Paediatric lung disease

Lung function prior to viral lower respiratory tract infections in prematurely born infants

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Anne Greenough1

ABSTRACT

Objective Prematurely born infants who develop respiratory syncytial virus (RSV) lower respiratory tract infections (LRTIs) have lung function abnormalities at follow-up. The aim of this study was to determine whether prematurely born infants who developed symptomatic RSV, or other viral LRTIs, had poorer premorbid lung function than infants who did not develop LRTIs during the RSV season.

Methods Lung function (functional residual capacity (FRC), compliance (Crs) and resistance (Rrs) of the respiratory system) was measured at 36 weeks postmenstrual age. After neonatal unit discharge, nasopharyngeal aspirates were obtained whenever the infants had an LRTI, regardless of whether this was in the community or in hospital. Nasopharyngeal aspirates were examined for RSV A and B, rhinovirus, influenza A and B, parainfluenza 1, 2 and 3, human metapneumovirus and adenoivirus.

Results 159 infants with a median gestational age of 34 (range 23–36) weeks were prospectively followed. 73 infants developed LRTIs; 27 had at least one RSV LRTI and 31 had at least one other viral LRTI, but not an RSV LRTI. Overall, there were no significant differences in the FRC (p=0.54), Crs (p=0.11) or Rrs (p=0.12) results between those who developed an RSV or other viral LRTI and those who did not develop an LRTI. Infants with RSV or other viral LRTIs who were admitted to hospital compared with those who were not had higher Rrs results (p=0.033 and p=0.039, respectively).

Conclusion Diminished premorbid lung function may predispose prematurely born infants to severe viral LRTIs in infancy.

INTRODUCTION

Respiratory syncytial virus (RSV) infection in previously healthy infants1–4 and those born prematurely5 is associated with lung function abnormalities at follow-up. The abnormalities include elevated thoracic gas volume and airway resistance at 1 year of age,5–7 airways obstruction in children5 3 and lower forced expiratory volume over 1 s and maximal expiratory flow at 25% of forced vital capacity in young adults.4 It is possible, however, that those lung function abnormalities at follow-up may reflect that infants who develop symptomatic lower respiratory tract infections (LRTIs) have diminished premorbid lung function. In one study,6 term-born infants who developed bronchiolitis in the first year after birth, compared with those who did not, had a non-significant trend to a lower maximal flow at functional residual capacity (Vmax FRC) at 5 weeks of age—that is, before they developed bronchiolitis. In addition, in the Tucson Children’s Respiratory Study, compared with controls, infants who had wheezing LRTIs in their first year had lower respiratory conductances prior to the LRTI7 and infants who had wheezy LRTIs in the first 3 years had lower premorbid Vmax FRCs prior to the LRTI.8 Among infants born before 32 weeks of gestation, those who developed an RSV LRTI had a significantly higher resistance of the respiratory system at 36 weeks postmenstrual age (PMA) at follow-up compared with those who did not.9 The symptomatic ‘RSV LRTI’ group, however, had similar resistance results to the ‘RSV-negative LRTI’ group,9 perhaps suggesting that diminished lung function in prematurely born infants might predispose to other viral LRTIs, as well as RSV LRTI, but the numbers of infants included in the study was small.9 Our aim, therefore, was to test, in a larger cohort, the hypothesis that diminished premorbid lung function would predispose prematurely born infants not only to symptomatic RSV LRTIs, but also to other viral LRTIs. In addition, we wished to determine whether infants who developed severe LRTIs (ie, they required hospitalisation) had poorer lung function than those who did require hospitalisation for the LRTI. The results of such a study would be important to determine whether assessment of lung function might be useful to identify a high risk group who could be targeted for prophylaxis against viral infections. As a consequence, we have related premorbid lung function in prematurely born infants to the occurrence of LRTIs. We have studied infants with a wide range of gestational ages, so additionally we could determine whether maturity at birth influenced any functional predisposition to viral LRTIs.

METHODS

Infants born at <36 weeks of gestational age were eligible for entry into the study if they were born prior to the onset of the RSV season in 2008 or 2009. The RSV season was defined as 1 October to 31 March, consistent with the UK experience.10 Consecutive infants whose parents gave informed written consent were recruited. The study was approved by the Research Ethics Committee of King’s College Hospital NHS Trust.

