maximum complete expiratory flow at 40% and 50% of vital capacity respectively ($\dot{V}_{\max_{40\%}}$ and $\dot{V}_{\max_{50\%}}$), and FEV_1 from the maximum flow-volume curve. Dose-response curves were analysed for (1) provocative concentration (PC) of histamine causing a 20% fall in FEV_1 and a 40% change in the other measurements; (2) threshold concentration (TC)—the concentration at which changes in the measurement exceed 2 SD from control values; (3) reactivity (R)—the slope of the dose-response curve beyond TC. We found that PC_{20,FEV_1} was the most reproducible index, the 95% confidence intervals based on a single determination being ± 1.6 single two-fold concentration difference. PC_{20,FEV_1} was more reproducible than PC values for other measurements and more reproducible than any of the TC values. The 95% confidence intervals based on a single determination of R varied from $\pm 52\%$ to $\pm 74\%$ change/log histamine concentration. Both sGL and $\dot{V}_{\max_{40\%}}$ detected the bronchoconstrictor response assessed by PC and TC at a significantly lower histamine concentration than FEV_1 ($p < 0.01$ and $p < 0.05$ respectively). PC and TC results showed a significant correlation, but neither were correlated with R.

After Dautrebande and Philippot¹ introduced non-allergic bronchial inhalation challenges in 1941, Tiffeneau,² Curry,³ and De Vries⁴ used the tests for clinical purposes. Attempts to standardise the procedure have been made,^{5,6} but so far only a few studies of reproducibility have been published.⁷⁻⁹ In these studies the forced expiratory volume in one second (FEV_1) only was used to monitor response, and the dose-response curve was assessed by only one index, either the threshold concentration (TC) or the provocative concentration (PC) of the stimulus causing a predetermined fall in FEV_1 . We decided to study histamine dose-response curves in clinically stable asthmatic subjects to assess the reproducibility and sensitivity of different measurements of bronchoconstriction, using different indices from the dose-response curves.

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Accepted 16 February 1983

Methods

PATIENTS

We studied 18 patients (five male, thirteen female) aged 19-55 years (table 1). All satisfied the criteria for asthma of the American Thoracic Society¹⁰ and were clinically stable at the time of study, with no nocturnal awakening due to asthma and minimum symptoms by day. None of the subjects reported any respiratory infection in the six weeks preceding the test. Patients with immediate positive skin reactions to animal dander and pollen had had no exposure to these allergens in the month preceding the study. At the time of study all subjects were taking bronchodilator medication on a regular basis or less frequently, and 12 were taking inhaled beclomethasone regularly. Bronchodilators were withheld before the inhalation tests for the time interval suggested by the special committee of the American Academy of Allergy.⁵ Written consent was obtained from each subject and the project was accepted by the local ethical committee.

Table 1 Baseline anthropometric, functional, and clinical data

No	Sex	Age (y)	Height (cm)	sGL ($s^{-1} kPa^{-1}$)	FEV ₁		$\dot{V}_{max_{50c}}$		Medication
					Obs (l)	%p	Obs ($l s^{-1}$)	%p	
1	M	48	180	0.80	3.2	89	5.7	105	B; BDT reg
2	F	42	158	2.09	3.5	140	3.7	80	B; BDT reg
3	M	25	158	2.50	3.8	104	4.6	74	B; BDT reg
4	F	32	165	1.08	2.4	86	5.1	104	BDT reg
5	F	33	158	2.37	2.7	104	3.8	79	BDT reg
6	F	38	165	1.27	3.0	115	4.3	91	B; BDT reg
7	F	23	155	0.79	2.4	89	4.0	80	BDT prn
8	M	28	180	0.50	3.4	81	4.7	77	B; BDT reg
9	F	19	168	2.55	3.2	93	4.0	74	BDT prn
10	F	39	161	1.06	2.6	104	3.0	66	B; BDT reg
11	F	55	157	1.55	2.2	116	4.0	94	B; BDT reg
12	M	21	170	1.02	2.5	62	2.0	33	B; BDT reg
13	F	26	168	1.65	3.0	101	2.4	45	B; BDT reg
14	F	38	170	1.05	2.9	107	3.4	71	BDT prn
15	F	49	160	1.01	2.2	99	3.1	70	B; BDT reg
16	F	52	155	1.16	1.6	82	1.9	43	B; BDT reg
17	M	39	180	0.44	2.3	60	1.6	27	B; BDT reg
18	F	48	161	1.31	2.3	106	2.4	53	B; BDT reg
Mean	—	36.4	164.9	1.34	2.7	96.6	3.5	70.3	
SD	—	11.2	8.4	0.65	0.6	19.1	1.2	22.5	

