ORIGINAI ARTICLE

Pseudomonas aeruginosa LasB protease impairs innate immunity in mice and humans by targeting a lung epithelial cystic fibrosis transmembrane regulator–IL-6–antimicrobial–repair pathway

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ABSTRACT

Background Pseudomonas aeruginosa lung infections are a huge problem in ventilator-associated pneumonia, cystic fibrosis (CF) and in chronic obstructive pulmonary disease (COPD) exacerbations. This bacterium secretes virulence factors that may subvert host innate immunity.

Objective We evaluated the effect of P. aeruginosa elastase LasB, an important virulence factor secreted by the type II secretion system, on ion transport, innate immune responses and epithelial repair, both in vitro and in vivo.

Methods Wild-type (WT) or cystic fibrosis transmembrane conductance regulator (CFTR)-mutated epithelial cells (cell lines and primary cells from patients) were treated with WT or ΔLasB pseudomonas aeruginosa O1 (PAO1) secretomes. The effect of LasB and PAO1 infection was also assessed in vivo in murine models.

Results We showed that LasB was the most abundant protein in WT PAO1 secretomes and that it decreased epithelial CFTR expression and activity. In airway epithelial cell lines and primary bronchial epithelial cells, LasB degraded the immune mediators interleukin (IL)-6 and trappin-2, an important epithelial-derived antimicrobial molecule. We further showed that an IL-6/STAT3 signalling pathway was downregulated by LasB, resulting in inhibition of epithelial cell repair. In mice, intranasally instillated LasB resulted in significant weight loss, inflammation, injury and death. By contrast, we showed that overexpression of IL-6 and trappin-2 protected mice against WT-PAO1-induced death, by upregulating IL-17/IL-22 antimicrobial and repair pathways.

Conclusions Our data demonstrate that PAO1 LasB is a major P. aeruginosa secreted factor that modulates ion transport, immune response and tissue repair. Targeting this virulence factor or upregulating protective factors such as IL-6 or antimicrobial molecules such as trappin-2 could be beneficial in P. aeruginosa–infected individuals.

INTRODUCTION

Cystic fibrosis (CF) is the most common genetically inherited disease in Caucasian populations (1 in 3500 newborns) and 70%–90% of CF individuals harbour the F508del mutation, resulting in misfolding and incorrect trafficking of the cystic fibrosis transmembrane conductance regulator (CFTR) molecule to the epithelial membrane.1–3 CFTR is an anion channel that regulates fluid homeostasis on epithelial surfaces.4 In the airways, a loss of function and/or stability3 of this protein is thought to sequentially induce hypohydration, mucus accumulation, bacterial infections (eg Pseudomonas aeruginosa, Burkholderia cepacia) and chronic inflammation via the recruitment of neutrophils.3 Although CFTR mutations are responsible for disease in individuals with CF and lead to chronic P. aeruginosa infections (a key hallmark of the disease), CFTR expression has recently also been reported as downregulated in epithelial cells treated with cigarette smoke.6–7 In vivo also, genetically CFTR-sufficient cigarette smokers showed a decrease in CFTR function,8 and similarly, in biopsies from non-CF chronic obstructive pulmonary diseases (COPD)/emphysemae
patients, CFTR protein expression was significantly decreased with disease activity and was associated with inflammation. We have shown recently that neutrophil elastase (NE) is able to degrade CFTR in vitro and in vivo, through the activation of intracellular calpains, potentially explaining infectious and inflammatory exacerbations in CF and COPD. Importantly, when comparing the effect of *Pseudomonas* infection on CFTR in wild-type (WT) and NE−/− mice, we found NE to account for only part of CFTR degradation. We therefore hypothesised that other factors, of *P. aeruginosa* origin, may also target and have deleterious effects on CFTR and on innate immune responses downstream of CFTR. Here, we identify LasB, a *P. aeruginosa* type II secretion system metalloprotease and an important virulence factor present in CF secretions as the main secreted protein in the secreteme from the *pseudomonas aeruginosa* O1 (PAO1) strain. Using biochemical and functional assays, we investigated its effects on CFTR function and on the regulation of innate immune responses, in vitro and in vivo.

