Carbon in airway macrophages from children with asthma

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ABSTRACT

Background Airway macrophage (AM) phagocytosis is impaired in severe asthma. Prostaglandin (PG) E₂ and D₂ are increased in severe asthma and suppress AM phagocytic function in vitro. In this study, we sought evidence for PG-mediated impairment of phagocytosis of inhalable carbonaceous particulate matter (PM) by AM in children with severe asthma compared with mild asthmatics and healthy controls.

Methods AM were obtained from children with asthma and healthy controls using induced sputum. AM carbon area (μm²) was assessed by image analysis. In a subgroup of asthmatics, urinary PGE₂ and PGD₂ metabolites were measured by high-performance liquid chromatography, and PM exposure at the home address was modelled. Phagocytosis of PM by human monocyte-derived macrophages and rat AM was assessed in vitro by image analysis.

Results AM carbon was 51% lower in children with moderate-to-severe asthma (n=36) compared with mild asthmatics (n=12, p<0.01). PGE₂ 10⁻⁶ M, but not PGD₂ 10⁻⁶ M, suppressed phagocytosis of PM₁₀ by human macrophages in vitro (p<0.05 vs control). PGE₂ 10⁻⁶ M also suppressed phagocytosis of PM₁₀ by rat AM in vitro (p<0.01 vs control).

Conclusions Phagocytosis of inhaled carbonaceous PM by AMs is impaired in severe asthma. PGE₂ may contribute to impaired AM phagocytic function in severe asthma.

INTRODUCTION

Studies in children and adults suggest that the capacity of airway macrophages (AMs) to phagocytose inhaled material is impaired in severe asthma. First, Fitzpatrick et al.¹ reported a 50% reduction of phagocytosis of Staphylococcus aureus by AM from children with poorly controlled asthma. Second, Alexis et al.² reported that phagocytosis of opsonised yeast by AM is impaired in adults with eosinophilic asthma compared with healthy controls. Third, Huynh et al.³ reported impaired phagocytosis of apoptotic human T cells by AM from adults with severe asthma compared with either healthy controls or adults with less-severe asthma. To date, the mechanism for impaired AM phagocytosis in asthma is unclear, but severe asthma is associated with increased airway levels of eicosanoids that suppress macrophage phagocytosis in vitro. For example, prostaglandin E₂ (PGE₂) is increased in induced sputum from adults with eosinophilic asthma and inhibits the phagocytosis of opsonised Escherichia coli and sheep red blood cells by rat AM in vitro.⁴ Furthermore, PGE₂ inhibits the phagocytosis of sheep red blood cells by murine AM in vitro,⁷ and PGE₂ has been shown to decrease phagocytosis of apoptotic neutrophils by human macrophages in vitro.⁸ To date, it is unknown whether PGE₂ and PGD₂ suppress macrophage phagocytosis of fossil fuel-derived particulate matter (PM) in vitro.

Carbonaceous PM is phagocytosed in a dose-dependent manner by AM.⁹ ¹⁰ Studies in animals and humans suggest that the amount of PM in AM is a valid marker of chronic exposure to...
carbonaceous PM from fossil fuel combustion. For example, in rats, the amount of carbon in AM reflects exposure to diesel PM over a 13-week period, and in human adults the amount of carbon in AM is most strongly associated with the modelled mean exposure to inhalable PM (PM <10 microns in aerodynamic diameter; PM_{10}) over the previous 6 months. We, and other groups, have therefore used the area of carbon in AM (AM carbon) to assess the health effects of chronic exposure to PM in children and adults. The utility of AM carbon as a marker of personal exposure to PM is based on the assumption that the phagocytic capacity of AM is broadly similar between individuals. However, in a previous small pilot study, we unexpectedly found low to absent AM carbon in a small number of children with severe eosinophilic asthma and speculated that phagocytosis of AM carbon in vivo is impaired in severe asthma. In the present study, we sought to establish definitive evidence for (i) lower AM carbon in children with severe asthma compared with children with either mild asthma or healthy controls, and (ii) a role for PGE_2 and PGD_2 in suppressing phagocytosis.

