

Introduction The development and use of nanoparticles for medical applications is rapidly increasing. One of the most commonly used nanoparticles is silver, due to its antimicrobial properties. At present the use of silver nanoparticles is unregulated and there is increasing concern regarding their potential adverse health effects, particularly when inhaled. The alveolar region is the primary site of nanoparticle deposition following inhalation, thus it is important to consider the effects they will exert on the alveolar epithelium, including potential toxicity. The alveolar epithelium is covered with a surfactant layer, acting as a barrier to inhaled particulate matter and pathogens, with which nanoparticles will interact before reaching the epithelium. Thus, we hypothesise that the surfactant layer will influence the toxicity and inflammatory response of alveolar epithelial cells to inhaled AgNPs.

Methods Immortalised human alveolar type 1 epithelial (TT1) cells were exposed to increasing concentrations of 20nm or 110nm silver nanoparticles (AgNP) in the presence and absence of porcine surfactant (Curosurf) for 24 hours. Cell viability was measured using the MTS assay and IL-6 and IL-8 release was measured by ELISA. In addition, the effect of the antioxidant glutathione on cytokine release was assessed.

Results Exposure to 20nm and 110nm AgNP did not significantly affect cell viability in the presence or absence of Curosurf (0–50 g/ml). However, IL-6 and IL-8 release was significantly increased ($P < 0.0005$) for all AgNP at concentrations above 10 g/ml. Furthermore, preincubation of AgNP with Curosurf significantly inhibited this response ($P < 0.001$). Pre-treatment of cells with glutathione also significantly inhibited IL-6 and IL-8 release following AgNP exposure. When glutathione and Curosurf were combined there was a further inhibition of cytokine release.

Conclusion This study demonstrates that AgNP, whilst not overtly toxic, induce an inflammatory response in human alveolar epithelial cells that is driven by oxidative stress. Furthermore, we have shown that the presence of surfactant significantly attenuates the inflammatory response suggesting a protective effect against inhaled nanoparticles. Thus, this study demonstrates the importance of studying AgNP bioreactivity in the presence of lung secretions to accurately represent the likely effects of inhalation.

P140 BPIFB1/LPLUNC1 IS A NOVEL MARKER FOR THE BRONCHIOLISED EPITHELIUM IN IPF

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Idiopathic pulmonary fibrosis (IPF) is an irreversible and progressive lung disease with limited life expectancy after diagnosis. Histopathological studies of IPF lungs reveal the typical “Usual Interstitial Pneumonia” (UIP) pattern, with epithelial hyperplasia, areas of scarring with fibroblast foci and characteristic morphological abnormalities, including bronchiolization of alveolar ducts and honeycomb cysts. Although it seems likely that bronchiolar abnormalities are caused by changes in epithelial cell differentiation, specific markers of this process remain elusive. By analysis of published array data sets from IPF patients, we identified BPIFB1/LPLUNC1 as a potential candidate marker for the disease. Indeed, in the largest published study the gene was the most differentially expressed transcript. This putative innate defence protein is normally expressed in the airway submucosal

glands and in a population of MUC5A/C positive goblet cells in the upper airways. Analysis of lung tissue from UIP revealed strong staining of BPIFB1/LPLUNC1 within the bronchiolized epithelium lining the honeycomb cysts as well as in the mucous substance filling these regions. MUC5B was localized to the same cells as BPIFB1/LPLUNC1 whereas the related protein, BPIFA1/SPLUNC1, was not co-expressed. This pattern of staining was not seen in other chronic lung diseases, suggesting a degree of specificity for IPF. To shed light on a temporal association of expression of these markers with fibrosis development we studied mice exposed to the pro-fibrotic agent bleomycin (Bleo). MUC5B and LPLUNC1 were co-expressed in a population of goblet cells in the airways of mice within 3–7 days of Bleo exposure, prior to the onset of fibrosis. Continued expression was seen during the development of fibrosis between 14–21 days post treatment. In contrast, in mice treated with PBS neither protein was seen (due to mouse airways being essentially free of goblet cells). Staining was absent from the fibrotic regions and the lung parenchyma, as is the case in IPF. Our data show that the ectopic expression seen in human IPF, is mirrored by that seen in the fibrotic mouse model. Furthermore it suggests that BPIFB1/LPLUNC1 may be worthy of further study as potential marker for the disease.