We demonstrate that LasB degrades CFTR and downregulates an interleukin (IL)-6–antimicrobial–lung repair pathway in vitro and ex vivo in primary airway cells from patients. Furthermore, we show that this pathway can be rescued in vivo in mice by overexpressing IL-6 and the antimicrobial molecule trappin-2.

Because *P. aeruginosa* infections are a common feature in CF and COPD/emphysema exacerbations, and since LasB is invariably found in inflammatory secretions (particularly in CF), our data suggest that targeting *P. aeruginosa* LasB and/or stimulating the IL-6/antimicrobial/repair axis maybe an interesting and novel approach for tackling the inflammatory processes underlying *P. aeruginosa*-induced lung disease exacerbations.

### MATERIALS AND METHODS

**Materials**

Phosphoramidon (PA, R7385), amiloride, forskolin, IBMX and CFTRinh172 were obtained from Sigma-Aldrich. Purified LasB (pLasB) was a kind gift from Pr. Gerd Döring and recombinant human IL-6 and 1β were purchased from R&D Systems. Taq II substrate was obtained from Enzolife Science.

**Preparation and analysis of WT PAO1 and ΔLasB PAO1 secretomes (WT-SEC and ΔLasB-SEC)**

PAO1 WT, a strain expressing LasB, and PAO1 ΔLasB strain (gift from Pr. D. Ohman) were grown overnight in Luria Broth (LB) medium (1% Bactotryptone, 0.5% Bacto Yeast Extract, 0.5% NaCl) under agitation. Bacterial suspensions were then centrifuged at 4000 g (15 min, 4°C), 6000 g (10 min, 4°C) and 12000 g (10 min, 4°C) before the remaining supernatants (secretomes (SEC)) were filter-sterilised using 0.2 μm pore syringe filters, aliquoted and stored at −80°C until use.

Purified SECs were then analysed by sodium dodecyl sulfate (SDS-PAGE) or zymography. For the latter, SECs were prepared in β-mercaptoethanol-free 4X Laemmli buffer and separated on 7% acrylamide gels containing 2% gelatin. After migration, gels were washed and incubated overnight in the developing buffer, stained in 0.05% Coomassie brilliant blue for 1 hour and destained with appropriate Coomassie R-250 destaining solution (methanol:acetic acid:water (50:10:40)).

**Cytokine and trappin-2 secretion measurements**

Media were collected and assayed for IL-8 and IL-6 by ELISA kits (R&D Systems, Minneapolis, Minnesota, USA) following the manufacturer’s instructions. Human trappin-2 was measured using an ELISA available in-house (Ref S4).

Cells, cell culture and analysis, scratching protocols, short-circuit current measurements, adenovirus constructs

These are described in detail in the online supplementary file 1.

### In vivo experiments

Procedures involving mice were approved by our local ethical committee (Paris-Nord/No 121) and by the French Ministry of Education and Research (agreement number 04537.03). Eight-week-old male C57BL/6 mice and trappin-2 transgenic mice (hereafter called eTg mice) were from Janvier (Le Genest-Saint-Isl, France) and generated by our group, respectively. Mice were anaesthetised using intramuscular injection of ketamine 500 and xylazine 2% in 0.9% NaCl (20:10:70). Different quantities of pLasB, Ad-vectors or PAO1 bacteria were instilled either intranasally or intratracheally (final volume of 40 μL max), followed by either monitoring their survival, or by humanely killing the animals (overdose of 100 μL intraperitoneally injected pentobarbitral) for mechanistic studies (see online supplementary file 1). Bronchoalveolar lavages (BALs) fluid were obtained by cannulating the trachea and instilling 2×1 mL of phosphate-buffered saline (PBS). Typically, a volume of 1.7 mL of BALF was retrieved and centrifuged at 2000 rpm for 10 min. Supernatants were used for protein, cytokine (ELISA) and haemoglobin (used as a surrogate for lung damage, absorbance reading at 405 nm) measurements. Cell pellets were used for cell differential analysis (Diff-Quick, Dade Diagnostika GmbH, Unterschleissheim, Germany).