**METHODS**

**Participants**

Asthmatic children were recruited from the Royal London Children’s Hospital, the University Hospitals of Leicester, the Royal Brompton Hospital (London) and schools in east London. The major source of carbonaceous PM exposure in children in these urban areas is fossil fuel-derived PM. Current asthma control was established from a review of the clinical notes or a parent-completed International Study of Asthma and Allergies in Childhood questionnaire. Asthma severity was categorised by British Thoracic Society/Scottish Intercollegiate Guidelines Network (BTS/SIGN) criteria. Children with asthma at BTS steps 1–2 were classified as ‘mild’, and BTS steps 3–5 classified as ‘moderate-to-severe’. Healthy children were recruited from east London schools and from siblings of children attending asthma clinics. A subgroup of asthmatic children recruited at the Royal London Children’s Hospital also had a urine sample obtained and exposure of their home to PM modelled using (i) the London Air Quality Toolkit (LAQT) and (ii) the home to main road distance. The LAQT is an established emissions dispersion model capable of producing annual mean pollutant concentrations at a resolution of 20 m×20 m. The LAQT models the mean exposure of children’s home addresses to PM with mean aerodynamic diameter <2.5 μm (PM_{2.5}), for annual, 7-day and 24 h PM_{2.5} prior to sputum induction (see online supplement). Children living in homes within 50 m of a main road were considered to have increased exposure to traffic-derived PM. Written informed assent and consent was obtained from children and parents (Research Ethics Committees approval reference; 08/H0704/139, 11/LO/1732 and 09/H0403/92).

**Spirometry and induced sputum**

Spirometry was performed according to standard criteria using a Microlab spirometer (Care Fusion, Kent, UK). Post bronchodilator lung function was measured 15 min after 400 μg inhaled salbutamol via a metered dose inhaler and large volume spacer. SD scores (z scores) for FEV_1 and FVC were calculated using Global Lung Initiative software. Children with asthma underwent sputum induction during a clinician-assessed period of stability. Induced sputum was not done within 1 month of an asthma exacerbation requiring oral corticosteroids. Sputum induction and processing was done as previously described using nebulised 4.5% saline via a Multisonic Profi nebuliser (Schill, Germany) (see online supplement).

**AM carbon**

The area of carbon in AM was determined as previously reported. Briefly, digital images of 50 randomly selected AM from each child were obtained at ×100 magnification, and the mean area (μm^2) of carbon determined. Analysis was blinded to asthma status. Additional details are given in the online supplement.

**Urinary PGE_2 and PGD_2 metabolites**

Urine was obtained from children at sputum induction, transported on ice and stored at −80°C within 1 h of collection. Analysis was by high-performance liquid chromatography–tandem mass spectrometry (HPLC-MS). Negative ionisation was used to detect 13,14-dihydro-15-keto-tetranor-PGE_2 (tetranor-PGE_2) and 13,14-dihydro-15-keto-tetranor-PGD_2 (tetranor-PGD_2). Metabolite concentrations were expressed in proportion to urinary creatinine (see online supplement).

**Phagocytosis of PM_{10} in vitro**

The effect of PGE_2 and PGD_2 on the phagocytosis of urban PM_{10} by human monocyte-derived macrophages and rat AM was assessed using a modification of the in vitro assay described by Aronoff et al (see online supplement). The aim was to first assess the effect of PGE_2 and PGD_2 on human macrophages at the optimal suppressive concentration (10^{-6} M) described by Aronoff et al then confirm any positive findings using rat AM. Briefly, either human monocyte-derived macrophages from healthy adult donors or rat AM were adhered to bovine serum albumin-coated coverslips and cultured with PM_{10} (10 μg/mL) collected from urban filters in Leicester city centre with or without PGE_2/PGD_2 (Sigma-Aldrich, St Louis, Missouri, USA) for 1 h. Phagocytosis of PM_{10} was determined by image analysis of 50 cells per well by an operator blinded to exposure status (described above). Experiments were done on at least six separate occasions.

**Statistical analysis**

Data are presented as median (IQR). Correlations were done by Spearman’s rank correlation (rs) and proportions compared by χ^2 test. Data between two groups were compared by either Mann–Whitney test (unpaired) or Wilcoxon matched-pairs signed-rank test (paired). Comparisons between more than two groups were done by Kruskal–Wallis test and Dunn’s multiple comparison test. Analyses were performed using Prism 5.00 for Windows (GraphPad Software, California, USA). Results were considered significant at p<0.05.

