Systemic elastin degradation in chronic obstructive pulmonary disease

John D Maclay, David A McAllister, Roberto Rabinovich, Imran Haq, Scott Maxwell, Stephen Hartland, Martin Connell, John T Murchison, Edwin J R van Beek, Robert D Gray, Nicholas L Mills, William MacNee

ABSTRACT

Background Development of emphysema and vascular stiffness in chronic obstructive pulmonary disease (COPD) may be due to a common mechanism of susceptibility to pulmonary and systemic elastin degradation.

Objectives To investigate whether patients with COPD have evidence of systemic elastin degradation in the skin.

Methods The authors measured cutaneous elastin degradation using immunohistochemistry (percentage area of elastin fibres) in sun-exposed (exposed) and non-sun-exposed (non-exposed) skin biopsies in 16 men with COPD and 15 controls matched for age and cigarette smoke exposure. Quantitative PCR of matrix metalloproteinase (MMP)-2, -9, -12 and tissue inhibitor of metalloproteinases (TIMP)-1 mRNA and zymography for protein expression of MMP-2 and -9 were performed on homogenised skin. Arterial stiffness and emphysema severity were measured using carotid-femoral pulse wave velocity and quantitative CT scanning.

Results Skin elastin degradation was greater in exposed and non-exposed skin of patients with COPD compared with controls (exposed, mean (SD); 43.5 (12.1)% vs 26.3 (6.9)%), p<0.001; non-exposed 22.4 (5.2)% vs 18.1 (4.3%), p=0.02). Cutaneous expression of MMP-9 mRNA and proMMP-9 concentrations was increased in exposed skin of COPD patients (p=0.004 and p=0.02, respectively) and was also associated with increased skin elastin degradation (r=0.62, p=0.001 and r=0.47, p=0.01, respectively). In the entire cohort of ex-smokers, cutaneous elastin degradation was associated with emphysema severity, FEV1, and pulse wave velocity.

Conclusions Patients with COPD have increased skin elastin degradation compared with controls, which is related to emphysema severity and arterial stiffness. Systemic elastin degradation due to increased proteolytic activity may represent a novel shared mechanism for the pulmonary, vascular and cutaneous features of COPD.

INTRODUCTION

Chronic obstructive pulmonary disease (COPD) is defined as a condition with extra-pulmonary effects and systemic consequences, including cardiovascular disease. Systemic vascular dysfunction is emerging as a specific mechanism that may contribute to the increased cardiovascular risk in COPD. We have demonstrated that systemic arterial stiffness correlates with the severity of emphysema in COPD patients. Thus, we postulated that elastin degradation in the lung parenchyma and systemic arterial walls is the mechanistic link between the pulmonary and systemic vascular manifestations of COPD.

Elastin is an essential structural protein in the lungs, maintaining airway patency and ensuring elastic recoil. Emphysematous changes are, in part, due to elastin fibre breakdown which causes parenchymal destruction, reduced lung compliance and airway collapse. An imbalance of proteases and antiproteases in COPD, with a net increase in elastolytic activity, is thought to play an important role in the pathogenesis of emphysema, and recent work has focused on the role of matrix metalloproteinases (MMPs) and their equivalent antiproteases. MMPs have also been implicated as a pathogenic mechanism for the increased arterial stiffness that occurs with ageing, caused in part by elastin breakdown in the arterial wall.

The hypothesis that COPD may be associated with both local lung and systemic abnormalities in connective tissue was addressed by Smith et al in 1967. Having shown abnormalities in the dermis of patients with obstructive lung disease, primarily in collagen, the authors postulate that these changes may reflect a primary defect of connective tissue in...
the body as a whole. However, the smoking history of the subjects is not reported. More recent studies have suggested that elastin degradation may occur outside the pulmonary compartment in COPD. Lee et al. have reported increased circulating elastin antibodies in the plasma of emphysematous compared with non-emphysematous subjects. Furthermore, Patel et al. reported that skin wrinkling was associated with emphysema in smokers, and postulated that this may be due to changes in collagen and elastin.

We hypothesise that the increase in elastin degradation and protease–antiprotease imbalance responsible for emphysema is present in systemic tissue, causing increased arterial stiffness and skin wrinkling. We therefore examined skin biopsies from patients with COPD and matched control ex-smokers for evidence of elastin degradation.