In parallel, RNA isolation, followed by reverse transcription and quantitative PCR (RT-q-PCR), was performed as described in online supplementary file 1. Finally, lung tissue was also used for quantifying bacteria after plating extracts on agarose plates.

### Statistical analysis

Data were analysed with GraphPad Software with either one-way analysis of variance (followed by Bonferroni post hoc tests) or non-parametric analysis, followed by post-Dunn’s test for multiple comparisons.

Survival curves in murine models experiments were plotted using Kaplan-Meier curves and statistical tests were performed using the Log-rank (Mantel-Cox) test.

### RESULTS

LasB is the main protein in the *P. aeruginosa* secreteme, degrades epithelial cells CFTR and decreases CFTR activity in vitro

We first demonstrated, using Coomassie blue staining of SDS-PAGE gels and zymography techniques, that LasB was the major protein secreted in WT PAO1 secreteme and that it was active as a metalloprotease (see relevant information on in online supplementary file 1).

We then investigated the effect of PAO1 WT and ΔLasB-SEC on CFTR expression in different cell lines. In NCI-H292 cells (cells expressing minimal amounts of endogenous CFTR) transfected with Ad-GFP-WT-CFTR, PAO1 WT SEC (but not ΔLasB-SEC) induced CFTR degradation (figure 1A). Similar results were observed in polarised Calu-3 cells, which express high amounts of endogenous CFTR (figure 1B). This was not due to an increase in CFTR endocytosis as these results were reproduced in the presence of dynasore (figure 1C).

We then evaluated if this decrease in expression induced a decrease in activity in WT-CFTR cystic fibrosis bronchial epithelial (CFBE) cells (stably expressing CFTR). WT-SEC (but not ΔLasB-SEC) significantly decreased CFTR activity as shown...
LasB downregulates CFTR protein levels in epithelial cells. (A) Western blot analysis of CFTR expression in NCI-H292 cells overexpressing GFP-WT-CFTR, as detected by anti-GFP antibody after a 24-hour treatment with either 5% LB medium, 5% WT-SEC or 5% ΔLasB-SEC. (B) Western blot analysis of CFTR expression in polarised Calu-3 cells after a 24-hour treatment with either 5% LB medium, 5% WT-SEC or 5% ΔLasB-SEC, as detected by anti-CFTR(596) antibody. (C) Western blot analysis was performed as in (B), except that cells were preincubated with dynasore (80 µM), a GTPase inhibitor that targets dynamin and blocks endocytosis, at 37°C for 30 min. Each image is representative of n=3 independent experiments. CFTR, cystic fibrosis transmembrane conductance regulator; GAPDH, glyceraldehyde-3 phosphate dehydrogenase; LB, Luria Broth; SEC, secretomes.
LasB downregulates IL-6 and the antimicrobial molecule trappin-2 in human lung epithelial cells

It has previously been suggested that the absence of CFTR ‘per se’ may provide an ‘inflammatory phenotype’, with modulated secretion of IL-6 and IL-8 for example as well as that of antimicrobial molecules. In addition, downregulation of CFTR affects acid/base homeostasis of epithelial surfaces and can therefore also affect antimicrobial function. Our results shown above therefore prompted us to test whether LasB might modulate these important epithelial cell innate immune mediators.

