Revisiting tuberculous pleurisy: pleural fluid characteristics and diagnostic yield of mycobacterial culture in an endemic area

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INTRODUCTION

Tuberculous pleurisy is the second most common form of extrapulmonary tuberculosis (TB)1 2 and a common cause of pleural effusion in endemic TB areas.3 4 A definitive diagnosis of tuberculous pleurisy usually requires mycobacterial culture of pleural fluid, pleural biopsy or other diagnostic tests.5 6 A high index of suspicion from a pleural fluid analysis is essential for diagnosis. However, the traditional concepts about the pleural fluid findings in tuberculous pleurisy mainly come from studies conducted decades ago that have not been validated in current practice.7–10 Pleural fluid has been frequently studied owing to the popular use of ultrasound, which may modify the pleural fluid findings at disease presentation.10 11 In addition, the data from previous studies on the diagnostic yields for tuberculous pleurisy have varied widely and are not consistent. These studies have suggested the following diagnostic yields: 12–70% for effusion culture,9 12 0–52% for sputum culture12 13 and 50–90% for histological examination of pleural biopsy specimens.14–17 We consider that variations in the study results are related to the patient selection criteria, the small number of cases in most of the studies and variable TB prevalence. Furthermore, advances in culture techniques have improved the yield of mycobacterial culture in this disease. Liquid culture media have been shown to have higher diagnostic yields and faster results than solid media.18 19

The objective of the present study was to provide updated information about the pleural fluid characteristics and diagnostic yields of mycobacterial culture and pleural biopsy. Differences between the cases with positive and negative effusion culture were also evaluated.
mycobacterial cultures were compared to determine the predictors of a positive effusion culture.

METHODS

Study setting
This was a retrospective cohort study conducted at the National Taiwan University Hospital and the National Taiwan University Hospital Yun-Lin Branch, two university-affiliated teaching hospitals in Taiwan. Taiwan is an endemic area of TB, with an incidence of 62–74 per 100,000 population from 2004 to 2008.\(^2^0\) The National Tuberculosis Program of Taiwan requires that all cases of culture-confirmed TB or clinically suspicious TB should be reported to the Taiwan Centers for Disease Control. Such cases are also recorded in the TB registration databases of the two study hospitals. Because of the high incidence of TB, effusion mycobacterial culture is a routine examination for all undiagnosed pleural effusions regardless of the clinical presentation and pleural fluid findings in both study hospitals. This study was approved by the Research Ethics Committee of the National Taiwan University Hospital.

Diagnosis of tuberculous pleurisy
Pleural effusion was diagnosed as tuberculous according to the following criteria: positive mycobacterial culture in pleural fluid or pleural biopsy tissue samples; granulomatous inflammation in pleural biopsy tissue samples; positive mycobacterial culture of sputum samples and pleural effusion that were resolved with anti-TB treatment; or clinically suspected tuberculous pleurisy and pleural effusion that were resolved with anti-TB treatment. In the fourth criterion, ‘clinical suspicion’ means that these patients had no systemic diseases, and were excluded from malignancy and other pleural diseases. Moreover, there were suggestive clues of TB from contact history, radiographical findings or clinical presentation.

We obtained and merged the lists of patients with tuberculous pleurisy or other tuberculosis diseases with concurrent pleural effusion from the following three sources: our TB registration database; our mycobacteriology laboratory database to identify cases with a positive TB culture in any specimen; and our pathology database to identify cases with granulomatous inflammation in pleural tissue specimens. The study period was from January 2004 to June 2009. The identified cases were further evaluated if they satisfied one of the four criteria. All cases that failed to satisfy one of these criteria were excluded from the study. Patients with TB empyema, defined as pus-like appearance of pleural fluid or positive acid-fast bacilli (AFB) smear on pleural fluid, were also excluded.

