Pravastatin attenuates allergic airway inflammation by suppressing antigen sensitisation, interleukin 17 production and antigen presentation in the lung

M Imamura, K Okunishi, H Ohtsu, K Nakagome, H Harada, R Tanaka, K Yamamoto, M Dohi

ABSTRACT

Background: Statins are widely used to treat hyperlipidaemia. Their immunosuppressive effect has recently been confirmed in various immune mediated disease models. However, relatively few studies have been conducted on allergic inflammation, so the precise mechanisms of their actions against allergies have not been fully clarified. On the other hand, the role of interleukin (IL)17 in immune responses has been recently highlighted, but whether statins affect IL17 production has not been well studied. The effect of pravastatin on allergic airway inflammation in a mouse model was examined to elucidate the mechanism of action, focusing on its effect on IL17 production.

Methods: BALB/c mice were immunised with ovalbumin (OVA) and then challenged with OVA aerosol. Pravastatin was delivered by intraperitoneal injection during either sensitisation or the challenge.

Results: When delivered during systemic sensitisation, pravastatin suppressed OVA induced proliferation and production of Th2 type cytokines such as IL5 in spleen cells ex vivo and in vitro. IL17 production was also suppressed. Furthermore, pravastatin delivered during the inhalation of OVA attenuated eosinophilic airway inflammation, OVA specific IgE production in serum and OVA induced IL17 production in the thoracic lymph node. We also found that pravastatin attenuated the antigen presenting capacity of CD11c+ cells obtained from the OVA challenged lung.

Conclusion: Pravastatin suppresses the systemic sensitisation to allergen with downregulation of IL17 production. It also suppresses an ongoing immune response in the airway partly by suppressing antigen presentation in the lung. Therefore, statins could be a novel therapeutic option for treatment of asthma.

Statins are inhibitors of the 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase, and have been widely prescribed to treat hyperlipidaemia. Large clinical trials have demonstrated that statins can be used for primary and secondary prevention of cardiovascular diseases.

In addition to their conventional lipid lowering properties, statins are now recognised as possessing a variety of immunomodulatory effects. Statins downregulate various functions of antigen presenting cells (APCs), such as dendritic cells and B cells. Furthermore, statins promote in vitro T helper (Th)2 polarisation from naïve CD4+ T cells via suppression of Th1 development. In animal models of Th1 biased autoimmune disorders such as experimental autoimmune encephalomyelitis and collagen induced arthritis, statin attenuates disease severity with a shift from a Th1-type to Th2-type cytokine profile.

Allergic bronchial asthma is a Th2 mediated disease, and statins will exaggerate allergic inflammatory responses in the airway. However, in several reports, statins suppressed Th2 type airway inflammation. Simvastatin and pravastatin reduced production of Th2-type cytokines, thus attenuating allergic airway inflammation. However, the precise mechanisms of action have not yet been fully clarified. Further details should be clarified for clinical application.

A significant role for interleukin (IL)17 producing CD4+ T cells (Th17 cells) has recently been highlighted. Generally, IL17 is critical to host defence against various infections. Furthermore, IL17 affects various immune responses. Many immunological processes have recently been re-evaluated in the context of Th17 and regulatory T cells. For example, both the encephalomyelitis model and collagen induced arthritis model have been considered Th17 mediated models rather than conventional Th1 models. IL17 also plays an important role in neutrophil dominated inflammatory responses in the lung. Recently, the role of IL17 in asthma has also been intensively studied. For example, elevated sputum IL17 concentrations correlate with clinical severity. The role of IL17 in an animal model of allergic airway inflammation has been investigated in several studies. In one report, IL17 was essential for sensitisation to the antigen whereas it attenuated airway inflammation in the effector phase. At present, the precise role of IL17 in allergic airway inflammation is not fully clear. In addition, whether statins affect IL17 mediated responses has not been well studied.

This study was performed to examine the hypothesis that pravastatin attenuates allergic airway inflammation through suppression of IL17 production. We conducted research in vitro and in vivo using an animal model. We found that pravastatin suppressed systemic sensitisation to the antigen with downregulation of IL17 production, and thus suppressed antigen induced eosinophilic airway inflammation, partly by regulating antigen presentation in the lung.