P141 DIFFERENTIAL EXPRESSION OF CONVENTIONAL AND INHIBITORY VEGFA ISOFORMS IN NORMAL AND FIBROTIC FIBROBLASTS—A POTENTIAL ROLE IN IPF PATHOGENESIS?

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Introduction Vascular endothelial growth factor (VEGFA) has been implicated in the pathogenesis of Idiopathic Pulmonary Fibrosis (IPF). Two families of endogenous isoforms exist formed by alternative splicing of mRNA transcripts: the conventional potent angiogenic and mitogenic isoforms (VEGF_{xxx}a family) and the VEGF_{xxx}b family that is thought to have contrasting inhibitory functions.

Hypothesis We hypothesise that differential expression of VEGF_{xxx}a and VEGF_{xxx}b isoforms by fibroblasts may influence the development of IPF.

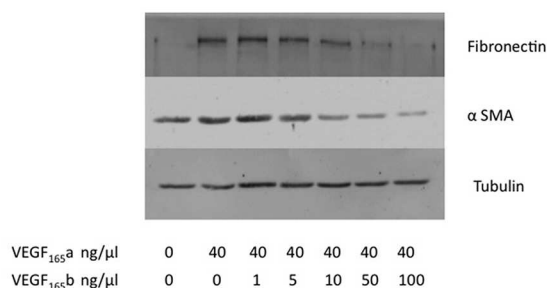
Methods Normal (NF) and fibrotic fibroblasts (FF) (from patients with proven UIP) were extracted from lung samples using the explant method. The expression of VEGF_{xxx}a and VEGF_{xxx}b by NF and FF was analysed at the mRNA level by RT-PCR and quantified by qPCR. Protein expression was determined by western blotting (WB) and ELISA. We sought to establish a potential functional effect of recombinant VEGF₁₆₅a and VEGF₁₆₅b proteins on fibroblasts by assessing the expression of a) the extracellular matrix (ECM) protein fibronectin and b) -SMA, a marker of myofibroblast differentiation.

Results Both NF and FF expressed VEGF_{xxx}a and VEGF_{xxx}b isoforms at the mRNA level as determined by RT-PCR with confirmation by direct sequencing. There was no statistical difference in total VEGF mRNA expression between the two cell types by qPCR ($p = 0.9307$, NF $n = 5$, FF $n = 6$), but FF expressed significantly more VEGF₁₆₅b mRNA than NF ($p = 0.05$, NF $n = 5$, FF $n = 6$). Total VEGF protein expression was significantly increased in FF (mean expression

NF = 180.5pg/ml vs FF 332.0pg/ml, $p = 0.0012$) by ELISA and confirmed by WB. Furthermore, increased VEGF_{165b} protein expression was also observed in FF by WB. Recombinant VEGF_{165b} had no effect on fibronectin or α -SMA expression in NF, but VEGF_{165a} (10ng/ l) significantly increased expression of fibronectin ($p < 0.05$). Interestingly, co-administration of VEGF_{165a} with VEGF_{165b} inhibited both α -SMA and fibronectin expression in these cells (Figure 1).

Conclusion Differential expression of VEGF isoforms between NF and FF suggests a potential role in the development of IPF. Furthermore, results suggest that factors altering the balance of splice variants may influence the surrounding fibrotic milieu.

Effect of rhVEGF_{165a} + rhVEGF_{165b} on fibronectin and α -SMA expression in normal fibroblasts



Abstract P141 Figure 1.

P142 SRC KINASE INHIBITION ATTENUATES NEUTROPHIL DEGRANULATION WITHOUT IMPAIRING BACTERIAL KILLING: A POSSIBLE THERAPEUTIC STRATEGY FOR ACUTE LUNG INJURY?

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Background A key mechanism in the pathogenesis of acute lung injury (ALI) is excessive neutrophil degranulation in response to an overwhelming inflammatory or infective insult. To date, no pharmacological therapy for ALI has proven beneficial.