Data extraction
Medical records were reviewed to obtain data, including age, gender, comorbidities, date of the first visit for pleural effusion, date of initiation of anti-TB treatment and date of the last visit. The laboratory data included initial blood cell counts, renal function tests, liver function tests, protein levels, lactate dehydrogenase (LDH) levels, glucose levels in serum and pleural fluid, and white blood cell and differential cell counts of pleural fluid. Chest radiographs were reviewed by a pulmonologist and a radiologist to assess the initial radiographical presentation and response to treatment. The results of the AFB smears and the mycobacterial cultures of pleural fluid and sputum were obtained.

Bacteriological procedures
The AFB smears were performed with auramine-rhodamine fluorochrome and examined using standard procedures.\(^2^1\) The specimens for the mycobacterial cultures were processed using the standard N-acetyl-L-cysteine and sodium hydroxide method. The mycobacterial cultures were performed by inoculating 0.5 ml of the processed specimens into MGIT 960 culture tubes according to the manufacturer’s instructions (BACTEC Mycobacteria Growth Indicator Tube 960 System, BD Diagnostic Instrument Systems, Sparks, Maryland, USA). The fluorometric BACTEC technique was used to detect positive tubes for a maximum of 6 weeks. For the positive tubes, a subculture was made on Middlebrook 7H11 selective agar (Remel Inc, Lexena, Kansas, USA).

Statistical analysis
The categorical variables were compared using the \(\chi^2\) test or Fisher’s exact test, as appropriate, while the continuous variables were compared using t tests or the Mann–Whitney U test. A multivariate logistic regression analysis was conducted to identify the predictors of a positive effusion mycobacterial culture. The analysed variables included age, gender, underlying diseases, effusion levels of protein, LDH levels, glucose levels, white blood cell counts and the lymphocyte percentages of the effusions. Variables with a p value of <0.2 in the univariate regression were added in a stepwise manner and selected to determine the final model. A generalised additive model was used to assess the linearity of the relationship between effusion culture and parameters of interest.\(^2^2\) The time-to-event curves for each variable were generated using the Kaplan–Meier method and compared by the log-rank test. The data were analysed using Stata software, V.10 (StataCorp).

RESULTS

Cases of tuberculous pleurisy
There were 3263 patients with definite or suspicious TB in the study period. Of these patients, 792 patients had tuberculous pleurisy or another form of TB disease with concurrent pleural effusion. Three patients with positive AFB smears on their pleural fluids were considered to have tuberculous empyema and excluded for analysis. Among the 789 patients, 382 patients were diagnosed as having tuberculous pleurisy, including 241 with effusion cultures positive for \textit{Mycobacterium tuberculosis} and 141 with negative effusion cultures. In the 141 patients with negative effusion cultures, 48 received a diagnosis by pleural biopsy, 45 had positive sputum cultures and had pleural effusion that resolved with anti-TB treatment, and 48 were clinical suspicions and had pleural effusion that resolved with anti-TB treatment. Table 1 summarises the characteristics of the 382 patients.

Pleural fluid findings
Table 2 shows the laboratory data for these patients. In general, the patients had no leukocytosis but had an elevated serum level of LDH. Protein-rich exudate was a common feature of the pleural fluid analysis. Figure 1A shows the distribution of the effusion lymphocyte percentages. A lymphocyte percentage of <50% was observed in 17% of the patients. The effusion cell differential of the three excluded patients with TB empyema showed 93%, 92% and 6% neutrophils. The case with 6% neutrophils had been treated with levofloxacin before thoracentesis.

Compared with the culture-negative effusions, the culture-positive effusions had a lower lymphocyte percentage (median 92% vs 78%, p<0.001). The probability of a positive effusion culture was negatively associated with the effusion lymphocyte...
Table 1 Characteristics of 382 patients with tuberculous pleurisy

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Pleural effusion culture, n (%)</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Positive, n=241</td>
<td>Negative, n=141</td>
</tr>
<tr>
<td>Age (mean ± SD)</td>
<td>62±20</td>
<td>63±21</td>
</tr>
<tr>
<td>Gender (men/women)</td>
<td>270/112</td>
<td>173/68</td>
</tr>
<tr>
<td>Visit to diagnosis, median (IQR)</td>
<td>15 (7–29)</td>
<td>17 (8–30)</td>
</tr>
</tbody>
</table>