METHODS

Mice

Male BALB/c mice, 7 weeks of age, were obtained from Charles River Japan (Kanagawa, Japan). Male BALB/c T cell receptor transgenic DOI11.10 mice,
Aged 6–8 weeks, were purchased from The Jackson Laboratory (Bar Harbor, Maine, USA). They were maintained under conventional animal housing conditions in a specific pathogen free setting. All experiments were approved and performed according to the guidelines of the University of Tokyo.

**Study protocol in vivo**

BALB/c mice were immunised with an intraperitoneal injection of 2 µg of ovalbumin (OVA; Sigma-Aldrich, St Louis, Missouri, USA)/2 mg of aluminium hydroxide (alum) on days 0 and 11. Control mice received saline instead of the OVA/alum solution. The animals were challenged with an aerosolised solution of 5% w/v OVA in phosphate buffered saline (PBS) for 10 min from day 18 to day 20. Pravastatin (10 mg/kg, kindly provided by Sankyo Co, Tokyo, Japan) dissolved in 0.5 ml of saline, or saline alone, was given by intraperitoneal injection 30 min before inhalation of OVA on days 18, 19 and 20. Control mice inhaled PBS on days 18–20. On day 21, samples of serum, bronchoalveolar lavage fluid (BALF) and lungs were obtained. The lungs were cut out and used for analyses of antigen presentation. In another experiment, left lungs were homogenised in 1.0 ml of PBS containing 0.5% Triton X-100 and complete protease inhibitor cocktail (Roche, Basel, Switzerland). The lung homogenates were cleared of debris and cells by centrifugation at 10 000×g for 10 min. Thoracic lymph nodes were also obtained and used for ex vivo analyses. Cell counts and cell differentials of BALF were determined as previously reported.28–30 Cytokine concentrations in BALF, lung homogenates and immunoglobulin levels in sera were measured by enzyme linked immunosorbent assay (ELISA). The numbers of animals were six in the saline treated or pravastatin treated sensitised groups, respectively. We conducted this experiment more than four times and confirmed the reliability of the data.

**Quantification of cytokines, cell proliferation and cholesterol**

Cytokine concentrations in BALF, serum and cell culture supernatants were determined by ELISA31 (see methods in the online data supplement).

**Preparation of single cell suspensions of spleen and lymph node cells**

Spleens were collected and incubated at 37°C for 15 min after treatment with 0.1% (w/v) collagenase (Sigma-Aldrich)/complete Dulbecco’s modified Eagle’s medium (DMEM) solution and then minced. Lymph node cells were collected and minced and then incubated at 37°C for 30 min with 0.035% collagenase/complete DMEM solution. Single cell suspensions were prepared with a cell strainer. Red blood cells were removed by hypotonic lysis. After two washes, the cells were used for experiments. Complete DMEM was used for incubation throughout the study.28–30

**Response of spleen and lymph node cells to OVA**

Spleen or lymph node cells (2.5×106 cells/ml) were cultured in a 96 well, flat bottomed microtitre assay plate with OVA (100 µg/ml) in an incubator (37°C, 5% CO2, 90% humidity). Cell proliferation was measured on day 3. On day 4, cytokine production was measured by ELISA.

**Animal preparation for ex vivo analyses**

BALB/c mice were immunised with OVA/alum on days 0 and 11. On days 11–17, some mice received pravastatin (10 mg/kg) dissolved in 0.5 ml of saline or saline alone by intraperitoneal injection. Control mice received saline on days 0 and 11. On day 18, spleens were collected and cell proliferation and cytokine production were measured.

