ACUTE LUNG INJURY

In vitro and in vivo effects of salbutamol on neutrophil function in acute lung injury

G D Perkins, N Nathani, D F McAuley, F Gao, D R Thickett

Background: Intravenous salbutamol (albuterol) reduces lung water in patients with the acute respiratory distress syndrome (ARDS). Experimental data show that it also reduces pulmonary neutrophil accumulation or activation and inflammation in ARDS.

Aim: To investigate the effects of salbutamol on neutrophil function.

Methods: In vitro effects of salbutamol on neutrophil function were determined. Blood and bronchoalveolar lavage (BAL) fluid were collected from 35 patients with acute lung injury (ALI)/ARDS, 14 patients at risk from ARDS and 7 ventilated controls at baseline and after 4 days’ treatment with placebo or salbutamol (ALI/ARDS group). Alveolar–capillary permeability was measured in vivo by thermodilution (PiCCO). Neutrophil activation, adhesion molecule expression and inflammatory cytokines were measured.

Results: In vitro, physiological concentrations of salbutamol had no effect on neutrophil chemotaxis, viability or apoptosis. Patients with ALI/ARDS showed increased neutrophil activation and adhesion molecule expression compared with at-risk-patients and ventilated controls. There were associations between alveolar–capillary permeability and BAL myeloperoxidase (r = 0.4, p = 0.038) and BAL interleukin 8 (r = 0.38, p = 0.033). In patients with ALI/ARDS, salbutamol increased numbers of circulating neutrophils but had no effect on alveolar neutrophils.

Conclusion: At the onset of ALI/ARDS, there is increased neutrophil recruitment and activation. Physiological concentrations of salbutamol did not alter neutrophil chemotaxis, viability or apoptosis in vitro. In vivo, salbutamol increased circulating neutrophils, but had no effect on alveolar neutrophils. These data suggest that the beneficial effects of salbutamol in reducing lung water are unrelated to modulation of neutrophil-dependent inflammatory pathways.

MATERIALS AND METHODS

In vitro studies

Under-agarose chemotaxis, cell viability and apoptosis

Neutrophils from non-smoking, healthy controls were purified from peripheral blood by discontinuous Percoll density gradients. The effect of salbutamol on the chemotactic activity of purified neutrophils was measured using the under-agarose method.

Abbreviations: ALI, acute lung injury; ARDS, acute respiratory distress syndrome; BAL, bronchoalveolar lavage; FMLP, N-formyl-l-leucin-methionyl-l-phenylalanine; LPS, lipopolysaccharide; TNF, tumour necrosis factor
Effects of salbutamol on neutrophil function

Method. The freshly harvested neutrophils were resuspended at 5 × 10⁷ cells/ml in RPMI (Sigma, UK) 1640 culture medium or 10⁻⁵-10⁻¹⁰ M salbutamol in RPMI (with or without propranolol 10⁻⁸ M) before seeding in 10-μl aliquots in the central wells of a freshly prepared agarose plate. In all, 10 μl of RPMI was placed in each of the inner wells and 10 μl of chemotractant (10⁻⁷ M formyl-L-leucin-methionyl-l-phenylalanine (FMLP, Sigma, UK) in RPMI) in each of the outer wells. Negative controls contained RPMI in the inner and outer wells. The plates were incubated at 37°C (5% CO₂) for 2 h and then flooded with methanol for overnight fixation of the cells. The gel was carefully removed and the plates washed under slow running tap water. The plates were then stained with Gram stain and left to dry. For each well, the chemotactic and chemokinetic responses were read using an eyepiece graticule. The overall response to the chemotractant (chemotactic differential) was calculated by subtracting the chemokinetic response from the chemotactic response.

Purified neutrophils, suspended in RPMI at a concentration of 1 × 10⁶ ml, were incubated with 10⁻⁵ and 10⁻⁷ M salbutamol or RPMI control for 2 h at 37°C in humidified 5% CO₂. Cell proliferation was assessed by adding 20 μl of CellTiter 96 AQueous one solution (Promega, Southampton, UK) to 100 μl of cell suspension in a 96-well culture plate (Nunc). The reaction was allowed to proceed for 2 h at 37°C in 5% CO₂. The cell titre solution contains a tetrazolium compound that is metabolised by healthy cells to a formazan product, the absorbance of which was read at 495 nm on an MRX-II 96-well plate reader (Dynex Technologies, Sussex, UK).