Underlying diseases

- COPD or current smoker: 92 (24)
- Diabetes mellitus: 73 (19)
- Cancer: 54 (14)
- Renal replacement therapy: 33 (9)
- Heart failure: 23 (6)
- Past history of tuberculosis: 17 (4)
- Liver cirrhosis: 16 (4)
- Autoimmune diseases: 10 (3)
- HIV seropositive: 8 (2)
- Post transplantation: 4 (1)

Effusion sites

- Right: 237 (62)
- Left: 149 (39)

p Values were calculated using χ² or Fisher’s exact test unless otherwise specified.
*Student t test.
†Mann–Whitney U test.
‡Bilateral tuberculous pleurisy in four cases.
COPD, chronic obstructive pulmonary disease.

percentage (figure 1B). The multivariate logistic regression model identified four independent predictors of the effusion culture results, including cancer as an underlying disease (OR 2.66; 95% CI 1.14 to 6.19), effusion lymphocyte percentage (OR 0.97; 95% CI 0.96 to 0.99), effusion protein level (g/dl) (OR 0.6; 95% CI 0.45 to 0.79) and effusion LDH level (U/litre) (OR 1.001; 0.97; 95% CI 0.96 to 0.99), effusion protein level (g/dl) (OR 0.6; 95% CI 0.45 to 0.79) and effusion LDH level (U/litre) (OR 1.001; 95% CI 1.00 to 1.002).

Diagnostic yields

The yields of effusion culture, sputum culture and pleural biopsy in the diagnosis of tuberculous pleurisy are summarised in table 3. The diagnostic yields were 65% and 48% for the effusion and sputum cultures, respectively. A combination of the effusion and sputum cultures achieved a diagnostic sensitivity of 79%. In contrast, the diagnostic yield was 74% for histological examination of pleural biopsies.

Outcome

Figure 2 shows the 180-day survival curve of the patients with positive and negative effusion cultures. The probability of survival significantly differed according to the effusion culture results (log-rank test, p<0.001). In the patients with positive effusion cultures, there were 21 (9%) deaths before the time of diagnosis.

DISCUSSION

This study reveals several important findings regarding the diagnosis of tuberculous pleurisy. The diagnostic yield of

Table 2 Biochemical characteristics and cells differential of 382 tuberculosis pleurisy cases