sGL—specific lung conductance; $\dot{V}_{max_{50c}}$ —maximum complete expiratory flow rate at 50% vital capacity; %p—% predicted values for FEV₁¹⁶ and $\dot{V}_{max_{50c}}$ ¹⁷; BDT—bronchodilator (β_2 -adrenergic agents or xanthine preparations or both); B—beclomethasone inhaler; reg—regularly (two or four times daily); prn—only if needed.

MEASUREMENTS

All measurements were made in a flow-displacement body plethysmograph as described by Leith and Mead.¹¹ After pressure compensation of the integrated flow signal, the volume signal was flat ($\pm 5\%$) to 12 cycles s^{-1} . Lung resistance (RL) was calculated by the subtraction technique described by Mead and Whittenberger¹² and lung volume by using Boyle's law. Flow at the mouth was measured with a No 4 Fleisch pneumotachograph connected to a Validyne pressure transducer. The flow signal was electrically integrated to derive volume, from which FEV₁ was calculated. Pleural pressure was estimated from an oesophageal balloon¹³ 10 cm long connected to a Validyne differential pressure transducer via a 100-cm polyethylene catheter. The other side of the transducer was connected to an oral pressure tap to give transpulmonary pressure. RL was measured as the ratio of change in transpulmonary pressure to change in mouth flow during tidal breathing.

Partial flow-volume curves were initiated from residual volume. Partial and complete maximum expiratory flow-volume curves were drawn on a Hewlett-Packard X-Y recorder. Maximum flow from the partial and complete flow-volume curves was measured at 40% ($\dot{V}_{max_{40p}}$) and 50% ($\dot{V}_{max_{50c}}$) of the forced vital capacity (FVC) respectively. When FVC changed by more than 5% a correction was made for total lung capacity as proposed by Habib *et al.*¹⁴ When changes in TLC were 10% or

less maximum expiratory flow-volume curves were matched at TLC. A volume equivalent to 60% of control FVC was subtracted from TLC and flow rates at this volume were obtained from the partial flow-volume curve. A similar procedure was used to determine flows at 50% of FVC from the complete flow-volume curve. When TLC changes were more than 10% the results were discarded.

Reference values were taken from Goldman and Becklake¹⁵ and from Chermiack¹⁶ for FEV₁ and $\dot{V}_{max_{50c}}$ respectively.

STUDY DESIGN

Subjects were studied on two (10 subjects), three (one subject), or four (seven subjects) occasions, at the same time of day on separate days, with a maximum interval between studies of two weeks. Baseline measurements of RL and thoracic gas volume (TGV) were carried out to derive specific lung conductance (sGL), and partial and complete maximum expiratory flow-volume curves were then produced. Subjects then inhaled phosphate buffer saline from a Wright's nebuliser (output 0.15 ml/min) through a face mask for two minutes,⁶⁻⁸ breathing tidally at a rate of 14 breaths a minute in time with a metronome. RL and TGV were measured one to two minutes after the end of nebulisation. The subject then performed two maximum partial expiratory manoeuvres, initiated from the end-inspiratory position. These were followed immediately by two complete forced expiratory manoeuvres

initiated from TLC. All measurements were completed within three minutes of the end of nebulisation. This interval is within the period of the maximum bronchoconstrictor effect of histamine, which has been estimated to last for a mean of 16.8 minutes (range 4–37 min).¹⁷ Two inhalations of diluent were followed by increasing concentrations of histamine, from 0.03 to a maximum dose of 32 mg/ml. The histamine was given at five-minute intervals to prevent any cumulative effect of histamine.⁷ The test was stopped when the patient showed a 30–50% fall in FEV₁.