Aim To investigate our hypothesis that extracellular neutrophil degranulation can be inhibited without necessarily impairing phagocytosis or live bacterial killing.

Methods Whole blood or purified neutrophils from healthy volunteers were pre-treated with a *src* kinase inhibitor (PP1) or vehicle control, before stimulation with either Phorbol 12-myristate 13-acetate (PMA), cytochalasin B + N-formylmethionyl-leucyl-phenylalanine (fMLP), lipopolysaccharide (LPS), live serum-opsonized *Staphylococcus aureus* (SA) or *Pseudomonas aeruginosa* (PA), to induce degranulation. Degranulation was measured in whole blood using CD63/CD66b expression and in purified neutrophils by extracellular release of myeloperoxidase (MPO) and lactoferrin (LTF). Neutrophil phagocytosis of fluorescent killed bacteria, cell viability, apoptosis and bacterial killing (by serial dilution and colony counting) were also measured. All experiments carried out using $n = 4-6$ healthy volunteers.

Results PP1 pre-treatment using concentrations above 10 μ M significantly attenuated primary and secondary granule exocytosis from healthy neutrophils in response to LPS, cytochalasin B/ fMLP, SA and PA but not to PMA. The same effect was observed in whole blood assays and in purified neutrophils, both in free suspension and when adhered to tissue culture plastic. PP1

treatment did not increase neutrophil death in response to the stimuli, nor did it significantly alter baseline apoptosis rates. Importantly, PP1 did not impair neutrophil phagocytosis or live bacterial killing of SA and PA.

Conclusions Our study supports our hypothesis and proposes *src* kinases as an attractive target for anti-inflammatory therapy in conditions such as ALI.

P143 HYPOXIA INDUCES HYPOTHERMIA AND SICKNESS BEHAVIOUR IN MICE FOLLOWING SUBCUTANEOUS INJECTION OF LIVE STAPHYLOCOCCUS AUREUS

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Infections frequently cause or complicate illnesses associated with hypoxaemia and local tissue hypoxia. The influence of hypoxia on the interaction between host and pathogen is therefore of considerable interest. *S. aureus* is a major pathogen in critical care where patients may have profound hypoxaemia and at a tissue level, *S. aureus* frequently infects ischaemic wounds. We investigated the effect of systemic hypoxia on host-pathogen interactions using a subcutaneous *S. aureus* infection model in mice.

C57BL/6 mice were shaved, injected with a low dose of *S. aureus* (5×10^7 SH1000) and placed in a hypoxic chamber (10% O₂) or left in room air. At 6 or 12 hours mice were assessed clinically and rectal temperature recorded. Clinical assessment of mouse sickness behaviour was made by two independent observers blinded as to which oxygen tension the mice had been exposed. Mice were anaesthetised and tissue samples (blood, skin, lung, spleen, kidney, liver and brain) obtained for analysis.

Mice injected with live bacteria and placed in hypoxia developed a phenotype of sickness behaviour and hypothermia. Infected hypoxic mice had significantly higher sickness scores and lower body temperature than infected normoxic mice or hypoxic mice injected with PBS (rectal temperature at 12 hours: hypoxic $33.4^\circ\text{C} \pm 0.74$, normoxic $37.7^\circ\text{C} \pm 0.24$, hypoxic PBS-injected $38.9^\circ\text{C} \pm 0.26$, $p < 0.0001$). Surprisingly, we found no evidence of bacteraemia, enhanced cytokine production, vascular leak or lung injury in the hypoxic infected mice. However, these animals had significant circulatory dysfunction, with hypotension, bradycardia and echocardiographic evidence of impaired left ventricular function. Interestingly, myeloid-cell deficiency of either HIF-1 or HIF-2 protected mice from the adverse systemic phenotype in this model, implicating the host innate immune response in the pathogenesis of the phenotype.

These findings imply that hypoxia may adversely alter the host response to a minor bacterial challenge, leading to profound systemic illness and that in such a setting modulation of the HIF pathway may be a possible therapeutic option.

P144 INFLUENZA INFECTION AFFECTS THE DEGREE OF FIBROSIS AND APOPTOSIS IN THE BLEOMYCIN MOUSE MODEL

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