Because NCI-H292 produced low amounts of trappin-2 at steady state, its production was upregulated with IL-1β (figure 3A–C), whereas no upregulation was needed in Calu-3 (figure 3D–F) and CFBE cells (figure 3G–I). In NCI-H292, LB medium + IL-1β and ΔLasB-SEC + IL-1β treatments induced an increase in IL-6 (figure 3B) and IL-8 (figure 3C) compared with IL-1β alone, indicating that LB and secretome components were stimulatory. However, WT-SEC, despite containing the same components as ΔLasB-SEC, completely abolished IL-1β-induced trappin-2 (figure 3A) and IL-6 (figure 3B) secretions but had no effect on IL-8 production (figure 3C). In addition, we showed in these cells that purified pLasB had a similar effect than the WT-SEC and that PA, a metalloprotease inhibitor, inhibited pLasB-induced and WT-SEC-induced downregulation of trappin-2 and IL-6 (figure 3A–B). In Calu-3 cells, ΔLasB-SEC similarly increased trappin-2, IL-6 and IL-8 production (figure 3D–F), compared with LB medium-treated cells, demonstrating as above that the bacterial pathogen associated molecular patterns (PAMPs) contained in ΔLasB-SEC can up-regulate these mediators. By contrast, WT-SEC, again, completely inhibited trappin-2 and IL-6 protein recovery in the supernatants (figure 3D–E), whereas IL-8 protein levels were unaffected (figure 3F).
LasB reduces trappin-2 and IL-6 recovery, but not that of IL-8, in the supernatants of NCI-H292, Calu-3 and CFBE cells (WT-CFTR and ΔF508). (A–C): NCI-H292 cells were preincubated with IL-1β for 1 hour prior to the addition of diluted SEC (5%) or purified LasB (pLasB) for 4 hours, supplemented or not with the metalloprotease inhibitor phosphoramidon (PA, 8.5 µM). DMSO was also added as a control, since it was used as a diluent for PA. Trappin-2, IL-6 and IL-8 protein levels were measured by ELISA in cell supernatants (n=4, *p<0.05, **p<0.01, ***p<0.001, ns: not significant; ANOVA). (D–F): Calu-3 cells were incubated with LB medium or with diluted SECs (5%) for 4 hours. Trappin-2, IL-6 and IL-8 protein levels were measured as above (n=3, ANOVA). (G–I): CFBE-WT-CFTR and CFBE-F508del-CFTR cells were either untreated (CTL), treated with LB medium, or with diluted SEC (1%) for 24 hours and ELISAs performed as above (n=3–4, two-way ANOVA). Results are shown as mean±SEM. ANOVA, analysis of variance; CFBE, cystic fibrosis bronchial epithelial; CFTR, cystic fibrosis transmembrane conductance regulator; DMSO, dimethyl sulfoxide; IL, interleukin; LB, Luria Broth; SEC, secretomes.

Respiratory infection

Similar results were obtained from CFBE cells over-expressing WT-CFTR (figure 3G-I) where WT-SEC, but not ΔLasB-SEC, downregulated apical secretion of IL-6 (figure 3H) and trappin-2 (figure 3G), whereas IL-8 levels remained unchanged (figure 3I). Since we have shown above that LasB decreased CFTR expression and activity, it appeared interesting to evaluate the effect of this metalloprotease on the immune mediators in a CFTR-deficient context. Interestingly, when mutant CFTR (F508del-CFTR) overexpressing CFBE cells were compared with WT-CFTR CFBE cells, the latter had higher levels of IL-6 and trappin-2 protein (but not IL-8), compared with the former (figure 3I). Notably, as in WT-CFTR cells, WT-SEC (but not ΔLasB-SEC) drastically downregulated IL-6 (but not IL-8) secretion in F508del-CFTR CFBE; because the levels of trappin-2 in the latter cells were almost undetectable at baseline, the potential further effect of WT-SEC on this mediator was likely not detected in these cells.

Interestingly, WT-SECs had similar effects on IL-6 and trappin-2 secretion in cells from patients with CF (see online supplementary figure 2).

IL-6 up-regulates trappin-2, and LasB disrupts this functional pathway by affecting STAT-3 activation and lung epithelial repair

Having shown that LasB downregulates IL-6 and trappin-2, two anti-inflammatory mediators, we then investigated if there was any functional link between these two molecules. We showed that IL-6 upregulated trappin-2 secretion in NCI-H292 cells (figure 4A) and that WT-SEC (but not ΔLasB-SEC) was able to downregulate this increase (figure 4B).