The infants underwent lung function measurements at 36 weeks PMA while still inpatients on the neonatal unit. The infants were studied while supine and asleep, they were not sedated and none
was ventilated at the time of study. Lung volume was assessed by measurement of FRC, using a commercially available helium gas dilution system (EBS 2615, Equilibrated Bio Systems, New York, USA). The FRC system contained a 500 ml rebreathing bag, the system reservoir, enclosed in an airtight cylinder, and a helium analyser, with a real-time digital display. A facemask (Rendell Baker, Laerdal Norway, facemask size 0 or 1) was held snuggly over the infant’s nose and mouth; silicone putty was used around the mask to achieve an airtight seal. The facemask was connected to the rebreathing bag via a three-way valve. The three-way valve was switched at the end of expiration, so that the infant subsequently breathed from the rebreathing bag. Digital display of the helium dilution curve allowed determination of when equilibration occurred. During the measurement, if there was no change in the helium concentration over a 15 s period, equilibration was deemed to have occurred. The initial and equilibration helium concentrations were used to calculate FRC, which was corrected for oxygen consumption (7 ml/kg/min)\(^1\) and converted to body temperature and water vapour saturated conditions. FRC was measured at least twice in each infant to obtain two results which were within 10% of each other. The paired FRC results were then averaged. The mean intra-subject coefficient of variability of the measurement of FRC was 5%.

Compliance (Crs) and resistance (Rrs) of the respiratory system were measured using the single breath occlusion technique. A facemask was placed over the infant’s nose and mouth; silicone putty was used around the face mask to ensure an airtight seal. A pneumotachograph (Mercury F10L; GM Engineering, Kilwinning, UK) connected to a differential pressure transducer (range: ±2 cm H2O, MP 45, Validyne Engineering, Northridge, California, USA) was inserted into the facemask. The flow signal from the pneumotachograph was integrated to give volume (Validyne CD280: Validyne Engineering). From a sideport on the pneumotachograph, mouth pressure was measured using a differential pressure transducer (range: ±100 cm H2O, MP 45, Validyne Engineering). The signals were amplified (Validyne CD280: Validyne Engineering) and displayed in real time on a computer (Dell Inspiron running Labview software (version 4.0), with 100 Hz analogue to digital sampling (DAQ 16XE-50; National Instruments, Austin TX, USA)).

Oclusions were made at end inspiration, which was identified from the flow signal. The distal end of the pneumotachograph was briefly occluded manually. Only oclusions during which there was no flow, a mouth pressure plateau of at least 100 ms in duration and a linear flow–volume plot after the occlusion was terminated were considered acceptable and analysed. Crs was calculated from the inspiratory volume and the pressure plateau duration and a linear plot, and Rrs were given by the time constant divided by the Crs. The mean Crs and Rrs results were calculated from at least five technically acceptable oclusions. The mean intra-subject coefficients of variability of the Crs and Rrs measurements were 12% and 11%, respectively.

Following neonatal unit discharge, infants were followed prospectively. For the purposes of this study, we assessed their outcomes in their first RSV season. The parents were asked to contact the research team when their infant was symptomatic with signs consistent with an LRTI: cough, wheeze and/or shortness of breath.\(^{12}\) In addition, parents were telephoned every 2 weeks by researchers to ascertain whether their infant had been or was symptomatic. A researcher visited the home on every occasion that an infant had an LRTI, and a nasopharyngeal aspirate (NPA) was obtained. NPAs were also obtained from all infants admitted to hospital with an LRTI. Real-time reverse transcription–PCR was performed on the NPAs for nine viruses (RSV A and B, human metapneumovirus, rhinovirus, influenza A and B, and parainfluenza 1–3) in three multiplexes with a monoplex varicella–zoster virus (VZV) RNA internal control.

The multiplexes tested were:

- **Multiplex 1**: influenza A and B and human metapneumovirus
- **Multiplex 2**: parainfluenza virus types 1, 2 and 3
- **Multiplex 3**: RSV A, RSV B and human rhinovirus

Monoplex: VZV RNA internal control.

The primers were all used at 5 pmol, and the probes at 1 pmol, respectively (see Supplementary table online).