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>n</th>
<th>Mean±SD</th>
<th>Median (IQR)</th>
<th>Effusion culture</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Positive, mean±SD</td>
<td>Negative, mean±SD</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>or median (IQR)</td>
<td>or median (IQR)</td>
</tr>
<tr>
<td>Blood test</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>WBC (µ/l)</td>
<td>349</td>
<td>7549±3150</td>
<td>6960 (5700–8530)</td>
<td>7755±3536</td>
<td>7189±2296</td>
</tr>
<tr>
<td>Platelet (K/µ/l)</td>
<td>345</td>
<td>282±114</td>
<td>272 (208–347)</td>
<td>262±115</td>
<td>315±106</td>
</tr>
<tr>
<td>Protein (g/dl)</td>
<td>171</td>
<td>6.8±1</td>
<td>6.8 (6.1–7.5)</td>
<td>6.6±1</td>
<td>7.1±1</td>
</tr>
<tr>
<td>LDH (U/litre)</td>
<td>149</td>
<td>565±271</td>
<td>489 (413–644)</td>
<td>509 (416–660)</td>
<td>453 (379–578)</td>
</tr>
<tr>
<td>Pleural effusion test</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>WBC (µ/l)</td>
<td>369</td>
<td>2275±2000</td>
<td>1800 (900–3100)</td>
<td>1600 (800–3060)</td>
<td>1900 (1000–3100)</td>
</tr>
<tr>
<td>Lymphocyte %</td>
<td>367</td>
<td>76±25</td>
<td>84 (64–95)</td>
<td>78 (55–91)</td>
<td>92 (82–97)</td>
</tr>
<tr>
<td>Neutrophil %</td>
<td>367</td>
<td>15±21</td>
<td>5 (1–19)</td>
<td>9 (2–28)</td>
<td>2 (1–8)</td>
</tr>
<tr>
<td>Protein (g/dl)</td>
<td>362</td>
<td>4.8±1</td>
<td>5 (4.2–5.4)</td>
<td>4.6±1</td>
<td>5±0.8</td>
</tr>
<tr>
<td>LDH (U/litre)</td>
<td>362</td>
<td>988±1259</td>
<td>680 (455–1055)</td>
<td>849 (500–1313)</td>
<td>550 (389–763)</td>
</tr>
<tr>
<td>Glucose (mg/dl)</td>
<td>325</td>
<td>105±53</td>
<td>97 (77–124)</td>
<td>101±56</td>
<td>112±49</td>
</tr>
<tr>
<td>Effusion-serum ratio</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Protein (effusion/serum)</td>
<td>170</td>
<td>0.7±0.1</td>
<td>0.7 (0.6–0.8)</td>
<td>0.7±0.1</td>
<td>0.7±0.1</td>
</tr>
<tr>
<td>LDH (effusion/serum)</td>
<td>144</td>
<td>1.9±1.6</td>
<td>1.3 (0.9–2.3)</td>
<td>2.1±1.8</td>
<td>1.4±1.1</td>
</tr>
</tbody>
</table>

p Values were calculated using Student t test unless otherwise specified.
*Mann–Whitney U test.
LDH, lactate dehydrogenase; WBC, white blood cell.
Figure 1 Lymphocyte percentage in tuberculous pleurisy. (A) Distribution of cases by effusion lymphocyte percentage. (B) Generalised additive model analysis of relationship between lymphocyte percentage of pleural fluids and effect on probability of culture positivity of pleural fluids, which equals ln(p/1− p). Data are presented as fitted regression (line) and 95% CI (grey area).

The lymphocyte percentage in pleural fluids was found to be an important marker for the diagnosis of tuberculous pleurisy. Furthermore, a high yield of pleural fluid lymphocytes was associated with the probability of a positive effusion culture. Prospective studies are needed to clarify this issue. Furthermore, previous studies suggested that most patients with tuberculous pleurisy have more than 90% lymphocytes in their pleural fluid. A highly cited study reported that only 6.7% of patients had fewer than 50% of lymphocytes in their pleural fluid. In contrast, the degree of lymphocyte predominance in our patients was lower than those of previous studies. This discrepancy may be partly related to the popular use of ultrasound for thoracentesis and early effusion studies.

Tuberculous pleurisy may resolve spontaneously without treatment, but the patient frequently develops active TB at a later date. A pleural effusion that resolves with anti-TB therapy rather than undergo invasive diagnostic procedures tends to improve the yield of pleural fluid lymphocytes. However, removing these cases does not alter the main study findings but increases sensitivity of all diagnostic methods (online supplementary tables 1 and 2). In TB endemic areas, some patients with suspected tuberculous pleurisy prefer to receive a therapeutic trial with anti-TB therapy rather than undergo invasive pleural biopsy after malignancy and other common diseases have been excluded.

Mycobacterial culture of sputum has been suggested in the management of suspected tuberculous pleurisy, though the diagnosis of TB pleurisy in this study cohort, which is much higher than the yield of less than 35% reported in most of the previous studies. The combination of the effusion and sputum cultures contributed to a good diagnostic yield of up to 79% compared with a yield of 74% in the histological examination of pleural biopsies. In addition, our study provides updated data about pleural fluid findings in tuberculous pleurisy (figure 1A and table 2) and shows that the lymphocyte percentage of pleural fluid was negatively associated with the probability of a positive effusion culture (figure 1B). Furthermore, the differences between effusion culture-positive and culture-negative tuberculous pleurisy were highlighted in our study.