Because IL-6 signalling induces STAT-3 activation²³–²⁵ and because some antimicrobial molecules are under the control of this transcription factor,²⁶,²⁷ the effect of LasB was also studied on this pathway. Echoing the results showing that LasB could
downregulate IL-6-induced secretion of trappin-2, LasB inhibited STAT-3 activation in NCI-H292 cells (figure 4C).

The IL-6/STAT3 pathway has been shown to be involved in epithelial repair in the lungs and the intestine. We showed, in an epithelial scratch assay on NCI-H292 cells, that IL-6 increased epithelial repair (figure 5A) and that WT-SEC (but not ΔLasB-SEC) very significantly hampered this effect (figure 5B). This was also demonstrated in polarised overexpressing WT-CFTR CFBE cells where, in the absence of IL-6, WT-SEC significantly impaired spontaneous repair following cell scratching (figure 5C–D).

LasB is an important P. aeruginosa virulence factor in mice lungs and in vivo transgenic expression of IL-6 and trappin-2 enhances innate immune protection against acute P. aeruginosa infection.

It has been shown in several organ systems that LasB can be very damaging as reviewed in ref31. Therefore, we evaluated the effect of LasB in our C57BL/6 mouse model. At high doses (≥40 µg), pLasB killed 100% of mice within 5 hours (figure 6A). With a lower dosage of pLasB (5 and 10 µg), significant weight losses were observed 24 hours postintra-

nosal instillation (figure 6B). At that time point, significant lung inflammation and lung injury were apparent in these mice as shown by an increase in cellularity (neutrophils, figure 6C–E), blood (figure 6F) and molecules involved in the host response to infection (figure 6G–J). We then performed ‘rescue experiments’ and showed that Ad-mediated IL-6 (5.10^8 pfu) overexpression was protective in an acute model of PAO1 (2.10^7 cfu) infection. Notably, all control PBS and Ad-null-treated C57BL/6 mice died within 50 hours, whereas more than 60% of IL-6 overexpressers survived (statistical significance p=0.01, figure 7A). We reasoned that overexpressing trappin-2 with this low dosage of PAO1WT may not be a significant advantage and we therefore performed a further independent experiment with a higher PAO1WT load to test the effect of trappin-2. In that setting, using the same dosage of Ad vectors (5.10^8 pfu) and a higher dose of PAO1 (4.10^9 cfu), we showed that after infection with PAO1, trappin-2 transgenic (eTg) survived significantly longer than C57BL/6 controls (p=0.007, figure 7A).
LasB decreases basal and IL-6-induced epithelial repair. (A) Epithelial repair, shown as mean±SEM and as measured by scratch assay as the percentage initial (Time 0, T0) wound 16 hours postscratch in NCI-H292 treated with increasing concentrations of IL-6 (n=4, ANOVA). (B) Effect of WT-SEC and ΔLasB-SEC on basal and IL-6-induced epithelial repair. Injured NCI-H292 cells were pretreated with IL-6 (1 or 10 ng/mL) for 1 hour, and secretomes were added for the following 16 hours (n=5, two-way ANOVA). Results are shown as mean±SEM. Panel C shows representative images, at time 0 and time 16 hours, of the wounded polarised WT-CFTR-CFBE cells that were apically treated with LB, WT-SEC or ΔLasB-SEC. (D) shows the mean±SEM of the percentage of wound closure of three independent experiments of WT-CFTR-CFBE cells apically treated with LB, WT-SEC or ΔLasB-SEC (n=3, ANOVA). ANOVA, analysis of variance; CFTR, cystic fibrosis transmembrane conductance regulator; IL, interleukin; LB, Luria Broth; SEC, secretomes.

Figure 5

PBS, compared with infected C57BL/6 controls. Remarkably, trappin-2 and IL-6 double overexpressers had a very significant increased survival over controls and eTg simple overexpressers (all statistically significant, figure 7B).