**METHODS**

Thirty-five male ex-smokers were recruited for a study of vascular function from primary care and a hospital respiratory outpatient clinic. Of these, 16 men with COPD and 15 male controls agreed to undergo skin biopsies. Inclusion and exclusion criteria of these study subjects have been previously published.

In brief, the subjects with COPD had a history consistent with the disease and postbronchodilator spirometric evidence of airflow limitation (FEV₁ to FVC ratio <0.7), while the controls had normal spirometry and no symptoms of respiratory disease. All subjects were ex-smokers of at least 6 months and had a greater than 10 pack year smoking history. They had no history of vascular disease or any other systemic inflammatory condition, and were not prescribed statins, oral corticosteroids or long-term oxygen therapy. Prebronchodilator and postbronchodilator spirometries were measured (Alpha Spirometer; Vitalograph, Buckingham, UK) according to American Thoracic Society/European Respiratory Society standards. Serum C reactive protein concentrations were measured using a highly sensitive immunonephelometric assay (Behring BN II nephelometer; Global Medical Instruments, Ramsay, Minnesota, USA). All studies were conducted at the Wellcome Trust Clinical Research Facility, Royal Infirmary, Edinburgh, approved by Lothian Regional Ethics Committee and conducted with the written informed consent of all participants.

**Image processing and analysis**

A single section was used for each subject. Images of each section were obtained using 10× magnification and processed with QCapture Pro software (Media Cybernetics, Bethesda, Maryland, USA). Consecutive images were merged using Adobe Photoshop CS4 (San Jose, California, USA) to enable analysis of the entire section. As elastin fibres degrade, they thicken and occupy a greater relative area, which can be quantified to assess elastin degradation in the skin. This phenomenon is called elastosis and is extensively reported in ageing and in response to UV light exposure. In a review of photoageing, Gilchrest described the histological hallmark of dermal elastosis as ‘thickened, tangled and granular amorphous elastin fibres’. We measured the area covered by elastin fibres in the reticular dermis using semiautomated quantification with ImageJ (NIH, Bethesda, Maryland, USA) in a random order by an investigator blinded to the subject identities (JM). This was expressed as a percentage of the total area measured.

In order to validate our semiautomated elastin degradation measurements, an investigator (JM) blinded to the identities of the subjects ranked each section from 1 (mild elastin degradation) to 4 (severe elastin degradation). This visual scoring was concordant with the semiautomated measurements (Spearman’s rank; r=0.86, p<0.001). Using visual scoring of elastin degradation as the outcome measure, we again showed higher elastin...
degradation in the COPD group in comparison with the control subjects (p<0.001).

**Protein and RNA extraction**

Samples of skin tissue were homogenised in lysis buffer and an EDTA-free antiprotease cocktail (Roche, Welwyn Garden City, UK) using the Qiagen tissue lyser (Crawley, UK). Protein quantification was performed using a bicinchoninic acid assay. RNA was isolated from skin following homogenisation using the RNeasy Mini Kit (Qiagen, Crawley, UK). Quantity and quality of RNA were calculated using a Nanodrop spectrophotometer (Nanodrop, Wilmington, Delaware, USA).

**Gene expression**

Total RNA (0.1 μg) was reverse transcribed in a reaction mix containing 4 μl of 5× reaction buffer, 2 μl dithiothreitol (0.1 M), 0.5 μl dNTPs (10 mM), 2 μl of random hexamers, 0.5 μl (200 U) Superscript II reverse transcriptase and 0.5 ml (40 U) RNase inhibitor (Amersham, Buckinghamshire, UK). A quantity of 0.1 μl of undiluted cDNA was used per reaction to assess expression of MMP-2, -9, -12 and tissue inhibitor of metalloproteinase (TIMP)-1; the primer and probe sets were custom probes (University of Edinburgh). These predefined primers are tested and standardised for reproducible expression analysis. Primer and cDNA were added to the TaqMan universal PCR master mix (Applied Biosystems) containing all the reagents for PCR. Experiments were performed in duplicate. Quantification of the PCR products was performed with an ABI prism 7500 (Applied Biosystems) using a relative standard curve made up of cDNA of a known quantity. The mean quantity of target gene expression (MMP-2, -9 and TIMP-1) was determined from the generated standard curve. The correlations were r=0.97, r=0.99 and r=0.99, respectively, indicating good reproducibility of these measurements. All samples were normalised using an internal standard reference gene, 18S (r=0.99 for standard curve). All data are presented as a quotient relative to the control data.