**RESULTS**

AM carbon and induced sputum eosinophil differential count (figure 1) were determined in 47 healthy controls, 13 children with asthma at BTS steps 1–2 and 36 children with asthma at BTS steps 3–5 (figure 2, table 1). Subject numbers were determined by the study duration (October 2011–December 2012, see online supplementary figure S1). For asthmatic and healthy children recruited at the Royal London Children’s Hospital and in east London schools, 195 children were approached, 129 consented to undergo a sputum induction and a technically acceptable induced sputum (IS) sample was obtained from 72 (56%). One asthmatic child with sufficient number of cells for AM carbon had insufficient leucocytes for an induced sputum differential count. All 33 asthmatic children recruited at the
Royal London Children’s Hospital provided a urine sample for PG analysis. At the University Hospitals of Leicester, 20 children with asthma and 58 healthy controls were approached for IS, 20 asthmatic children and 17 controls agreed to undergo IS. Ten randomly sampled slides from each group were assessed for AM carbon, and AM carbon determined in eight asthmatics and eight controls (see online supplementary figure S1). At the Royal Brompton Hospital, 65 children with asthma were approached for IS, 55 agreed to undergo IS. Eight randomly sampled slides were assessed and AM carbon determined in 8/8 (see online supplementary figure S1). For analysis, data from all three sites were pooled.

Compared with healthy controls, children with moderate-to-severe asthma (BTS steps 3–5), but not those with mild asthma (BTS steps 1–2), had lower postbronchodilator FEV1 z score and increased induced sputum eosinophils (table 1). In healthy controls, there was no association between AM carbon and age, and no difference in AM carbon between males and females. There was also no significant difference in AM carbon when stratified by site of recruitment in either control or asthmatic groups.

Asthma and AM carbon

Asthmatic children with moderate-to-severe asthma had 51% less AM carbon compared with both healthy controls (p<0.01) and mild asthmatics (p<0.01, table 1, figure 2). There was no difference in AM carbon between healthy controls and children with mild asthma (table 1, figure 2). In asthmatics (n=49), there was an inverse association between AM carbon and sputum eosinophils (%) (rs=−0.32, p<0.05), but not with sputum neutrophils (%) (p=0.14). AM carbon was therefore lower in asthmatic children with sputum eosinophilia defined as ≥2.5% (0.16 vs 0.25 μm², p<0.01, figure 3). AM carbon in asthmatics was also inversely associated with daily inhaled corticosteroid dose (rs=−0.31, p<0.05) and age (rs=−0.44, p<0.01).

Urinary PG metabolites and AM carbon

In the 33 asthmatic children who had a urine sample taken for PG analysis, exposure to air pollution at the home address was determined by the LAQT air pollution model for 23/33, and the home–road distance assessed in 33/33. There was no difference in modelled annual mean PM2.5, 7-day PM2.5 or 24 h PM2.5 between mild and moderate-to-severe asthmatics (table 2), and no difference in the proportion of homes within 50 m of a main road between the two asthmatic groups (table 2). There was no association between modelled PM exposure and AM carbon in 33 asthmatics who had a urine sample. Urinary PG metabolites (13,14-dihydro-15-keto-tetranor-PGE2 and 13,14-dihydro-15-keto-tetranor-PGD2) were highly correlated (rs=0.89, p<0.001), and both metabolites were increased in moderate-to-severe asthma (p<0.05, p<0.01 vs mild asthmatics for -tetranor-PGE2 and -tetranor-PGD2, figure 4, table 2). AM carbon was inversely associated with urinary metabolites of PGE2 (rs=−0.40, p<0.01) and PGD2 (rs=−0.44, p<0.01). There was an association between sputum eosinophils and urinary PGE2 and PGD2 metabolites (rs=0.35, p<0.05, and rs=0.39, p<0.05, respectively). Compatible with results from all children, AM carbon in the subgroup with a urine sample was lower in moderate-to-severe asthmatics compared with mild asthmatics (p<0.01, table 2, see online supplementary figure S2).
**Paediatric lung disease**