Purified neutrophils, suspended in RPMI with 10% heat-inactivated fetal calf serum (Life Technologies and 100 units/ml penicillin and 100 mg/ml streptomycin (Sigma, Poole, UK) at a concentration of 1 × 10⁶/ml, were incubated with 10⁻⁵ and 10⁻⁷ M salbutamol or RPMI control for 18 h at 37°C in humidified 5% CO₂. Preliminary experiments showed that 18 h was the optimal time point for assessing apoptosis. The number of normal and apoptotic neutrophils were determined by morphological patterns. After centrifugation, samples were stained using a commercial May–Grunwald Giemsa stain (Diff-Quick, Baxter Healthcare Product, Deerfield, Illinois, USA). The percentage apoptotic cells compared with total number of cells were calculated. The morphological results were confirmed using flow cytometry and the annexin V/propidium iodine apoptosis kit (Dako, Carpinteria, California, USA). The percentages of live (annexin and propidium iodine negative) and early apoptotic (annexin positive, propidium iodine negative) or late apoptotic/dead (annexin positive, propidium iodine positive) cells were calculated.

Clinical study
Mechanically ventilated adult patients enrolled in the BALTI-1 study were eligible for inclusion. This study recruited patients within 48 h of the onset of ALI and ARDS and randomised them to 7 days of treatment with intravenous salbutamol (15 μg/kg/h). ALI was defined according to the American European Consensus Conference definition as the acute onset of respiratory failure with a arterial oxygen tension:fractional oxygen tension (PaO₂:FiO₂) ratio of <300 mm Hg and bilateral infiltrates on the chest radiograph in the absence of clinical evidence of left atrial hypertension. ARDS was considered present when the PaO₂:FiO₂ ratio was <200 mm Hg. The exclusion criteria were as follows: age <18 years; participation in other intervention trials; severe obstructive airway disease requiring nebulised or intravenous β₂-agonist; treatment with β-blockers within 48 h; neutrophil count >5 × 10⁷/l; brain stem death; treatment withdrawal within 24 h; immunosuppression (steroids >20 mg/day, chemotherapy or other immunosuppressive agents within 2 weeks); lobectomy/pneumonectomy; burns >40% body surface area; assent declined from the next of kin.

Patients with identifiable risk factors for ALI/ARDS, but who, at the time of recruitment had not met the criteria for
ALI/ARDS, were included as an at-risk population. Non-smoking, age-matched ventilated patients undergoing elective surgery were recruited as controls.

**Biological sample collection and processing**

BAL using 150 ml cold saline was performed immediately after recruitment in at-risk patients and ventilated controls and at baseline and on day 4 in patients with ALI/ARDS. BAL fluid was kept on ice until transferred to the laboratory for processing. Blood was simultaneously collected in lithium heparin tubes (Becton-Dickinson, Plymouth, UK) and placed immediately on ice until processed.

The BAL fluid was filtered through coarse surgical gauze to remove mucus and other debris. The fluid was then centrifuged at 500 g for 5 min in a centrifuge prechilled to 4°C. The supernatant was removed and immediately frozen to −80°C and stored for subsequent analysis. The cell pellet was resuspended in 10 ml phosphate-buffered saline supplemented with 1% human serum albumin. Total cell count was determined using a haemocytometer. Cell viability was measured by the ability of live cells to exclude Trypan Blue. Cell purity was measured on a cytospin preparation stained with DiffQuick (Baxter, Berkshire, UK) as described previously. ²³

**Patient characteristics**

Patient demographic characteristics were recorded at baseline. The acute physiology and chronic health evaluation II (APACHE II) and simplified acute physiology score II (SAPSII) scores, and predicted intensive care unit mortality were recorded as global markers of disease severity. ²⁶ The Murray Lung Injury Score and PaO₂/FIO₂ ratio were collected as markers of the severity of lung injury.

**Alveolar–capillary permeability**

The single indicator transpulmonary thermodilution system (PiCCO; Pulsion Medical Systems, Munich, Germany) was used to calculate an in vivo alveolar–capillary permeability index. The permeability index was derived from the ratio of extravascular lung water to pulmonary blood volume. Previous studies have shown that this index can separate cardiogenic (low permeability) and inflammatory (high permeability) causes of pulmonary oedema. ²⁵