The samples were tested using a Rotor-Gene 6000 (Qiagen, Crawley, UK) and the following conditions: (1) hold at 50°C for 15 min; (2) hold at 95°C for 15 min; and (3) 50 cycles of (a) step 1, 95°C hold for 30 s; (b) step 2, 57°C hold for 30 s, acquiring to cycle A (FAM, Cy5, JOE, ROX).

Adenovirus (DNA virus) was tested by real-time PCR in monoplex with the VZV RNA internal control (also a monoplex). The adenovirus primers were used at 10 pmol and the probe at 2 pmol. The VZV internal control was used as for the multiplexes above. The samples were tested using a Rotor-Gene 6000 (Qiagen) and the following conditions: (1) hold at 98°C for 15 min; and (2) 45 cycles of (a) step 1, 98°C hold for 15 s; (b) step 2, 57°C hold for 20 s, acquiring to cycle A (Cy5, ROX); and (c) step 5, 65°C hold for 20 s.

The internal control was derived from a plasmid construct consisting of a 147 bp fragment of Promega’s pGEM-T (Promega UK Ltd, Southampton, UK) vector with the VZV forward and reverse primer sequences incorporated into the 5\(^\prime\) and 3\(^\prime\) ends forming a chimeric molecule, amplified by the VZV primers and detected by a sequence-specific TaqMan probe (Applied Biosystems, Warrington, UK), based on the original method described previously.\(^{13}\) To produce the RNA internal control, the fragment was cloned into the pGEM-T vector, reverse transcribed with Promega’s Riboprobe in vitro Transcription System, as described by the manufacturer, isolated, purified and quantified using standard molecular biology techniques. Each reaction included the internal control to monitor each sample for potential inhibition and to avoid reporting false-negative results. In a true negative, the internal control will amplify, whereas if the internal control fails to amplify, this would indicate inhibitory compounds in the sample which were preventing the amplification of the nucleic acid from any potential pathogen in the sample.

The neonatal notes were examined to determine if the infant had been exposed to antenatal steroids and/or antenatal infection (maternal positive blood culture, histologically proven chorioamnionitis, maternal temperature with a positive culture from a high vaginal swab or rupture of the membranes of infection (positive blood culture or suspected clinical infection with a raised C-reactive protein, increased or decreased neutrophil count and/or decreased platelet count\(^{15}\)), developed bronchopulmonary dysplasia (BPD; oxygen dependency beyond 28 days) and/or received palivizumab. During the study period, only infants with BPD who had required supplementary oxygen until at least 1 week before neonatal intensive care unit (ICU) discharge and were being discharged during the RSV season were given palivizumab. The decision to admit infants during the follow-up period was made by the clinicians caring for the infants, who were unaware of the lung function test results.
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Analysis
The infants were divided into four groups: (i) infants who never had a symptomatic LRTI—no LRTI; (ii) infants who had symptomatic LRTI(s), but no virus was detected from an NPA—viral-negative LRTI; (iii) infants who had at least one LRTI, from which RSV was detected from the NPA—RSV LRTI; and (iv) infants who had symptomatic LRTI(s) with viruses other than RSV detected from the NPA—other viral LRTI.

Data were tested for normality using the Shapiro–Wilk test. Birth weight and FRC results were normally distributed and differences were assessed for statistical significance using the one-way analysis of variance (ANOVA); all the other results were not normally distributed, hence differences were assessed for statistical significance using the Kruskal–Wallis test. Multiple comparisons were performed using a post hoc Dunn test for multiple comparisons or the Kruskal–Wallis test with a post hoc Bonferroni test for multiple comparisons. The relationship between Rrs and the occurrence of either RSV LRTI or other viral LRTI correcting for gestational age was assessed using analysis of covariance. The threshold for significance was p<0.05. To give an estimate of the value of lung function as a predictor of subsequent severe RSV LRTI (ie, hospitalisation) we calculated and compared areas under receiver operating characteristic curves (AUCs) using SPSS version 15 for statistical significance using the Kruskal–Wallis test.

Sample size
In our previous study we found a difference in the lung function results equivalent to 1 SD between each of the groups. Our planned sample size, therefore, was 160 infants to ensure we had at least 20 infants in each of the RSV and other viral LRTI groups to give 90% power at the 5% level to detect a difference in the lung function results equivalent to at least 1 SD between each of those two groups and the no LRTI group.