**Gelatin zymography**

Sample separation was performed using precast 10% Tris-Glycine gels (7.5%), containing 0.1% gelatin (Invitrogen, Paisley, UK), in 2× non-reducing sample buffer at 120 V and Novex Tris Glycine SDS running buffer (Invitrogen, Paisley, UK). SDS was removed using zymogram renaturing buffer (Invitrogen, UK) for 60 min at room temperature. The gels were incubated overnight at 57°C in developing buffer (Invitrogen) and then stained with 0.1% Coomassie blue in 40% methanol and 10% acetic acid and destained until clear proteolytic bands appeared. Densitometry was performed using ImageJ (NIH). Normalisation between gels was performed using 10 ng of recombinant MMP-2 (R&D Systems, Abingdon, UK) before adjusting for total protein, allowing comparison between gels.

**CT scanning**

All 15 healthy controls and 13 subjects with COPD agreed to low-dose CT scanning performed at full inspiration using a 320-multidetector row CT scanner (Aquilion One, Toshiba Medical Systems, Nasushiobara, Japan). Non-contrast enhanced CT scans were obtained at 120 kV, 100 mAs during coached inspiratory breath-hold to total lung capacity. Images were reconstructed at 0.5-mm intervals with 0.5-mm thick slices using an FC-08 filter. The histogram of CT Hounsfield Units (HU) was corrected for the air offset in Toshiba CT scanners (about −985 HU instead of the nominal −1000 HU with the FC03 filter) using an extra-thoracic air calibration method. Emphysema was quantified using inhouse software as percentage low attenuation voxels below −910 and −950 HU (% LAA-910 and %LAA-950). While the latter, higher threshold distinguishes well between those subjects with COPD and healthy controls, using the former, lower cut-off allows more uniform distribution of emphysema across this cohort of former smokers. We therefore performed an analysis using both measures of lung density.

**Arterial stiffness**

We measured arterial stiffness using carotid-femoral pulse wave velocity as per the Expert Consensus Document on Arterial Stiffness in these cases and controls and presented the findings in a previous study. Carotid-femoral (aortic) pulse wave velocity is increased with increasing arterial stiffness.

**Data analysis**

COPD and control groups were compared using Student’s unpaired t tests or an appropriate non-parametric alternative for variables not normally distributed. Associations among skin, postbronchodilator spirometry, emphysema severity using % LAA-910 and arterial stiffness measures were explored using Pearson correlations and associations with emphysema severity using %LAA-950 were explored using Spearman’s rank correlation. C reactive protein was log-transformed for all analysis. All analyses were performed using SPSS V18.0 and Graphpad prism (La Jolla, California, USA). Statistical significance was taken at p<0.05.

**RESULTS**

Men with COPD and controls were well matched for age and smoking history. Patients with COPD had a range of severity of airflow limitation (GOLD stage 1–4), were taking medication consistent with their disease severity and had considerably more emphysema than controls (table 1).

Elastin degradation was greater in sun-exposed skin (exposed) in patients with COPD in comparison with controls (mean (SD); 43.5 (12.1)% vs 26.3 (6.9)% p<0.001). Elastin degradation was also greater in non-sun-exposed skin (non-exposed) in patients with COPD (mean (SD); 22.4 (5.2)% vs 18.1 (4.3)%, p=0.02; table 2; figure 2).

MMP-9 mRNA expression and proMMP-9 protein concentrations were higher in the exposed skin of COPD patients in comparison with controls (table 2). There was little detectable MMP-9 activity present in any subject. There was greater MMP-2 and TIMP-1 mRNA expression in the exposed skin of COPD patients, but these differences were not statistically significant. MMP-2 and -9 mRNA expression were positively associated with elastin degradation in exposed skin (MMP-2 r=0.55, p=0.002; MMP-9 r=0.62, p<0.001; figure 3). Differences in MMP expression between COPD and controls were not seen in the non-exposed skin. There was little MMP-12 mRNA expression in the skin of any of the subjects.