**Flow cytometry analysis**

Flow cytometry was performed using the whole-blood technique. ²⁶ Aliquots of 100 μl of whole blood or BAL cells (suspended in 1% human serum albumin (Sigma Chemicals) at a final concentration of 1×10⁶/ml) were fixed by adding 100 μl of 1% paraformaldehyde for 15 min. Immunofluorescent staining was performed by adding fluorescin isothiocyanate isomer-conjugated monoclonal antibodies directed against the following: CD11B (IgG1, Dako, Ely, UK); CD18 (IgG1, Dako); CD49 (IgG1, Serotec, Oxford, UK); CD64 (IgG1, Serotec) and l-selectin, IgG1, (Becton-Dickinson). Stimulated neutrophil l-selectin expression was determined by incubating whole blood with FMLP (10⁻⁶ M) for 15 min before fixation. Controls were included using isotype-matched, irrelevant antibodies to human IgG1 (Dako). After incubation with the appropriate monoclonal antibody for 30 min at room temperature (protected from light), 1.5 ml of FACS brand lysing solution (Becton-Dickinson) was added to each tube for 5 min. The cells were then washed twice with 1.5 ml of wash buffer (500 ml phosphate-buffered saline, 0.1 g sodium azide (Sigma Chemicals), 5 g bovine serum albumin (Sigma)). After the second wash, the cell pellet was resuspended in 500 μl of 1% paraformaldehyde. Samples were analysed on a Becton-Dickinson 440 flow cytometer. The neutrophil cell populations were identified and gated from the forward and side light scatter. The median intensity of fluorescence for cells labelled with specific antibody was determined relative to the median intensity of fluorescence for cells labelled with the non-specific isotype control antibody.

**Tumor necrosis factor α, interleukin 8, MPO and salbutamol assays**

Tumor necrosis factor α (TNFα) and interleukin (IL)8 levels in BAL fluid were measured using a commercially available ELISA (R&D systems, Abingdon, UK). BAL MPO was measured using the chromogenic substrate assay as described previously. ²⁷ Salbutamol levels were measured using a commercially available ELISA (Biox diagnostics, Jemelle, Belgium).

**Statistical analysis**

The study was powered to detect a 60% reduction in alveolar neutrophil sequestration based on recent data showing that β₂-agonists reduce pulmonary neutrophil sequestration in human volunteers exposed to LPS by 60%. ²⁸ We calculated that nine patients would need to be recruited in each arm to detect this difference with 80% power at a significance level of 0.05. Differences between groups were examined by analysis of variance or the Kruskal–Wallis test. Where significant differences were identified, Tukey’s test or the Mann–Whitney U test was used to further examine the differences. Repeated measures of neutrophil counts were non-parametric and analysed by Friedman’s repeated measures test. Linear associations were tested using Pearson’s correlation test. Data are expressed as mean (standard deviation) unless otherwise stated; p <0.05 was considered significant.

**RESULTS**

**In vitro studies**

Neutrophil migration towards the chemotactant stimulus FMLP was greater than that towards the RPMI control (11.6 mm (1.2) versus 3.6 mm (0.2), p<0.001). Salbutamol at

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<th>Table 1 Baseline demographics for patients at risk from acute respiratory distress syndrome and patients with established acute respiratory distress syndrome</th>
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<td>P:F ratio mm Hg (kPa)</td>
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<td>Lung injury score</td>
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APACHE, acute physiology and chronic health evaluation; ARDS, acute respiratory distress syndrome; SAPS, simplified acute physiology score; SOFA, sequential organ failure assessment.

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$10^{-4}$ and $10^{-5}$ M significantly reduced neutrophil chemotaxis (fig 1A). There was no effect on random neutrophil chemokinesis (fig 1B). The effect of salbutamol on neutrophil chemokinesis was abolished by propranolol (fig 1C). Salbutamol $10^{-4}$ to $10^{-6}$ M had no effect on cellular viability (data not shown) or neutrophil apoptosis (fig 1D).

Clinical studies

Patient baseline demographics
Forty patients with ALI/ARDS were initially recruited, of whom 35 had BAL performed at baseline and were included in this study. Fourteen patients with risk factors for ARDS but who had not developed ARDS at the time of BAL formed the at risk group. Some of the data from the at risk group (BAL MPO levels and neutrophil counts) have been published previously.23 Seven ventilated controls were recruited. Table 1 shows the baseline data on severity of illness and aetiology of lung injury for the patients with ARDS and those at risk.

Neutrophilic inflammation is present in the alveolar space in patients with ARDS
Evidence of increased accumulation of neutrophils and neutrophilic inflammation was found in the BAL fluid of patients with ARDS. Total neutrophil count, MPO, TNFα and IL8 levels were raised in patients with ARDS compared with patients at risk from ARDS and normal ventilated controls (fig 2 A–D). We found significant, albeit weak, linear associations between the PiCCO alveolar–capillary permeability index and MPO ($r = 0.4$, $p = 0.038$) and IL8 ($r = 0.37$, $p = 0.033$) levels. Compared with normal controls, there was evidence of increased adhesion molecule expression CD11B, CD18, CD49, reduced l-selectin expression and increased neutrophil activation (CD64) on circulating and alveolar neutrophils (fig 3).