RESULTS
A total of 251 infants were eligible for inclusion in the study (figure 1). Seventy-four parents did not consent to their infant taking part in the study, and nine parents consented but then defaulted from follow-up. One infant had a tracheostomy and was not taking part in the study, and nine parents consented but then defaulted from follow-up. One infant died before discharge and two infants died shortly after discharge. The 159 (88 males) infants who completed the study had a median gestational age of 34 weeks (IQR 31–35) and a median birth weight of 1890 g (IQR 1446–2308); none was lost during the follow-up period. The infants who completed the study were of lower gestational age (median gestational age 34 weeks vs 35 weeks, p=0.03) and birth weight (median birth weight 1890 g vs 2132 g, p=0.08) than those who did not take part. Overall, 32 (20%) mothers had had an antenatal infection and 110 (69%) had received antenatal steroids. Thirty-two (20%) infants had received surfactant, 55 (35%) infants had had a postnatal infection and 16 (10%) developed BPD. Seven infants received palivizumab (table 1). One of the seven infants who received palivizumab was admitted to hospital and that was because of an RSV LRTI. Two infants had a nosocomial infection due to rhinovirus while on the neonatal unit; in both cases this was after lung function testing. One of the infants subsequently developed an RSV LRTI and their results were analysed in the RSV group; the other did not subsequently have an RSV infection and their results were analysed in the other viral LRTI group.

The only significant differences in the demographics of the four groups were with regard to birth weight (p=0.02) and the number of infants receiving surfactant (p=0.03) (table 1). Those differences were due to the other viral LRTI group being of lower birth weight and the others having received surfactant.

Table 1 Demographic data

<table>
<thead>
<tr>
<th></th>
<th>No LRTI (n=86)</th>
<th>Viral-negative LRTI (n=15)</th>
<th>RSV LRTI (n=27)</th>
<th>Other viral LRTI (n=31)</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gestational age (weeks)</td>
<td>34 (32–35)</td>
<td>34 (31–35)</td>
<td>33 (30–34)</td>
<td>33 (29–35)</td>
<td>0.11</td>
</tr>
<tr>
<td>Birth weight (g)</td>
<td>2070 (1605–2413)</td>
<td>2026 (1586–2312)</td>
<td>1830 (1335–2085)</td>
<td>1565 (1261–2152)</td>
<td>0.02</td>
</tr>
<tr>
<td>Males</td>
<td>46 (56%)</td>
<td>9 (60%)</td>
<td>14 (52%)</td>
<td>17 (55%)</td>
<td>0.97</td>
</tr>
<tr>
<td>Antenatal smoking</td>
<td>14 (16%)</td>
<td>2 (13%)</td>
<td>4 (15%)</td>
<td>7 (23%)</td>
<td>0.82</td>
</tr>
<tr>
<td>Antenatal steroids</td>
<td>52 (60%)</td>
<td>11 (73%)</td>
<td>22 (81%)</td>
<td>25 (81%)</td>
<td>0.07</td>
</tr>
<tr>
<td>Surfactant</td>
<td>12 (14%)</td>
<td>2 (13%)</td>
<td>6 (22%)</td>
<td>12 (39%)</td>
<td>0.03</td>
</tr>
<tr>
<td>Duration of ventilation (days)</td>
<td>0 (0–2)</td>
<td>1 (0–1.5)</td>
<td>1 (0–5)</td>
<td>1 (0–4.5)</td>
<td>0.25</td>
</tr>
<tr>
<td>Bronchopulmonary dysplasia</td>
<td>6 (7%)</td>
<td>1 (7%)</td>
<td>3 (11%)</td>
<td>6 (19%)</td>
<td>0.25</td>
</tr>
<tr>
<td>Parental atopy</td>
<td>43 (50%)</td>
<td>6 (40%)</td>
<td>9 (33%)</td>
<td>12 (42%)</td>
<td>0.42</td>
</tr>
<tr>
<td>Day care</td>
<td>2 (2%)</td>
<td>0 (0%)</td>
<td>1 (4%)</td>
<td>2 (6%)</td>
<td>0.61</td>
</tr>
<tr>
<td>Number of siblings</td>
<td>1 (0–2)</td>
<td>2 (1–2)</td>
<td>1 (0–2)</td>
<td>1 (0–2)</td>
<td>0.17</td>
</tr>
<tr>
<td>Palivizumab</td>
<td>1 (1%)</td>
<td>1 (7%)</td>
<td>1 (4%)</td>
<td>4 (13%)</td>
<td>0.05</td>
</tr>
</tbody>
</table>

Data are expressed as median (IQR) or n (%). LRTI, lower respiratory tract infection; RSV, respiratory syncytial virus.
birth weight ($p<0.05$) and a greater proportion of them having received surfactant ($p<0.05$) than the no LRTI group.