ANALYSIS OF DOSE-RESPONSE CURVES

Dose-response curves were drawn on a semi-log scale, the abscissa representing the concentration and the ordinate the percentage change in each measurement. The following indices were obtained from each dose-response curve: (1) TC—the concentration of histamine which produced a change in excess of 2 SD from mean post-diluent values (four post-diluent values for FEV₁ and maximum flows and 10 post-diluent values for sGL); for the 10 subjects who made only two visits there were insufficient measurements to calculate TC for sGL; (2) PC—the concentration of histamine causing a predetermined fall in each measurement, 20% for FEV₁ (PC₂₀) and 40% for the other measurements (PC₄₀). These percentages were well outside the range of measurements seen after inhalation of diluent (table 2); (3) reactivity (R)—the slope of the

Table 2 Reproducibility of post-diluent assessments

	FEV ₁	sGL	$\dot{V}_{max_{40p}}$	$\dot{V}_{max_{50c}}$
Coefficient of variation (%):				
Mean	2.6	10.0	8.2	5.5
SD	1.2	3.1	3.8	3.2
Range	0–5.3	2.8–16.2	2.9–15.8	0–15.3

Individual coefficients of variation obtained at each visit from four post-diluent measurements of FEV₁, $\dot{V}_{max_{40p}}$ and $\dot{V}_{max_{50c}}$, and 10 measurements of sGL (51 assessments in 18 subjects for each measurement except sGL, for which there were 31 measurements in eight subjects. Abbreviations as in table 1.

dose-response curve beyond TC, including at least three points for each curve. For the 10 subjects for whom TC sGL could not be obtained R was calculated from points in excess of a 30% fall in sGL. This is close to the upper limit of values for sGL in other subjects after inhalation of diluent (that is, twice the coefficient of variation) (table 3). The slope of the dose-response curve was calculated by the method of least squares, in keeping with previous studies of FEV₁,^{18,19} specific airway conductance,^{20,21} and maximum expiratory flows.¹⁴ Curves were retained for analysis only if $p < 0.05$. Slope a from the formula $y = ax + b$ was used to characterise reactivity.

STATISTICAL MEASUREMENTS

A one-way analysis of variance was used to compare within-subject and between-subject variance for log TC, log PC, and R, calculating intraclass correlation

Table 3 Reproducibility of dose-response curves: results from one-way analysis of variance

Functional index	F	IC correlation*	95% CI overall mean*	95% CI single determination*
Provocative concentration (PC)				
PC ₂₀ -FEV ₁	23.5†	0.88	±0.72	±1.59
PC ₄₀ -sGL	13.4†	0.81	±1.00	±2.14
$\dot{V}_{max_{40p}}$	12.3†	0.80	±0.81	±1.83
$\dot{V}_{max_{50c}}$	15.5†	0.84	±0.76	±1.71
Threshold concentration (TC)				
TC				
FEV ₁	11.6†	0.79	±0.85	±1.70
sGL	22.1†	0.85	±2.10	±2.24
$\dot{V}_{max_{40p}}$	4.6†	0.57	±1.65	±1.92
$\dot{V}_{max_{50c}}$	6.7†	0.67	±1.45	±1.74
Reactivity (R)				
R				
FEV ₁	11.4†	0.79	±12	±52
sGL	9.2†	0.76	±16	±60
$\dot{V}_{max_{40p}}$	2.7‡	0.38	±14	±74
$\dot{V}_{max_{50c}}$	5.4†	0.62	±15	±71

*Intraclass (IC) correlation—between-subject variance/total variance; 95% CI overall mean—95% confidence interval for the overall mean over subjects; 95% CI single determination—95% confidence interval for subjects based on a single determination. The 95% CI overall mean and 95% CI single determination are expressed as single two-fold concentration difference for PC and TC and as slopes (% change/log histamine concentration) for R. Other abbreviations as in table 1.

† $p < 0.001$; ‡ $p < 0.01$.

Table 4 Comparison between the use of provocative concentration (PC), threshold concentration (TC), and reactivity (R): correlation coefficients (r) for paired indices (21–51 assessments for each comparison)

PC ₂₀ -FEV ₁	*	0.82†	0.73†	0.81†										
PC ₄₀														
sGL	—	*	0.84†	0.91†										
Vmax _{40p}	—	—	*	0.95†										
Vmax _{50c}	—	—	—											
TC														
FEV ₁	0.87†	—	—	—	*	0.79†	0.75†	0.73†						
sGL	—	0.91†	—	—	—	*	0.53†	0.79†						
Vmax _{40p}	—	—	0.80†	—	—	—	*	0.61†						
Vmax _{50c}	—	—	—	0.63†	—	—	—	*						
R														
FEV ₁	0.16	—	—	—	0.22	—	—	—	*	0.30§	0.34§	0.30§		
sGL	—	0.21	—	—	—	0.23	—	—	—	*	0.90	0.74†		
Vmax _{40p}	—	—	0.03	—	—	—	0.19	—	—	—	*	0.46†		
Vmax _{50c}	—	—	—	0.08	—	—	—	0.24	—	—	—	*		
PC ₂₀ -FEV ₁														
PC ₄₀														
Vmax _{40p}														
Vmax _{50c}														
TC														
FEV ₁														
sGL														
Vmax _{40p}														
Vmax _{50c}														
R														
FEV ₁														
sGL														
Vmax _{40p}														
Vmax _{50c}														