In vivo transgenic expression of IL-6 and trappin-2 enhances innate immune protection against acute P. aeruginosa infection by increasing bacterial clearance, decreasing lung injury and engaging tissue repair pathways

Dissecting the mechanisms underlying the phenotype described above, we showed that C57BL/6 mice overexpressing IL-6 (group 2, figure 8A) and eTg mice (transfected or not with Ad-IL-6, group 3 and 4, respectively) had reduced PAO1 loads, when compared with Ad-null WT C57BL/6 infected mice (group 1). Notably, Ad-IL-6 treatment did not further decrease PAO1 load on the eTg background (groups 3 vs 4). This increase in P. aeruginosa clearance was also associated with a clear protection against lung damage, as assessed by measuring haemoglobin content in BALs (OD405nm, figure 8B and C), with the mice most protected being the double IL-6-trappin-2 overexpressers (group 4). When BAL cellularity was assessed, Ad-IL-6 treatment showed an ‘anti-inflammatory’ phenotype in the context of PAO1 infection, with a trend towards reduction and increase in neutrophils and lymphocytes, respectively (online supplementary figure 3). In addition, eTg mice treated with Ad-IL-6 showed an increase influx of lymphocytes, compared with WT mice. Furthermore,
a transcriptomic study indicated that Ad-IL-6 treatment had an overall downregulatory transcriptional effect in the context of PAO1 infection, in C57BL/6 WT, but above all in eTg mice (online supplementary figure 4).

To further dissect this, we analysed individually by real-time PCR (RT-PCR) a number of genes not represented in the PCR array described above. As expected, we demonstrated, in a non-infectious context, the induction of the IL-6 gene (lower dCT reflecting higher gene induction) following Ad-IL-6 treatment in both C57BL/6 WT and eTg mice (figure 9A). IL-6 was also induced by PAO1 infection and its expression was higher in eTg mice expressing Ad-IL-6 and infected with PAO1 when compared with similarly treated C57BL/6 mice. Trappin-2 expression was, as expected, only detected in eTg mice (since C57BL/6 mice do not express trappin-216 (as reflected here by a dCT>30 in C57BL/6 WT), and PAO1 upregulated its expression in eTg mice (figure 9B). In addition, there was a trend, which did not reach statistical significance, for an increase in trappin-2 expression following Ad-IL-6- (p=0.080) or Ad-IL-6 + PAO1 (p=0.09), respective to Ad-null or Ad-null + PAO1 eTg controls (figure 9B).

Neither Ad-IL-6 nor trappin-2 overexpression induced KC (figure 9C) or CCL-2 (figure 9D) expression, but as above, PAO1 infection upregulated these cytokines.

PAO1 also induced IL-17 and IL-22 in both C57BL/6 WT and eTg mice. IL-6 + PAO1 treatment was the most potent IL-17 and IL-22 inducer within the eTg group, but also when compared with C57BL/6 WT mice (figure 9E–F).

The existence of an IL-6-IL-17-IL-22-trappin-2 pathway was further demonstrated by showing strong correlations in the expression of these mediators, when all individual mice were analysed together (online supplementary figure 5, panels A–E).

Because the IL-17/IL-22 pathway has been linked to the expression of antimicrobial molecules,32 we tested the expression of other antimicrobials (than trappin-2) in our system. In

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**Figure 6** LasB triggers pulmonary innate immune responses in vivo and is important for pathogenesis. C57BL/6 male mice were intranasally instilled with increasing doses of LasB and survival was monitored (A, numbers in parenthesis represent the number of mice used). Alternatively, PBS (n=3) or moderate doses of LasB (5 µg, n=5 or 10 µg, n=4) or 40 µg were instilled. Mice weight was measured before and 24 hours postinillation (B). At that time point, mice lungs were recovered and bronchoalveolar lavage (BAL) fluid was analysed for cellularity (C–E) and haemoglobin content (absorbance at 405 nm (F), NB: the original absorbance was multiplied by 10, the BALF dilution factor. In parallel, RNA from lung extracts (homogenised in Trizol) were analysed by qPCR analysis for the expression of TNFα (G), MIP1α (H) and antimicrobial peptides: S100a8 (I) and Lcn2 (J). Results are shown as medians±IQR (n=3–5, ANOVA). ANOVA, analysis of variance.
addition to trappin-2, Ad-IL-6 expression had a clear upregulating effect on Reg-3γ in both WT and eTg mice. In eTg mice receiving Ad-IL-6 and infected with PAO1, there was a (not statistically significant) trend towards increased Reg-3γ expression compared with WT mice treated similarly (figure 9G, p=0.09). Furthermore, Ad-IL-6 and PAO1 strongly increased another antimicrobial, Lcn2, compared with the respective controls, with C57BL/6 and eTg behaving similarly in that respect (figure 9H and online supplementary figure 5 panels F and G).