On 158 occasions, 73 infants had an LRTI and on 94 occasions a virus was detected (table 2). Twenty-seven infants had at least one RSV-positive LRTI (some also had other viral LRTIs); 11 required hospital admission (nine for RSV LRTI (one of those infants required intensive care) and two for non-respiratory causes). The infants who were admitted compared with those who were not admitted at an earlier gestational age (median (IQR) 30 (29–35) weeks vs 34 (33–35) weeks, $p=0.004$), were of lower birth weight (1446 (1020–1515) g vs 1852 (1515–2100) g, $p=0.047$), had required more days of ventilation on the neonatal unit (6 (1.5–220) days vs 0 (0–1) days, $p=0.011$) and more days of supplementary oxygen on the neonatal unit (7 (1–49) days vs 0 (0–1) days, $p=0.011$), and a greater proportion had had BPD (35% vs 0%, $p=0.015$). They, however, had not had more NPIAs taken ($p=0.22$). The age at LRTI presentation also did not differ significantly between those who were and were not admitted, the mean age in both groups being 4.2 months ($p=0.91$).

Thirty-one infants suffered at least one other viral LRTI, but not an RSV LRTI; three required hospital admission (two for LRTI and one for non-respiratory causes). There were no significant differences between those who were and were not admitted with regard to gestational age at birth, birth weight, duration of ventilation and supplementary oxygen on the neonatal unit, BPD, number of NPIAs taken and age at LRTI presentation. Fifteen infants had LRTIs with no virus identified; four required hospital admission (two for LRTI and two for non-respiratory causes). Eighty-six infants did not have an LRTI and one for non-respiratory causes. Eleven infants had at least one dual infection and two had a triple infection (RSV A/human metapneumovirus/adenovirus and RSV B/parainfluenza 1/adenovirus). The infant with the triple infection due to RSV A/human metapneumovirus/adenovirus was the only infant who was admitted to an ICU.

The only significant differences between the lung function results of the four groups (table 3 and figure 2), even after correcting for gestational age, were the ‘raw’ $Crs$, $FRC$, $Crs$, $Rrs$, $Res$, $RLTI$, $LRTI$, $respiratory tract infection$, $rs$, $respiratory syncitial virus$.
admission. In this study, which included infants with a broader range of gestational ages, we highlight the Rrs results to be significantly higher in those infants who were admitted to hospital with an RSV or another viral LRTI—that is, they were likely to have had more severe infection.

We have demonstrated that infants with RSV or other viral LRTIs who were hospitalised compared with those who were not had significantly worse premorbid lung function—that is, higher Rrs results and significantly lower Crs results (not corrected for body weight). In the RSV LRTI groups, those who were admitted to hospital compared with those who were not were born at a significantly earlier gestational age and of lower birth weight, required significantly longer durations of ventilation and supplementary oxygen and a significantly greater proportion had developed BPD. It is, therefore, not surprising that they had significantly worse lung function at 36 weeks PMA. Comparison of the AUCs for gestational age and either Rrs or Crs—that is, they performed similarly. We thus suggest lung function testing at 36 weeks PMA may not only indicate a group at highest risk, but also indicates a mechanism by which prematurely born infants develop severe RSV LRTIs—that is, they are more likely to develop a severe RSV LRTI if they have significantly poorer premorbid lung function.