**Effects of pravastatin on antigen presentation in the lung**

On day 21, CD11c+ antigen presenting cells in the lungs of mice were positively selected, as reported previously.28–30 In brief, lung tissues were minced and then treated with 0.055% (w/v) collagenase/complete DMEM solution for 30 min. Single cell suspensions of the tissues were obtained, and CD11c+ cells were positively selected using MACS CD11c microbeads (Miltenyi Biotec, Auburn, California, USA). The populations of cells selected from lung tissues were routinely ~70% CD11c+, and there was no significant difference in the purity of CD11c+ cells between the groups of mice. Preliminary experiments demonstrated that almost 70% of the CD11c+ cells expressed the major histocompatibility complex class II molecule I-A/I-E. Some CD11c+ cells co-expressed F4/80 or Gr-1 (1–2%), an indicator of macrophages and granulocytes, respectively. Lung CD11c+ cells had negligible expression of CD4, CD8 and CD19 (data not shown). These results suggest that although lung CD11c+ cells contain some subsets, the main population of CD11c+ cells were dendritic cells. These CD11c+ cells were incubated with mitomycin C (10 µg/ml; Sigma-Aldrich) for 35 min at 37°C in a humidified atmosphere containing 5% CO2 and then washed four times with PBS. In addition, CD4+ T cells were obtained from spleen cells of the DO11.10 mice using anti-mouse CD4 colloidial superparamagnetic microbeads (Miltenyi Biotec), as previously reported.28–30 The purity of CD4+ cells, confirmed by flow cytometry, was >95%. For measuring antigen presenting capacity, lung CD11c+ cells (0.25×106, 0.8×104, and 2.5×104 cells/ml) obtained from each group of mice were cocultured with CD4+ T cells (2.5×104 cells/ml) selected from the spleens of DO 11.10 mice. After a 2 day coculture, proliferation of CD4+ T cells was measured.

**Statistical analysis**

We applied data from at least three sets of samples from individual animals to statistical analysis, while we did not apply data from individual culture wells. Values are expressed as mean (SD) or individual points. The Komolgorov–Smirnov test was used to check the normal distribution of data. For all variables, the numbers of lymphocytes and IL15 levels in BALF rejected normality. Differences in variables between the groups were determined by the Student’s t test or the Mann–Whitney U test for variables with or without normal distribution, respectively. Values of p<0.05 were considered significant.

**RESULTS**

Pravastatin decreased eosinophilic inflammation in the lung and suppressed total IgE, OVA specific IgE and IgG1 levels in serum

Treatment with pravastatin significantly suppressed eosinophilic airway inflammation induced in OVA treated mice (fig 1A, 1B). We conducted this experiment repeatedly, and the numbers of eosinophils in the BALF of mice treated by pravastatin were 30–60% of those of OVA treated mice. Concentrations of IL13 and transforming growth factor β in BALF were also significantly suppressed (fig 1C, 1D). Although the IL17 content of the lung increased in the OVA treated mice (fig 1A, 1B). We conducted this experiment repeatedly, and the numbers of eosinophils in the BALF of mice treated by pravastatin were 30–60% of those of OVA treated mice. Concentrations of IL13 and transforming growth factor β in BALF were also significantly suppressed (fig 1C, 1D). Although the IL17 content of the lung increased in the OVA treated mice compared with the saline treated control mice, it did not seem to be affected by pravastatin (fig 1E). Pravastatin treatment significantly reduced serum total IgE, OVA specific IgE and OVA specific IgG1 levels, while no significant difference was observed in the total IgG and asthma.
OVA specific IgG levels (fig 2A–E). Pravastatin did not affect total serum cholesterol levels (saline treated mice 110.4 (12.4) mg/dl; OVA treated mice 102.6 (7.0) mg/dl; pravastatin treated mice 106.3 (11.0) mg/dl). Therefore, the suppressive effect was independent of serum cholesterol levels.

Treatment with pravastatin during systemic sensitisation suppresses the antigen induced immune response of spleen cells

Cell proliferation and production of IL5, IL10 and IFN-γ by splenocytes obtained from OVA treated mice were increased compared with saline treated mice (fig 3A–D). IL17 production was also increased in OVA treated mice, and pravastatin seemed to suppress IL17 production (fig 3E). Furthermore, pravastatin treatment tended to reduce these responses to OVA except the production of IFN-γ (fig 3A–E). In another experiment, pravastatin treatment in vitro tended to suppress cell proliferation and production of IL5, IL17 and IL10 in a dose dependent manner (see supplementary fig 1 with method online). Hence treatment with pravastatin during a second systemic sensitisation attenuated the Th2-type antigen induced immune response with suppression of IL17 production.