Finally, we showed that overexpression of IL-6 and trappin-2 significantly induced a variety of molecules involved in lung repair. Notably, in C57/Bl6 mice, overexpression of IL-6 increased the expression of Mrc1, Hgf, fibronectin 1 (Fn1) and α1 type I collagen (Col1a1) (figure 10). Furthermore, in the context of PAO1 infection, overexpression of IL-6 and trappin-2 significantly increased mRNA levels of Fn1 and Col1a1. Significant correlations were established between these factors involved in epithelial repair and IL-6 and trappin-2 (online supplementary figure 6).

Altogether, these experiments (depicted in figures 7–10 and online supplementary figures S3–S6) demonstrate that IL-6 and trappin-2 overexpressors rescue the deleterious effect of WT-PAO1 (LasB expresser) in murine lungs. An outstanding interesting issue will be to assess whether this protective effect also occur in a situation where the less virulent ΔLasB-PAO1 strain infects the lung.

**DISCUSSION**

*P. aeruginosa* infections are a common feature of cystic fibrosis and COPD/emphysema exacerbations, and we and others have recently shown that the epithelial channel CFTR can be downregulated in vitro and in vivo by exogenous noxious stimuli such as cigarette smoke or endogenous products of innate immune activation, such as neutrophil elastase. However, the role of *P. aeruginosa* and that of its virulence factors on CFTR biology has not been extensively studied.

Among these virulence factors, LasB-bearing *P. aeruginosa* has previously been shown to be consistently present in mucoid and/or non-mucoid CF sputum secretions. However, its activity has been studied mainly in vitro where LasB was shown to degrade some cytokines, eukaryotic cell surface receptors and to participate in bacterial escape from the host immune system, in host colonisation and tissue destruction. We show here that LasB is the main protein in the PAO1 secretome (online supplementary figure 1) and that it is the main factor responsible for CFTR degradation in different lung epithelial cell lines, resulting in loss of function. Because CFTR integrity and function is essential to maintain an adequate hydration of mucosae and prevent mucus stasis and bacterial infections (through maintaining...
adequate acid/base buffering activity and antimicrobial function), the effect of \textit{P. aeruginosa} LasB on CFTR can be interpreted as a mechanism by which \textit{P. aeruginosa} breaches the epithelial cell innate immune shield and gains a foothold to colonise the lung of susceptible individuals. Indeed, CFTR degradation by LasB will likely have consequences in CFTR-sufficient patients with COPD but also in CF individuals, including the ones harbouring the F508del-CFTR mutation, since the latter still bears residual CFTR activity (estimated at 5\% according to Dekkers et al\cite{38}). Similarly, the downregulation of antimicrobial molecules such as trappin-2 shown here is likely to be beneficial for the bacterium. We show here, in epithelial cell lines and primary cells from patients with CF, that this is associated with a drastic downregulation of IL-6 (at both transcriptional and translational levels, not shown) but that IL-8 levels are not affected.