In vivo effects of salbutamol
From the initial group of patients randomised to salbutamol or placebo, follow-up samples of blood and BAL fluid were collected from 22 patients at day 4 (9 randomised to salbutamol and 13 to placebo). Salbutamol was detected by ELISA in the plasma of the treatment group at a concentration equivalent to $10^{-6}$ M (range 250–495 ng/ml). The ELISA did not perform reproducibly in the BAL matrix.

Circulating neutrophils
Treatment with intravenous salbutamol significantly increased the number of circulating neutrophils (fig 4). There was a trend towards reduced l-selectin expression on unstimulated neutrophils in the salbutamol group (placebo 55 (19) v salbutamol 39 (30), $p = 0.18$). When neutrophils were stimulated ex vivo with FMLP, neutrophils from salbutamol-treated patients had significantly lower l-selectin expression (placebo 43 (15) v salbutamol 22 (21), 95% confidence interval (CI) 1.2 to 39, $p = 0.038$; fig 5). We found no effect on integrin adhesion molecule expression measured at day 4 (CD11B, CD18 or CD49) or expression of the neutrophil activation marker CD64 (fig 3A).
the dose used and the experimental conditions. Some studies supporting data from animal studies,27 the response was not evident until 48 h after the initiation of treatment. Our study sought to identify whether salbutamol was having an effect on pulmonary neutrophil sequestration by 30%. In normal human volunteers, in a placebo-controlled trial, treatment with 300 μg inhaled salbutamol prevented platelet-activating factor-induced pulmonary sequestration of radiolabelled neutrophils.19 Pretreatment with salmeterol similarly inhibited LPS-induced neutrophil influx, neutrophil degranulation (myeloperoxidase) and TNFα release in human volunteers.18 Reduced neutrophil–endothelial adhesion, through the down regulation of neutrophil integrin molecule expression (CD11B/18) seen with β2-agonists may partly explain this finding.16

In our study, intravenous salbutamol considerably increased circulating neutrophil count but had no apparent effect on alveolar neutrophil counts. The increase in circulating neutrophils induced by β-agonists is thought to be due to detachment of cells from the marginating neutrophil pools rather than mobilisation of neutrophils from the bone marrow.7 Neutrophil i-selectin expression may be important in this effect because of a dominant role in the initial slowing, margination and rolling behaviour of neutrophils over the endothelium at the post-capillary venule in the systemic circulation.15 The shedding of i-selectin from the neutrophil surface allows neutrophils to break free from the vascular endothelium and return to the circulation. The mobilisation of neutrophils was not, however, associated with any difference in the degree of organ dysfunction between the two groups.7

In contrast to the systemic circulation, where neutrophil sequestration usually occurs at the post-capillary venule, neutrophil migration in the lung occurs at the pulmonary capillaries, without dependence on i-selectin-mediated rolling.20 In our study, we found a trend towards reduced i-selectin expression on circulating neutrophils in patients treated with salbutamol. When the neutrophils were maximally stimulated with FMLP, neutrophils primed by treatment with salbutamol showed a notably greater reduction in i-selectin expression than those treated with placebo. This finding is consistent with the observation that isoprotrenol decreases neutrophil I-selectin expression in human volunteers.40 Enhanced I-selectin shedding would increase neutrophil mobilisation from the systemic circulation and may explain the finding of increased circulating neutrophils. By contrast, the absence of a measurable effect of salbutamol on integrin molecule expression relevant to pulmonary neutrophil emigration may explain the observed lack of effect on pulmonary neutrophil sequestration.

Figure 3 Treatment with intravenous salbutamol significantly increased circulating neutrophil count (p = 0.004). Data are shown as mean (SE) for salbutamol (squares/grey dotted line) and placebo (circles/solid line).

DISCUSSION

In a double-blind randomised placebo-controlled trial, we showed that sustained treatment with intravenous salbutamol considerably reduced extravascular lung water in humans with ALI/ARDS.19 In contrast to our initial hypothesis and supporting data from animal studies,7 the response was not evident until 48 h after the initiation of treatment. Our study sought to identify whether salbutamol was having an effect on pulmonary neutrophil accumulation and alveolar inflammation. Our study confirms previous observations that there is increased neutrophil accumulation and inflammation in the alveolar space in people with ALI/ARDS. The novel finding of a positive association between the intensity of neutrophilic infiltration and an in vivo measurement of alveolar–capillary permeability adds additional support to the hypothesis that the neutrophil is a key inflammatory mediator in ARDS. Contrary to in vitro, animal and healthy volunteer studies, we found no evidence that intravenous salbutamol modulated alveolar neutrophil accumulation, activation or markers of alveolar inflammation in humans with ALI/ARDS.