*Results of one-way analysis of variance given in table 3.

†p < 0.001; §p < 0.01; §p < 0.05; for other values p > 0.05.

as between-subject variance/total variance. The 95% confidence interval for the overall mean over subjects (that is, the 95% confidence interval of the mean for all subjects) and the range for subjects based on a single determination (that is, the 95% confidence interval for which a single value sampled at random would belong to the population) were calculated.⁸ Log TC and log PC results for each measurement were compared by analysis of variance.

Linear regression analysis was used to compare log TC, log PC, and R at each visit.

Results

BASELINE FUNCTIONAL RESULTS

On the initial assessment 14 subjects had an FEV₁ above 80% of the predicted value, while only four had a Vmax_{50c} above 80% predicted. The difference in FEV₁ between assessments was within 10% for each subject; differences in sGL, Vmax_{50c}, and Vmax_{40p} were within 50%.

REPRODUCIBILITY OF DOSE-RESPONSE CURVES

The results for reproducibility derived from the one-way analysis of variance are listed in table 3. The F value and intraclass correlation show that between-subject variance was always larger than within-subject variance. The values of intraclass correlation (that is, the proportion of total variance due to real subject differences as opposed to measurement error) were satisfactory. Results of the 95% confidence interval for the overall mean over subjects and of the 95% confidence interval for subjects

based on a single determination are also given in table 3. These results are expressed as single two-fold concentration differences since doubling concentrations of histamine were nebulised (see under "Methods"). The 95% confidence interval of PC₂₀-FEV₁ based on a single determination was the observed value ±1.59 two-fold concentration difference (that is, ±3.18-fold difference). The range for PC₂₀ for FEV₁ was smaller than the ranges obtained for PC with other measurements. The confidence intervals for PC were smaller than those obtained for TC. The 95% confidence intervals of R based on a single determination were the observed values ±52–±74 (% change/log histamine concentration), depending on the measurement.

SENSITIVITY OF DIFFERENT INDICES

Comparison of TC and PC results for each measurement show that both sGL and Vmax_{40p} could detect a response to histamine at a lower concentration than FEV₁ (figs 1 and 2).

COMPARISONS BETWEEN PC, TC AND REACTIVITY

There was a significant correlation between PC results for each measurement (r values ranging from 0.73 to 0.95) and between TC values for each measurement (r values 0.61–0.79) (table 4). Comparison of reactivity results showed lower correlation coefficients and one (r sGL–R Vmax_{40p}) was not significant. TC and PC results for each measurement correlated significantly, r values varying from 0.63 to 0.91. Reactivity did not correlate significantly with PC or TC.

Fig 1 Comparisons of threshold concentration (TC) results obtained at each visit for FEV_1 , sGL , and $\dot{V}max_{40p}$. The TC for sGL and $\dot{V}max_{40p}$ detected the response at a lower concentration of histamine than TC_{FEV_1} (p values from one-way analysis of variance).

Fig 2 Comparisons of provocative concentration (PC) results obtained at each visit for FEV_1 , sGL , and $\dot{V}max_{40p}$. The PC_{40} for sGL and $\dot{V}max_{40p}$ detected a response at a lower concentration of histamine than PC_{20FEV_1} .