Lung function was assessed by measuring Crs, Rrs and FRC. The FRC and Crs results were corrected for body weight, as length can be difficult to measure accurately in small infants. In the ‘thinnest’ infants, FRC and Crs may be overestimated when a weight correction is used. We have also given the uncorrected data, and the median Crs, but not FRC results, differ significantly in the RSV group between those who were and were not admitted, further emphasising they had worse lung function. Rrs results are not routinely corrected for either body weight or length and differed significantly in both the RSV and the other viral groups between those who were and were not admitted. There was a wide range of results, reflecting the wide spread of gestational ages of the infants we examined. The low intra-observer coefficients of variability, however, demonstrate our results to be reliable. We studied the infants in the neonatal ICU and were unable to measure \( \text{T}_{\text{max}} / \text{T}_{\text{E}} \). A criticism, therefore, of our study is that we cannot exclude that there may be differences in small airway function between the two groups. Our study, however, has a number of strengths. We prospectively followed a large cohort of infants and thus had sufficient numbers to be able to appropriately compare results from infants with RSV LRTI or other viral LRTI with those with no LRTI. The infants, however, were only prospectively followed during the RSV season and it is possible the infants suffered other viral infections subsequently. In addition, only two infants were admitted to hospital with other viral LRTIs and thus it would be important to determine whether our results are replicated in a larger number of infants admitted to hospital with other respiratory viral infections. We used PCR techniques that could detect 10 respiratory viruses. The PCR techniques are more sensitive than conventional virus isolation or immunofluorescence techniques for the diagnosis of respiratory virus infections in children. A further strength of our study is that parents were asked to contact us whenever their infant was symptomatic and, in addition, they were reminded by 2-weekly telephone calls. As a consequence, we were able to document LRTIs not only in hospital but also in the community.

In conclusion, our results suggest that, overall, prematurely born infants are not predisposed by diminished premorbid lung function to RSV or other viral LRTIs, but rather they are predisposed to severe LRTIs—that is, those that require hospitalisation.

### Table 4 Previous studies reporting lung function prior to lower respiratory tract infections (LRTIs) in infancy

<table>
<thead>
<tr>
<th>References</th>
<th>Sample size</th>
<th>GA at birth</th>
<th>PMA at assessment (weeks)</th>
<th>Viral identification</th>
<th>Lung function test results</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Young et al</td>
<td>253</td>
<td>Term</td>
<td>45</td>
<td>RSV (n=2)</td>
<td>Bronchiolitis (n=10)</td>
<td>Control (n=236)</td>
</tr>
<tr>
<td>Martinez et al</td>
<td>124</td>
<td>Term</td>
<td>48</td>
<td>Virology not reported</td>
<td>LRRTI with wheeze (n=24)</td>
<td>No LRTI (n=88)</td>
</tr>
<tr>
<td>Broughton et al</td>
<td>39</td>
<td>28</td>
<td>36</td>
<td>RSV (n=15)</td>
<td>RSV LRTI (n=15)</td>
<td>No RSV LRTI (n=24)</td>
</tr>
</tbody>
</table>

Data are demonstrated as mean (SD).

Crs, compliance of the respiratory system; FRC, functional residual capacity; GA, gestational age; He, helium dilution; NS, non-significant; Rrs, resistance of the respiratory system; RSV, respiratory syncytial virus; \( \text{T}_{\text{max}} / \text{T}_{\text{E}} \), the time to maximal expiratory flow as a proportion of total expiratory time.
A role for cigarette fumes in the development of emphysema through reduced alveolar cell proliferation and upregulation of apoptosis

This paper describes a molecular mechanism for the loss of alveolar tissue in cigarette smoke-induced lung injury. Under the influence of adverse environmental conditions including hypoxia, the intracellular stress protein Rtp801 is transcribed. This study sought to demonstrate that Rtp801 downregulates the kinase mammalian target of rapamycin (mTOR) which is responsible for alveolar cell proliferation and upregulates the apoptotic and inflammatory pathways through the activation of the transcription factor nuclear factor kappa B (NF-kB).

Increased levels of Rtp801 were measured in alveolar cells of patients with chronic obstructive pulmonary disease compared with healthy individuals through histological staining and electrophoresis. Western blotting studies in mice showed significantly increased Rtp801 levels following exposure to cigarette smoke for 7 days. Mice treated with the antioxidant N-acetyl-L-cysteine followed by exposure to smoke showed no overall increase in Rtp801 levels. Overexpression of Rtp801 in mice without exposure to smoke led to significant alveolar apoptosis. Rtp801 knock-out mice were protected from alveolar inflammatory and apoptotic changes in the presence of smoke. Inhibition of NF-kB activity resulted in decreased levels of inflammation and apoptosis.

These results confirm the role of this cellular stress response in alveolar damage and provide further evidence for a genetic component to emphysema.


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