In exploratory analyses examining the whole cohort as a group of ex-smokers, we found that exposed and non-exposed skin elastin degradation were both related to FEV1 (exposed r=−0.56, p=0.001, non-exposed r=−0.45, p=0.01) and to emphysema severity (%LAA-910: exposed r=0.54, p=0.003, non-exposed r=0.50, p=0.007; %LAA-950: exposed r=0.46, p=0.01, non-exposed r=0.54, p=0.003; figure 4). Elastin degradation in non-exposed skin was associated with aortic pulse wave velocity...
Table 1 Subject characteristics

<table>
<thead>
<tr>
<th></th>
<th>COPD</th>
<th>Control</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>n</td>
<td>16</td>
<td>15</td>
<td></td>
</tr>
<tr>
<td>Age, years</td>
<td>65 (5.2)</td>
<td>64 (5.7)</td>
<td>0.83</td>
</tr>
<tr>
<td>Body mass index, kg/m²</td>
<td>26.5 (3.8)</td>
<td>27.9 (3.8)</td>
<td>0.33</td>
</tr>
<tr>
<td>Cigarette smoke exposure, pack years*</td>
<td>36 (24–54)</td>
<td>34 (28–46)</td>
<td>0.56</td>
</tr>
<tr>
<td>Inhaled medications, number of subjects (%)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Short acting beta agonist</td>
<td>15 (94%)</td>
<td>16 (94%)</td>
<td></td>
</tr>
<tr>
<td>Anticholinergic</td>
<td>10 (63%)</td>
<td>10 (63%)</td>
<td></td>
</tr>
<tr>
<td>Long acting beta agonist</td>
<td>2 (12%)</td>
<td>2 (12%)</td>
<td></td>
</tr>
<tr>
<td>Inhaled corticosteroid</td>
<td>1 (6%)</td>
<td>1 (6%)</td>
<td></td>
</tr>
<tr>
<td>ICS/LABA combination</td>
<td>13 (81%)</td>
<td>13 (81%)</td>
<td></td>
</tr>
<tr>
<td>Postbronchodilator spirometry, %</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>FEV₁ predicts</td>
<td>47.8 (18.5)</td>
<td>102.1 (10.2)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>FVC predicted</td>
<td>87.0 (13.9)</td>
<td>100.9 (11.1)</td>
<td>0.005</td>
</tr>
<tr>
<td>FEV₁/FVC</td>
<td>41.6 (11.8)</td>
<td>79.2 (5.6)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Emphysema severity*</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>%LAA-950</td>
<td>26.3 (10.3–42.4)</td>
<td>2.5 (1.1–3.8)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>%LAA-910</td>
<td>59.9 (43.4–64.9)</td>
<td>26.3 (10.3–42.5)</td>
<td>0.002</td>
</tr>
<tr>
<td>Aortic pulse wave velocity, m/s</td>
<td>10.4 (2.0)</td>
<td>8.5 (1.7)</td>
<td>0.009</td>
</tr>
<tr>
<td>hsCRP, mg/l*</td>
<td>2.1 (1.3–5.5)</td>
<td>1.0 (0.5–3.5)</td>
<td>0.04</td>
</tr>
</tbody>
</table>

Data presented as mean (SD) or n (%), except * median (IQR). Groups compared using Student t test except for * where groups compared with Mann–Whitney U test.