**Table 1** Demographics and airway macrophage carbon in healthy controls and children with asthma

<table>
<thead>
<tr>
<th></th>
<th>Healthy controls</th>
<th>Mild asthma (BTS steps 1 to 2)</th>
<th>Moderate-to-severe asthma (BTS steps 3 to 5)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>n</strong></td>
<td>47</td>
<td>13</td>
<td>36</td>
</tr>
<tr>
<td><strong>Age (year)</strong></td>
<td>9.34 (8.9 to 9.7)</td>
<td>9.41 (9.1 to 9.6)</td>
<td>11.63*** (9.8 to 13.4)</td>
</tr>
<tr>
<td><strong>Gender (M/F)</strong></td>
<td>21/26</td>
<td>11/2*</td>
<td>18/18</td>
</tr>
<tr>
<td><strong>Postbronchodilator FEV₁ (z-score)</strong></td>
<td>−0.16 (−0.89 to 0.46)</td>
<td>−0.25 (−0.78 to 0.23)</td>
<td>−1.26*** (−1.93 to −0.79)</td>
</tr>
<tr>
<td><strong>ICS dose (µg BDP equivalent/day)</strong></td>
<td>Not applicable</td>
<td>0 (0 to 150)</td>
<td>500 (400 to 750)</td>
</tr>
<tr>
<td><strong>LABA (n)</strong></td>
<td>Not applicable</td>
<td>0</td>
<td>27</td>
</tr>
<tr>
<td><strong>LTRA (n)</strong></td>
<td>Not applicable</td>
<td>0</td>
<td>16</td>
</tr>
<tr>
<td><strong>Induced sputum eosinophils (%)</strong></td>
<td>0 (0 to 0)</td>
<td>0 (0 to 0.60)</td>
<td>3.5*** (0.32 to 9.84)</td>
</tr>
<tr>
<td><strong>Induced sputum neutrophils (%)</strong></td>
<td>1.75 (0 to 22.5)</td>
<td>1.00 (0.13 to 16.8)</td>
<td>16.00** (6.5 to 39.3)</td>
</tr>
<tr>
<td><strong>Induced sputum airway macrophage carbon (µm²)</strong></td>
<td>0.37 (0.16 to 0.53)</td>
<td>0.37 (0.24 to 0.88)</td>
<td>0.19*** (0.08 to 0.26)</td>
</tr>
</tbody>
</table>

*Data are described as median (IQR). Comparison between groups by Kruskal–Wallis test and Dunn’s multiple comparisons test. There is a higher proportion of males with mild asthma (p<0.05 vs controls, χ² test). *p<0.05, **p<0.01, ***p<0.001 versus healthy controls.
BDP, beclomethasone dipropionate; BTS, British Thoracic Society; ICS, inhaled corticosteroid; LABA, long acting β₂ agonist; LTRA, leukotriene receptor antagonist.

**Phagocytosis of PM₁₀ in vitro**

Phagocytosis of PM₁₀ by human monocyte-derived macrophages was suppressed by PGE₂ 10⁻⁶ M (p<0.05 vs control, figure 5A), but not by PGD₂ 10⁻⁶ M (figure 5B). PGE₂ at 10⁻⁶ M also suppressed phagocytosis of PM₁₀ by rat AM (p<0.05, figure 6).

**DISCUSSION**

This study found, compatible with our previous pilot data,¹⁴ that AM carbon in children with moderate-to-severe asthma is approximately half that of mild asthmatics and healthy controls. It is unlikely that lower AM carbon in children with severe asthma reflects lower exposure to PM since no difference was found in either modelled exposure to PM at the home address or in the proportion of homes within 50 m of a main road in the subgroup where this was assessed. Children with more severe asthma were older than mild asthmatics, and as expected received higher doses of inhaled corticosteroids. Although there were inverse associations between AM carbon and both age and daily inhaled therapy in asthmatics, these are unlikely to be causal. First, as reported previously in healthy children,¹⁴ we found no effect of age on AM carbon in healthy controls in the present study. Second, Fitzpatrick et al¹¹ previously reported that phagocytosis of bacteria by AM in vitro is not attenuated in children receiving regular inhaled corticosteroid therapy for non-asthmatic symptoms. Third, AM phagocytic function is not impaired by β₂-adrenoreceptor agonists in vitro.²³

Lower AM carbon in children with severe asthma is compatible with previous studies of AM using other phagocytic targets. For example, Huynh et al,³ in a study of adults with severe asthma, reported a reduced number of apoptotic bodies in AM (reflecting decreased phagocytosis of apoptotic cells in vivo) compared with healthy controls, and in children with poorly controlled asthma, Fitzpatrick et al¹¹ reported an impaired capacity of AM to phagocytose *S aureus* in vitro. The lack of an effect of mild asthma on the capacity for AM phagocytosis was first suggested by Alexis et al,⁶ who reported no difference in the capacity of induced sputum AM to phagocytose opsonised yeast in mild asthmatics compared with healthy controls, and second by Lay et al,²⁵ who found that phagocytosis of inhaled particles by AM in vivo was not impaired in mild asthmatics compared with healthy controls. It is unlikely that lower AM carbon in severe asthma is due to less PM inhaled into the lower airway since studies in adults suggest increased lower airway deposition of PM in asthmatics compared with controls.²⁶ ²⁷ An increased number of AM in asthma could also result in lower AM carbon. To date, AM density in children with asthma has not been studied, but no increase in bronchoalveolar lavage AM concentration has been reported in a murine model of asthma.²⁸