Although other \textit{P. aeruginosa} PAMPs such as LPS and, to a lesser extent, flagellin, have been used extensively to model \textit{P. aeruginosa} infection in vivo, surprisingly, the direct effect of LasB has been, so far, clearly understudied, and little is known about its effects in the lung.\cite{41} Here, we showed LasB to be inflammatory at moderate doses (5–10\(\mu\)g), by inducing lung damage (figure \text{8E}), neutrophilic recruitment (figure \text{8D}) and by transcriptionally upregulating proinflammatory cytokines and antimicrobial molecules (figure \text{8F-I}). At higher doses (40\(\mu\)g), LasB was shown to be lethal (figure \text{8A}). In that context, echoing the in vitro data demonstrating the protective effect of IL-6 and...
trappin-2 following LasB exposure, we show that transgenic expression of IL-6 and trappin-2 enhanced protection against WT PAO1, a *P. aeruginosa* strain expressing LasB.42 Remarkably, all control mice died, whereas a significant proportion survived with either IL-6 or trappin-2 transgenic expression. Surprisingly, although IL-6 has been shown previously to be associated in vivo with reduced local lung and systemic inflammatory responses, following LPS pulmonary and intra-peritoneal instillation, respectively,43 this is, to our knowledge, the first report demonstrating the beneficial effect of IL-6 in any pneumonia model. Indeed, IL-6 is not classically described as a molecule with 'direct' anti-antimicrobial properties, and the latter may not fully account on its own for the results obtained here. It seems more likely that IL-6 acts through the induction of a variety of mediators (IL-17, IL-22, antimicrobials and molecules implicated in tissue repair, confirming in vitro data), which overall dissuade neutrophilia (no change in KC was observed here) and promote lymphocytic influx and tissue repair, as observed in other systems.23–25 44–46 Furthermore, although trappin-2 itself and defensins49 have been shown to be associated with repair processes (although mostly in other organs, and with little mechanistic information), we show here clearly, as stated above, that the IL-6 + trappin-2 dual regimen was the most effective in protecting mice against *P. aeruginosa*-induced pneumonia (figures 8 and 9). This suggests the existence of a synergy between these two mediators, both engaging anti-inflammatory/prorepair pathways, and ultimately leading to faster lung healing and removal of the microbial burden, improving tissue resilience and remodelling (figure 10; online supplementary figure 6).

Finally, and demonstrating the potential clinical relevance of the described IL-6-antimicrobial pathway, an additional interesting finding from our study is that, aside from the LasB effect, the levels of IL-6 and trappin-2 (but again not those of IL-8) were drastically reduced in mutant F508del-CFTR CFBE cells, compared with WT-CFTR-corrected CFBE cells. This advocates that the inhibition of the IL-6-trappin-2 pathway described here may be operative in cystic fibrosis. However, this will need to be confirmed in vivo since overexpression of CFTR in air liquid interface (ALI)-differentiated epithelial cells from four different F508del-CFTR individuals had no effect on the protein recovery of trappin-2, IL-6 and IL-8, when compared with Ad-null-infected control cells. Nonetheless, this could be explained by the short restoration of CFTR expression (48 hours treatment), which may not be optimal to modulate the F508del phenotype and restore the potentially deficient IL-6-trappin-2 pathway.

Regardless, a tantalising interpretation of our data could be that LasB, one of the major (and consistently found in CF secretions10–13) virulence factors of *P. aeruginosa*: (1) targets CFTR and, in doing so, modifies the lung mucosal milieu leading to ionic flux disturbances, increased acidity and antimicrobial...
loss of function; (2) targets trappin-2, an important bacterial/anti-inflammatory molecule; (3) hampers an IL-6/repair pathway while leaving untouched a ‘chronic neutrophilic program’, contributing to overzealous inflammation and lung damage.

Our novel findings hold important implications for both genetically CFTR-sufficient individuals infected with P. aeruginosa (eg, during nosocomial infections, or during COPD exacerbations), as well as for individuals with CF. Directly targeting the virulence factor LasB or upregulating the downstream host immune molecules IL-6 and trappin-2 represents new therapeutic strategies for such patients.

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REFERENCES


Pseudomonas aeruginosa LasB protease impairs innate immunity in mice and humans by targeting a lung epithelial cystic fibrosis transmembrane regulator–IL-6–antimicrobial–repair pathway

Vinciane Saint-Criq, Bérengère Villeret, Fabien Bastaert, Saadé Kheir, Aurélie Hatton, Aurélie Cazes, Zhou Xing, Isabelle Sermet-Gaudelus, Ignacio Garcia-Verdugo, Aleksander Edelman and Jean-Michel Sallenave

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