Table 2 Skin measurements

<table>
<thead>
<tr>
<th></th>
<th>COPD</th>
<th>Control</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Area of dermis occupied by elastin fibres, %</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sun-exposed skin</td>
<td>43.5 (12.1)</td>
<td>26.3 (6.9)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Non-sun-exposed skin</td>
<td>22.4 (5.2)</td>
<td>18.1 (4.3)</td>
<td>0.02</td>
</tr>
<tr>
<td>mRNA expression, % increase</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>SES MMP-2</td>
<td>+14 (1 to 30)</td>
<td>0.06</td>
<td></td>
</tr>
<tr>
<td>SES MMP-9</td>
<td>+48 (16 to 79)</td>
<td>0.004</td>
<td></td>
</tr>
<tr>
<td>SES MMP-12</td>
<td>Insufficient expression for analysis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>SES TIMP-1</td>
<td>+15 (1 to 31)</td>
<td>0.06</td>
<td></td>
</tr>
<tr>
<td>SES MMP-2</td>
<td>–3 (21 to 12)</td>
<td>0.6</td>
<td></td>
</tr>
<tr>
<td>SES MMP-9</td>
<td>+17 (35 to 74)</td>
<td>0.5</td>
<td></td>
</tr>
<tr>
<td>SES MMP-12</td>
<td>Insufficient expression for analysis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>SES TIMP-1</td>
<td>–4 (22 to 13)</td>
<td>0.6</td>
<td></td>
</tr>
<tr>
<td>Protein quantification, mg/ml*</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>SES proMMP-2</td>
<td>22.3 (16.7–24.7)</td>
<td>20.8 (16.5–33.5)</td>
<td>0.85</td>
</tr>
<tr>
<td>SES MMP-2 activity</td>
<td>13.6 (10.3–24.8)</td>
<td>22.7 (13.0–29.4)</td>
<td>0.25</td>
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<tr>
<td>SES total MMP-2</td>
<td>33.3 (22.9–36.5)</td>
<td>29.9 (23.7–45.8)</td>
<td>0.93</td>
</tr>
<tr>
<td>SES proMMP-9</td>
<td>11.7 (7.3–12.1)</td>
<td>7.1 (4.9–9.4)</td>
<td>0.02</td>
</tr>
<tr>
<td>SES proMMP-2</td>
<td>27.7 (19.5–37.6)</td>
<td>35.6 (28.6–47.6)</td>
<td>0.08</td>
</tr>
<tr>
<td>SES MMP-2 activity</td>
<td>10.1 (6.4–15.2)</td>
<td>9.8 (8.4–13.3)</td>
<td>1.0</td>
</tr>
<tr>
<td>SES total MMP-2</td>
<td>17.7 (12.9–26.0)</td>
<td>23.5 (18.6–36.6)</td>
<td>0.06</td>
</tr>
<tr>
<td>SES proMMP-9</td>
<td>9.1 (6.1–11.5)</td>
<td>10.9 (8.9–12.2)</td>
<td>0.12</td>
</tr>
</tbody>
</table>

mRNA expression calculated as the percentage difference in the COPD group relative to the control data. Data presented as mean (SD) or analysed using Student t test, except * median (IQR).

DISCUSSION

Patients with COPD have evidence of greater cutaneous elastin degradation than age- and smoking-matched controls. Our findings represent evidence of enhanced degradation of elastin outside of the lung in COPD, and we believe that systemic elastin degradation may be a novel systemic feature of COPD. Furthermore, we have found increased expression of MMP-9 in the skin of patients with COPD, a protease implicated in the pathogenesis of emphysema, and these data suggest a role in systemic elastin degradation in COPD.

Elastin degradation in non-sun-exposed skin was associated with both emphysema severity and aortic pulse wave velocity. We speculate that the systemic elastin degradation demonstrated in the skin may also occur in other elastin-rich structures, such as the walls of large conduit arteries, and is an explanation for the increased arterial stiffness in COPD.

Development of emphysema, arterial stiffness and elastin degradation are features of normal ageing, reinforcing the importance of matching our groups for age. Our findings support the hypothesis that COPD is a disease characterised by accelerated ageing. Recent manuscripts have suggested that the arterial stiffness associated with COPD may in part be due to increased vascular calcification and that vascular calcification is associated with emphysema severity. Indeed, this mirrors the medial elastocalcinosis seen in large vessels as a feature of normal ageing. As the elastin in the vessel wall is degraded, the calcium content is thought to increase. Thus, measures of vascular calcification in the aorta may also be a reflection of elastin degradation. In addition, as malnutrition is associated with elastin degradation in the lungs and vasculature, it is noteworthy that subjects with COPD had a similar mean body mass index to the controls.

Both the ATS/ERS consensus definition and the GOLD guidelines classify COPD as a disease with systemic consequences and extra-pulmonary effects. However, it is difficult determine whether this relationship is causal—that is, due to a direct consequence of COPD—or results from a combination of shared risk factors for COPD and cardiovascular disease such as smoking, social deprivation and a sedentary lifestyle. We have performed carefully controlled studies comparing patients with COPD and controls matched for age and smoking history that suggest arterial stiffness is an independent systemic manifestation of COPD. Furthermore, emphysema severity is closely related to arterial stiffness independent of cigarette smoking in patients with COPD. Together, these findings suggest a shared susceptibility to elastin degradation in the pulmonary and systemic tissues of COPD patients.