The mechanism causing impaired AM phagocytosis in severe asthma is unclear. Two studies to date have assessed the association between AM phagocytosis in asthma and airway inflammation. Simpson et al²⁹ reported that uptake of apoptotic bronchial epithelial cells by AM in vitro from adults with non-eosinophilic asthma is impaired compared with adults with eosinophilic asthma. By contrast, Alexis et al²⁶ reported that phagocytosis of opsonised yeast particles by AM in vitro is attenuated in adults with eosinophilic asthma compared with a non-eosinophilic asthmatic phenotype. Compatible with Alexis et al²⁶, we found that AM carbon was indeed lower in children with eosinophilic asthma when defined either by a cut-off of ≥2.5%³⁰ or by a cut-off of ≥5%³¹ (p<0.05, see online supplementary figure S2).

PGE₂ and PGD₂ are putative mediators of impaired phagocytosis in severe asthma since they both are associated with eosinophilic asthma and directly suppress AM phagocytosis in vitro.³³ ³⁴ ³⁵ Profta et al³³ reported increased induced sputum PGE₂ in asthmatic adults with bronchodilator reversibility >15% and peak
PM10 is compatible with Rossi in the plasma during sampling. However, the urinary metabolites obtained using induced sputum, and PGs are rapidly metabolised AM, and PGE2, and to a lesser extent PGD2, suppresses phagocytosis of apoptotic neutrophils by human monocyte-derived macrophages. In the present study, the inverse association between urinary metabolites of PGE2 and PGD2 and AM carbon provides indirect evidence of a role for both mediators in suppressing the phagocytosis of inhaled fossil-derived carbon. Further support for a role of PG is provided by our in vitro studies that show, for the first time, that PGE2 10−6 M markedly suppresses phagocytosis of PM10 by human macrophages and rat AM. The lack of effect of PGD2 at 10−6 M on phagocytosis of PM10 is compatible with Rossi et al., who reported that while PGE2 10−6 M suppresses 60% phagocytosis of apoptotic cells by human monocyte-derived macrophages, PGD2 at a concentration of 10−6 M suppresses phagocytosis by only 20%. A limitation of our study is that we did not measure airway PG directly since HPLC-MS is not suitable for small sample volumes obtained using induced sputum, and PGs are rapidly metabolised in the plasma during sampling. However, the urinary metabolites measured in the present study are plausible markers of systemic levels since they result from a common metabolic pathway that includes a reduction of the double bond between C-13 and C-14 and oxidation of the hydroxyl group at C-15, producing 13,14-dihydro-15-keto PGs—metabolites that are excreted unchanged in the urine. Although we have shown that PGE2-mediated suppression of phagocytosis is biologically plausible and have provided indirect evidence for this association in vivo, we cannot exclude a role for ‘intrinsic’ changes in phagocytic capacity of AM in severe asthma. For example, Fitzpatrick et al reported that AM from children with severe asthma are characterised by a distinct molecular phenotype that does not have a clear Th1 or Th2 pattern. In summary, in children with moderate-to-severe asthma with no evidence of lower exposure to fossil fuel-derived PM, we found lower AM carbon compared with mild asthmatics and healthy controls. Indirect evidence was found of increased systemic concentrations of both PGE2 and PGD2—mediators that suppress alveolar macrophage phagocytosis in vitro. Of these, our in vitro studies suggest that PGE2 is a potent suppressor of AM phagocytosis of PM in vivo. The consequences of impaired removal of inhaled PM by AM are unknown in humans, but in mice inhaled particles that are normally non-toxic to wildtype animals are toxic to animals deficient in the receptor required for efficient AM phagocytosis of PM.