OVA specific IgG levels (fig 2A–E). Pravastatin did not affect total serum cholesterol levels (saline treated mice 110.4 (12.4) mg/dl; OVA treated mice 102.6 (7.0) mg/dl; pravastatin treated mice 106.3 (11.0) mg/dl). Therefore, the suppressive effect was independent of serum cholesterol levels.

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Pravastatin treatment during OVA challenge suppresses antigen induced IL17 production of thoracic lymph node cells

Pravastatin treatment during OVA challenge attenuated antigen induced IL17 production of lymph node cells. (A, B) On day 21, lymph node cells from OVA mice treated with pravastatin (PR) or control vehicle (OVA) were incubated with OVA. After 3 days of incubation, production of IL5 (A) and IL17 (B) was measured. Data were obtained from six wells per group of mice. All results are representative of three independent experiments.

DISCUSSION

The results of the present study demonstrated that pravastatin suppresses sensitisation to OVA antigen with downregulation of IL17 production, and that it also attenuates the ongoing immune response in the airway. Pravastatin, delivered during systemic immunisation, attenuated the OVA induced Th2 type response with suppression of the production of IL17. In addition, pravastatin delivered during the airway challenge with OVA suppressed eosinophilic airway inflammation by affecting IL17 production, and suppressing the antigen presenting capacity of CD11c+ cells in the lung. As far as we know, this is the first study to clarify the effect of any statin treatment in vivo on IL17 production in an immune response and its effect on antigen presentation in the lung.

The immunomodulating effect of statins involves the inhibition of Th1-type responses. In addition, recent reports demonstrate that they can also suppress Th2-type responses. To date, two studies with simvastatin and one report with pravastatin demonstrated that statins suppressed the Th2-type allergic airway inflammation. In the current study, pravastatin tended to suppress OVA induced cell proliferation and cytokine production by spleen cells both ex vivo (fig 3) and in vitro (see supplementary fig 1 with method online). Pravastatin also suppressed eosinophilic airway inflammation, IgE production in serum and cytokine production from immunocytes in the thoracic lymph node (figs 1, 2, 4). These results are reasonably consistent with the findings of previous reports, confirming that statins certainly suppress Th2-type allergic airway inflammation in an experimental system.

To date, only one study has examined the effect of pravastatin on allergic airway inflammation. Yeh and Huang reported that treatment of mice with pravastatin from 2 weeks before antigen sensitisation to after an antigen challenge reduced airway eosinophilia. In a preliminary experiment,
CD4 or saline (ovalbumin (OVA)) were positively selected and cocultured with presenting cells (APCs) in the lung of mice treated with pravastatin (PR) muscle cells. These studies indicate that IL17 may enhance that IL17 plays a critical role in antigen sensitisation.

These findings support the idea that IL17 is important in the clinical situation. In a preliminary study, treatment with 2 mg/kg/day of pravastatin did not attenuate allergic airway inflammation (data not shown). The dose of 10 mg/kg/day of pravastatin in this study is higher than the dose used clinically in human. The clinical dose of pravastatin in Japan is about 0.2–0.4 mg/kg/day, and the concentration in serum is 0.1–1 μM. On the other hand, the doses of statins used in most animal studies were higher than the clinical dose, and high doses of statins could suppress the activities of autoimmune disease, but did not show cholesterol lowering effects in mice.

As animal models have limitations to adaptation to human diseases, clinical studies need to be performed. To date, only one clinical study has examined the effect of statins on chronic asthma. Menzies et al reported that simvastatin treatment for 1 month did not exhibit significant therapeutic anti-inflammatory effects. However, in their study, the number of patients enrolled in the trial was small, and simvastatin was administered during a stable period of asthma. To draw a definite conclusion, further clinical studies in various clinical situations are necessary.

In conclusion, we demonstrated that pravastatin suppresses eosinophilic airway inflammation by attenuating the sensitisation to allergens with suppression of IL17 production and by regulating lung dendritic cells in a murine model. Although further investigation is needed to confirm this possibility, our results indicate the therapeutic potential of statins as anti-inflammatory drugs for asthma.

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Competing interests: None.

Ethics approval: All experiments were approved by the Animal Research Ethics Board of the Department of Allergy and Rheumatology, University of Tokyo.

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