Vascular samples from conduit arteries are not readily available; however, previous research on the dermatological manifestations of COPD suggested that skin may be a suitable surrogate for assessing systemic elastin degradation. Using visual categorisation of facial wrinkling, skin wrinkling has been associated with emphysema severity and lung function in a cohort of patients with COPD. Our study provides the first direct evidence of increased elastin degradation in the skin in COPD patients, and importantly we are able to demonstrate that this observation is independent of cigarette smoke exposure. Two previous studies have reported increased cutaneous elastin degradation using biopsy specimens in cigarette...
Chronic obstructive pulmonary disease

Figure 2 Patients with chronic obstructive pulmonary disease (COPD) have increased elastin degradation in both sun-exposed and non-sun-exposed skin. Symbols represent individual values and the horizontal lines the means.

Figure 3 Sun-exposed skin elastin degradation was associated with both matrix metalloproteinase (MMP)-2 (panel A; r=0.55, p=0.002) and MMP-9 (panel B; r=0.62, p<0.001) mRNA expression. The y-axis represents relative expression based on an internal standard. Subjects with chronic obstructive pulmonary disease are represented by the closed circles (●) and the controls are represented by the open circles (○). MMP-2 expression was detectable in all subjects; however, MMP-9 expression was below the lower limit of detection in two subjects.
Figure 4  Non-sun-exposed skin elastin degradation (percentage of dermis occupied by elastin fibres) was associated with airflow obstruction (panel A: forced expiratory volume in one second (FEV1) %predicted; r=-0.45, p=0.01), arterial stiffness (panel B: aortic pulse wave velocity; r=-0.42, p=0.02) and emphysema severity (panel C: percentage low attenuation voxels below -910 Hounsfield units (%LAA-910); r=0.50, p=0.007). Arterial stiffness was associated with emphysema severity (panel D: %LAA-910; r=0.47, p=0.01). Pearson’s correlations were used across all subjects. Subjects with chronic obstructive pulmonary disease are represented by the closed circles (●) and the controls are represented by the open circles (○).

There was generally reduced expression of MMPs in non-sun-exposed skin in comparison with sun-exposed skin and production of cutaneous MMPs in response to ultraviolet radiation is well described.44 Thus, it may be that sunlight exposure is a complementary factor which increases elastin degradation in susceptible skin tissue, unmasking differences in protease production between the patients with COPD and the healthy subjects. However, the pathophysiology of both COPD and vascular disease is complex and the exact inflammatory mechanisms are as yet unclear. While we have specifically investigated cutaneous MMPs, it is important to acknowledge that other proteases may have a role in the pathogenesis of both vascular disease and the development of emphysema. Serine proteases (eg, neutrophil elastase) have been implicated in both athero-disease and the development of emphysema. Serine proteases (eg, cathepsin K) in animal models.47 48 Thus, these other proteases may be involved in systemic elastin degradation in COPD.

We did consider measuring elastin degradation products, but our protein extraction methods were suitable to extract MMPs and not the relatively insoluble desmosine/isodesmosine from samples. In any case, a consistent and reliable methodology for assessment of these proteins is yet to be established. Indeed, desmosine levels in the lung parenchyma are similar in patients with severe COPD and healthy donor lungs.49 Future studies could specifically assess desmosine levels, its relationship with elastin degradation and its prospective use as a local tissue biomarker.

Although we did exclude individuals on regular oral steroids, we were unable to adjust for inhaled corticosteroids use, taken by the majority of patients with COPD. Skin thinning, bruising and atrophy are recognised side effects of corticosteroid therapy. A small biopsy study suggested that inhaled corticosteroids may affect collagen synthesis,50 but in the largest study assessing the effects of long term inhaled corticosteroids on skin there was no effect on skin collagen or thickness.51 No specific effect of inhaled corticosteroids on elastin fibres has been described.

CONCLUSIONS

Patients with COPD have increased skin elastin degradation in comparison with age- and smoking-matched controls and cutaneous elastin degradation was related to both emphysema severity and arterial stiffness. Furthermore, we identified upregulation of MMP-9 in the skin of COPD patients, which is associated with elastin degradation. Systemic elastin degradation due to increased proteolytic activity may represent a novel shared mechanism for the pulmonary, vascular and cutaneous features of COPD.

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Contributors Conception and design: JM, DM, NM, RG, WM; Laboratory and radiological methodology and analysis: RR, IH, SM, SH, MC, JM, EvB; Drafting the manuscript for important intellectual content: JM, DM, NM, RG, WM.

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Competing interests None.

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