### Table 2 Prostaglandin E2 and D2 metabolites and modelled exposure to air pollution in asthmatic children in whom a urine sample was obtained

<table>
<thead>
<tr>
<th></th>
<th>Mild asthma (BTS steps 1–2)</th>
<th>Moderate-to-severe asthma (BTS steps 3–5)</th>
</tr>
</thead>
<tbody>
<tr>
<td>N</td>
<td>13</td>
<td>20</td>
</tr>
<tr>
<td>Induced sputum eosinophils (%)</td>
<td>0 (0 to 0)</td>
<td>2** (0 to 10)</td>
</tr>
<tr>
<td>Alveolar macrophage carbon (μm²)</td>
<td>0.37 (0.24 to 0.88)</td>
<td>0.19** (0.11 to 0.26)</td>
</tr>
<tr>
<td>Mean annual PM2.5 (μg/m³)</td>
<td>14.3 (14.1 to 14.6)</td>
<td>14.3 (13.7 to 14.5)</td>
</tr>
<tr>
<td>Mean 7-day PM2.5 (μg/m³)</td>
<td>15.3 (9.3 to 18.0)</td>
<td>12.4 (10.1 to 21.0)</td>
</tr>
<tr>
<td>Mean 24 h PM2.5 (μg/m³)</td>
<td>9.5 (7.9 to 10.1)</td>
<td>11.5 (8.9 to 18.1)</td>
</tr>
<tr>
<td>Home address ≤50 m from a main road (n)</td>
<td>2</td>
<td>4</td>
</tr>
<tr>
<td>Urinary 13,14-dihydro-15-keto-tetranor-PGE2 (pg/mg creatinine)</td>
<td>250 (119 to 450)</td>
<td>505* (218 to 937)</td>
</tr>
<tr>
<td>Urinary 13,14-dihydro-15-keto-tetranor-PGD2 (pg/mg creatinine)</td>
<td>225 (170 to 470)</td>
<td>779** (259 to 1149)</td>
</tr>
</tbody>
</table>

PM2.5: particulate matter <2.5 μm in aerodynamic diameter modelled at home address for 2393 children. Sputum eosinophil differential is derived from 400 induced sputum leucocytes per child. AM carbon is derived from 50 AM per child. Urine samples were obtained from all children recruited via the Royal London Hospital. Data are described as median (IQR). *p<0.05 **p<0.01 versus mild asthmatics by Mann–Whitney test.

AM, airway macrophage; BTS, British Thoracic Society; PG, prostaglandin.
Figure 6  Effect of prostaglandin E2 (PGE2) on phagocytosis of urban particulate matter <10 μm in aerodynamic diameter (particulate matter10\textsubscript{p} (PM\textsubscript{10})) by rat airway macrophages (AMs). AMs were obtained by bronchoalveolar lavage and adhered onto coverslips. Rat AMs were cultured with PGE\textsubscript{2} 10\textsuperscript{-6} M for 10 min, then incubated with PM\textsubscript{10} (10 μg/mL) for 1 h. Phagocytosis of carbon was assessed using image analysis. Paired data are from AM from the same animal. Preincubation with PGE\textsubscript{2} 10\textsuperscript{-6} M suppresses PM\textsubscript{10} phagocytosis by AM (\(p<0.01\) by Wilcoxon matched-pairs signed-rank test). Data are from 17 separate experiments in separate animals done at separate times.

Contributors  RE recruited participants, supervised sampling, analysed sputum samples and performed phagocytosis assays, and wrote the first draft of the manuscript. NM processed sputum samples, supervised phagocytosis assays and contributed to the manuscript. TR recruited participants, supervised sampling and contributed to the manuscript. DHG recruited participants and supervised sampling. ID recruited participants, and supervised sampling and spirometry. EG and LJF contributed to the manuscript. DHG recruited participants and supervised sampling. ISM participated in subject recruitment and sample collection. ISM contributed to the manuscript. EG and LJF supervised phagocytosis assays and contributed to the manuscript. DM contributed to the manuscript. EG and LJF contributed to the manuscript. NM differentiated monocytes, conducted phagocytosis assays on human cells and contributed to the manuscript. DG performed validation and contributed to the manuscript. ID recruited participants and supervised sampling. ISM contributed to the manuscript. EG and LJF contributed to the manuscript. NM recruited participants and supervised sampling. ISM contributed to the manuscript. EG and LJF contributed to the manuscript.

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Patient consent  Obtained.

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Provenance and peer review  Not commissioned; externally peer reviewed.

Data sharing statement  Additional anonymised data for analysis of carbon in individual macropahges, including stored images, as well as HPLC-MS results from urine and air pollution monitoring data are available upon request to the corresponding author.

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