Table 3 Diagnostic yields of effusion culture, sputum culture and pleural biopsy for tuberculous pleurisy

<table>
<thead>
<tr>
<th>Diagnostic procedures</th>
<th>n (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Positive effusion culture</td>
<td>241/382 (63)</td>
</tr>
<tr>
<td>Positive sputum culture</td>
<td>124/241 (51)</td>
</tr>
<tr>
<td>Negative sputum culture</td>
<td>89/241 (37)</td>
</tr>
<tr>
<td>Sputum culture not performed</td>
<td>28/241 (12)</td>
</tr>
<tr>
<td>Negative effusion culture</td>
<td>141/385 (37)</td>
</tr>
<tr>
<td>Positive sputum culture</td>
<td>60/141 (43)</td>
</tr>
<tr>
<td>Negative sputum culture</td>
<td>64/141 (45)</td>
</tr>
<tr>
<td>Sputum culture not performed</td>
<td>17/141 (12)</td>
</tr>
<tr>
<td>Positive sputum culture</td>
<td>184/382 (48)</td>
</tr>
<tr>
<td>Positive sputum culture/sputum culture performed</td>
<td>184/337 (55)</td>
</tr>
<tr>
<td>Positive AFB smear/positive sputum culture</td>
<td>39/184 (21)</td>
</tr>
<tr>
<td>Combining effusion and sputum cultures</td>
<td>301/382 (79)</td>
</tr>
<tr>
<td>Pleural biopsy performed</td>
<td>149/382 (39)</td>
</tr>
<tr>
<td>Observation of granuloma</td>
<td>110/149 (74)</td>
</tr>
<tr>
<td>Positive tissue culture/tissue culture performed</td>
<td>41/104 (39)</td>
</tr>
<tr>
<td>Either positive tissue culture or histology</td>
<td>112/149 (75)</td>
</tr>
</tbody>
</table>

AFB, acid-fast bacillus.

Figure 2 Kaplan–Meier estimates of survival for patients with positive and negative effusion cultures.
sensitivity of sputum culture varied widely from 0% to 52% in previous studies.\textsuperscript{12} \textsuperscript{13} Our results showed that 48% of patients had positive sputum cultures, which is comparable to previous studies with larger case numbers.\textsuperscript{6} \textsuperscript{12} Frequent lung parenchymal involvement in patients with tuberculosis pleurisy shown in a CT study illustrated the rationale of using sputum cultures for the diagnosis of tuberculosis pleurisy.\textsuperscript{26} Therefore, the combination of effusion and sputum cultures is a reasonable approach in the diagnosis of tuberculosis pleurisy and may achieve a good diagnostic yield. Our study documented the high yield of this diagnostic approach. Another advantage of this approach over histological examination and other novel molecular tests is that it allows for susceptibility testing, which is important in areas with a high rate of drug resistance.

The diagnostic yield of the effusion cultures in our study was much higher than the reported values of 7–53% in most of the previous studies.\textsuperscript{7} \textsuperscript{12} \textsuperscript{14} \textsuperscript{17} This discrepancy may be partly due to the difference in culture methods. Studies with low effusion culture yields mostly used solid culture media. However, liquid culture systems are known to have higher yields and faster results than those using solid media, especially for paucibacillary diseases, such as extrapulmonary TB.\textsuperscript{17} \textsuperscript{18} In addition, selection bias contributed to the varying yields of effusion culture in previous studies. Many prospective studies of tuberculosis pleurisy excluded patients with multiple comorbidities and poor performance. However, patients who are immunocompromised are more likely to have a higher mycobacterial burden in the pleural cavity and, therefore, have a higher probability of a positive effusion culture.\textsuperscript{19} These exclusion criteria consequently decrease the yield of effusion culture. In addition, if neutrophilic pleural effusion is not considered indicative of tuberculosis pleurisy and is excluded in a study, the sensitivity of the effusion culture also decreases. The negative association between lymphocyte percentage and effusion culture was well demonstrated in our study. Compared with previous studies, the selection bias was reduced in our study because of the comprehensive enrolment of all cases based on the backgrounds of a universal mycobacterial culture for all undiagnosed pleural effusions and a high index of suspicion due to endemic TB.