Discussion

In this study we assessed the reproducibility of different ways of measuring bronchoconstriction in response to inhaled histamine. We also evaluated the sensitivity of several indices used to monitor the induced bronchoconstriction. This approach stems from the proposals by Orehek²² and other workers that the complete dose-response curves should be studied rather than a single point. The use of measurements other than the FEV₁ has also been suggested.^{23–25}

We found that the concentration of histamine causing a 20% fall in FEV₁ (PC₂₀)^{6–8} was the most reproducible index, a 95% confidence interval based on a single determination being the observed value ± 1.59 two-fold concentration difference (± 3.18 -fold difference in PC₂₀). This range is larger than that described by Ryan and coworkers.⁸ We used a similar method of nebulisation and baseline FEV₁ varied less than 10% in both studies. There was a similar difference in reproducibility between two studies from the same group of investigators using a similar method of nebulisation.^{7,8} The reproducibility of airway hyperreactivity may therefore vary as a result of differences between the subjects studied. As suggested by Ryan and colleagues,⁸ this emphasises the importance of baseline data in studies which examine changes in PC_{20-FEV₁}. In our study the reproducibility of TC was less satisfactory than that of PC.

Some of the theoretical advantages of more complete characterisation of the dose-response curve²² are therefore countered by the fact that these measurements are less reproducible than is PC. The PC_{20-FEV₁} has been shown to correlate with the clinical state¹⁹ and need for medication of asthmatic subjects,²⁶ whereas the clinical relevance of reactivity is unknown. Orehek and colleagues claimed that reactivity distinguished normal from asthmatic subjects.²⁰ Their dose-response curves, however, were drawn on an arithmetic rather than the more usual scale. In another study using dose-response curves, the reactivity of normal subjects did not differ from that of asthmatic subjects.²⁷ Beaupré and Malo found no correlation between reactivity and the clinical state of asthma,¹⁹ while in our study and others^{14,20} reactivity correlated with neither PC nor TC. Finally, even if curves assessed by FEV₁, sGL, and maximal flows are linear in the range of selected changes, some have found that when assessed in terms of lung resistance they may reach a plateau in some individuals.²⁸ This may further complicate their interpretation and use.

One aim of the present work was to assess the

sensitivity of different measurements in monitoring induced bronchoconstriction. Bouhuys *et al* showed that maximum expiratory flow rates detected induced bronchoconstriction at an earlier stage than FEV₁,²³ while Fish and Kelly found that more methacholine was needed to produce a 20% fall in FEV₁ than a 35% change in specific airway conductance.²⁴ Orehek *et al* showed that FEV₁ is less sensitive than airway resistance as a measurement of provoked bronchoconstriction in asthmatic subjects because of the effect of a previous deep inspiration.²⁵ Our results are in agreement, since the PC for both $\dot{V}_{max_{40p}}$ and sGL detected bronchoconstriction at an earlier stage than did PC_{20-FEV₁}. There might thus be circumstances when the use of an index such as PC_{sGL} is preferable to PC_{20-FEV₁}. For example, the PC₃₀ for $\dot{V}_{max_{40p}}$ but not PC_{20-FEV₁} separated young symptomless cigarette smokers from non-smokers.²⁹ Nevertheless, in most circumstances requiring serial comparisons of airway responsiveness to non-allergic agents for clinical purposes, PC_{20-FEV₁} would seem to be preferable because of its better reproducibility.

Many factors need to be standardised for bronchial provocation tests,²⁹ including dose-response curves, which should seek to define the reactions of normal and asthmatic populations in the most satisfactory way, as pointed out by Orehek and Gayraud.³⁰ Few such studies have been done. In a study by Cockcroft *et al* no normal subject had a PC_{20-FEV₁} of less than 8 mg/ml, whereas all the asthmatic patients with symptoms had lower values.⁶ The same group of investigators,³¹ however, recently described a grey zone from 2 to 20 mg/ml: asthmatic subjects with such responses may have no symptoms, normal diurnal variation of peak flow rate, and no appreciable induced bronchoconstriction unless exposed to a vigorous stimulus.³² Orehek *et al* found that both TC (sensitivity) and reactivity differed between normal and asthmatic subjects, though there was considerable overlap.²⁰ In a smaller group of subjects PC_{20-FEV₁} appeared to be more specific than indices using sGL and maximum partial expiratory flow rates in distinguishing normal from asthmatic responses.³² Population studies are needed to evaluate the ability of these various indices to separate normal and asthmatic patients.

This work was supported by a grant from the Canadian Thoracic Society. AR is the recipient of a special summer grant from the Medical Research Council of Canada and RRM is a scholar of the Conseil de la Recherche en Santé du Québec. We want to thank the patients who kindly consented to take part and U Maag and L Senecal for the statistical analysis, as well as FE Hargreave and R Roberts

(McMaster University) for their helpful criticism. We also wish to express our gratitude to J Norman-din for the secretarial work.

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