Studies investigating the in vitro effects of β-agonists on neutrophil chemotaxis have produced conflicting results, probably due to differences in the specific β2-agonist tested, the dose used and the experimental conditions. Some studies showed a reduction in neutrophil chemotaxis,28 29 whereas others reported a biphasic response with increased neutrophil chemotaxis at low concentrations of β2-agonist and a reduction in chemotaxis at higher concentrations.30 Our study reports for the first time that the physiological concentration of salbutamol achieved in the plasma after an intravenous infusion of salbutamol (15 μg/kg/h) in patients with ALI/ARDS is 10^-6 M. At this concentration, no effect on neutrophil chemotaxis was observed. Supraphysiological concentrations (10^-4 and 10^-3 M), however, reduced neutrophil chemotaxis through activation of the β-receptor. Moreover, physiologically relevant concentrations of salbutamol had no effect on neutrophil viability or the rate of spontaneous apoptosis measured by morphology and annexin V/propidium iodine staining.

The mechanisms regulating pulmonary neutrophil sequestration have been well characterised. Pulmonary neutrophil sequestration occurs within minutes of exposure to an inflammatory insult.11 12 The insult causes an increase in neutrophil stiffness and reduction in deformability,13 leading to sequestration into the pulmonary capillaries, followed by emigration into the alveolar space. The process of neutrophil emigration occurs by at least two differentially regulated pathways: CD11/18 adhesion molecule interactions determine the response to Gram-negative organisms, IL1α and phorbol 12-myristate 13-acetate, whereas Gram-positive organisms, hyperoxia and the complement anaphylatoxins (C5a) seem to induce neutrophil emigration through a CD11/18-independent pathway.14

Animal models of direct13 and indirect14 lung injury have shown that pretreatment with intravenous β2-agonists reduces pulmonary neutrophil sequestration by 30%. In normal human volunteers, in a placebo-controlled trial, treatment with 300 μg inhaled salbutamol prevented platelet-activating factor-induced pulmonary sequestration of radiolabelled neutrophils.19 Pretreatment with salmeterol similarly inhibited LPS-induced neutrophil influx, neutrophil degranulation (myeloperoxidase) and TNFα release in human volunteers.18 Reduced neutrophil–endothelial adhesion, through the down regulation of neutrophil integrin molecule expression (CD11B/18) seen with β2-agonists may partly explain this finding.16

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Effects of salbutamol on neutrophil function

There are several other potential explanations for the absence of a measurable effect of salbutamol in vivo on alveolar neutrophil accumulation and inflammation. Firstly, pulmonary neutrophil emigration occurs within minutes of exposure to the toxic insult. In the previous studies that reported a neutrophil accumulation and inflammation, the drug was administered before the inflammatory insult. In our study, salbutamol was given many hours after the initial insult, potentially therefore too late to have an effect on pulmonary neutrophil recruitment. Although the study was relatively small, it is similar in size to previous studies, and was powered to detect a treatment effect of a magnitude similar to that observed in previous studies. We cannot, however, exclude that a smaller treatment effect would have been missed.

CONCLUSION

At the onset of ALI/ARDS, there is increased pulmonary neutrophil recruitment and activation in addition to positive correlations between IL8, myeloperoxidase and alveolar-capillary permeability, suggesting an association between neutrophil activation and the development of lung injury. In vitro, physiological concentrations of salbutamol failed to show an effect on neutrophil chemotaxis, viability or apoptosis. Treating patients with ALI/ARDS with intravenous salbutamol increased the number of circulating neutrophils, but had no effect on alveolar neutrophil numbers or on neutrophil activation or alveolar inflammation. The beneficial effects of salbutamol in reducing extravascular lung water seem to be unrelated to the modulation of neutrophilic inflammatory pathways.

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Funding: This study was supported by a grant from the West Midlands Intensive Care Society, Queen Elizabeth Hospital, Birmingham, UK.

Competing interests: GDP, DFMcA and DRT have received payment to attend scientific meetings and have given talks for pharmaceutical companies that manufacture β-agonists (Astra Zeneca, GlaxoSmithKline).

Ethical approval: This study was approved by the East Birmingham Research Ethics Committee (Reference SJR/LMH/0522).

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Thorax 2007 62: 36-42 originally published online August 23, 2006
doi: 10.1136/thx.2006.059410

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