Our study also adds new insight to the pathogenesis of tuberculosis pleurisy. The classical pathogenesis model, a rupture of subpleural caseous foci followed by a delayed hypersensitivity reaction to \textit{M tuberculosis} antigens,\textsuperscript{30} \textsuperscript{32} indicates two primary features of tuberculosis pleurisy, including lymphocyte predominance and a low yield of effusion culture. Some investigators even suggested that tuberculosis pleurisy was a disease caused by a hypersensitivity reaction rather than an infection of the pleura.\textsuperscript{14} \textsuperscript{23} However, this hypothesis can be challenged if effusion lymphocytosis and a low yield of effusion culture are not universally present in every case of tuberculosis pleurisy. Based on the findings of our study, we propose a new concept for tuberculosis pleurisy. We hypothesise that tuberculosis pleurisy is a continuous spectrum of disease (illustrated by online supplementary figure 1) rather than the two distinguishing clinical phenotypes of a delayed hypersensitivity reaction or true pleural infection.\textsuperscript{6} \textsuperscript{33} Tuberculous empyema represents an extreme condition, with a high percentage of neutrophils and a high mycobacterial load in pleural fluids. Previous studies have reported the presence of neutrophilic leukocytosis in tuberculosis empyema and an increased neutrophil percentage in culture-positive tuberculosis pleural effusions.\textsuperscript{34} \textsuperscript{35} Tuberculous para-pneumonic effusion represents the common paucibacillary presentation of tuberculosis pleurisy. The effusion lymphocyte percentage is negatively associated with the pleural mycobacterial burden and the likelihood of a positive effusion culture. It should be noted that this hypothesis is proposed solely based on the findings of our study and needs additional studies to test it.

The patients with positive effusion cultures were associated with a higher mortality rate in this study cohort (figure 2). However, a positive effusion culture was also associated with higher rates of cancer, renal failure, liver cirrhosis and a lower serum protein level. It is not clear whether the higher mortality rate in patients with positive effusion cultures was caused by a higher intrapleural mycobacterial burden or the poor health status of the host in this retrospective study. Further studies are needed to clarify the causality.

Our study has several limitations. This study was a retrospective analysis, and the effect of missing data on the results is unknown. In addition, this study was conducted in a TB endemic area and the results may not be generalisable to other areas.

In summary, our study yields several important findings that contribute to the current understanding of the diagnosis of tuberculosis pleurisy. Using liquid culture media, the sensitivity of effusion culture was higher than has been previously reported. The combination of effusion and sputum cultures offers a good diagnostic yield and may be an alternative approach to diagnose tuberculosis pleurisy in areas where pleural biopsy and other novel diagnostic tests are limited. Although further validation is needed, the concept of a continuous spectrum of tuberculosis pleurisy offers a new and more practical consideration for this disease.

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\section*{Contributors}

SYR, YCC, JYW: study concept and design, analysis of data and review of manuscript. LNL, CJY: review of manuscript. S T Yang, for assistance with the study, and the members of the Taiwan Anti-Mycobacteria Investigation (TAMI) group for advice on interpretation of data.

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\section*{Competing interests}

None.

\section*{Ethics approval}

The Research Ethics Committee of the National Taiwan University Hospital.

\section*{Provenance and peer review}

Not commissioned; externally peer reviewed.

\section*{Data sharing statement}

We would like to share the original data of our study. Dr J Y Wang is the corresponding author and has full access to all study data.

\section*{REFERENCES}


Tuberculosis

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**Supplementary Material**

Supplementary material can be found at: [http://thorax.bmj.com/content/suppl/2012/03/20/thoraxjnl-2011-201363.DC1](http://thorax.bmj.com/content/suppl/2012/03/20/thoraxjnl-2011-201363.DC1)

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