

CYSTIC FIBROSIS

Neutrophil cell death, activation and bacterial infection in cystic fibrosis

A P Watt, J Courtney, J Moore, M Ennis, J S Elborn

Thorax 2005;60:659–664. doi: 10.1136/thx.2004.038240

See end of article for authors' affiliations

Correspondence to: Dr A Watt, Department of Clinical Biochemistry and Metabolic Medicine, Institute of Clinical Science, The Queen's University of Belfast, Grosvenor Road, Belfast BT12 6BJ, UK; a.watt@qub.ac.uk

Received
26 November 2004
Accepted 9 May 2005

Background: Cystic fibrosis (CF) is characterised by chronic endobronchial bacterial infection and neutrophil mediated inflammation. Neutrophil apoptosis is essential for the resolution of inflammation. This study assessed the relationship between levels of neutrophil apoptosis and sputum microbiology in matched clinically stable patients with CF.

Methods: Sputum was induced from 34 patients (nine with no Gram negative infection, 10 colonised with *Pseudomonas aeruginosa*, 10 with *Burkholderia cenocepacia*, and five with other infections). Apoptotic neutrophils measured by flow cytometric Annexin V/propidium iodide staining and morphology were similar in all groups.

Results: Patients infected with *P aeruginosa* or *B cenocepacia* had a significantly lower percentage of viable neutrophils in the sputum than those with no Gram negative infection (Kruskal-Wallis $p=0.01$, median (interquartile range (IQR)) 14.2% (9.4–21.6), 15.8% (12.3–19.5), and 48.4% (23.0–66.4); $p=0.003$ and $p=0.002$, respectively). They also had significantly higher levels of secondary necrotic granulocytes in sputum than patients with no Gram negative infection (Kruskal-Wallis $p<0.0001$, median (IQR) 55.5% (48.4–64.5), 50.4% (44.6–61.9), and 24.8% (14.4–30.5); $p<0.0001$ and $p<0.0001$, respectively). Neutrophils ($\times 10^6/\text{g}$ sputum) in *P aeruginosa* infected patients (Kruskal-Wallis $p=0.05$, median (IQR) 6.3 (3.5–12.7)) and *B cenocepacia* infected patients (5.7 (1.5–14.5)) were significantly higher than in the group with no Gram negative infection (0.5 (0.5–4.3), $p=0.03$ and 0.04 , respectively).

Conclusion: These results suggest that cell death and clearance may be altered in patients with CF colonised with *P aeruginosa* and *B cenocepacia* compared with those with no Gram negative infection.

Lung damage due to bacterial infection remains the main cause of morbidity and mortality in cystic fibrosis (CF).¹ The lungs of patients with CF become colonised in early infancy with micro-organisms such as *Staphylococcus aureus* and *Haemophilus influenzae*, which may damage epithelial surfaces, leading to increased attachment of and eventual replacement by *Pseudomonas aeruginosa* (PA).² Once acquired, PA will invariably colonise the patient for life. The virulence and longevity of PA infections can be attributed to a number of factors including biofilm formation in the lung.¹ In recent decades, *Burkholderia cenocepacia* (BC) (formerly *B cepacia* genomovar IIIa) has also been identified as an important lung pathogen eliciting a range of disease states from asymptomatic carriage to the lethal "cepacia syndrome".³ Colonisation with both these micro-organisms is detrimental to the patient; however, pulmonary colonisation by BC is observed to lead to an accelerated decline in lung function. The mechanism by which BC causes this deterioration is unclear and, although virulence factors have been identified, none appear to account for its pathogenicity.^{4–6}

In the lungs, neutrophils are important in innate defence against infection, preventing spread of micro-organisms by phagocytosis and release of toxic agents such as pro-inflammatory cytokines, proteases, and reactive oxygen species.⁷ In a healthy lung inflammation resolves naturally and the tissue returns to normal.⁸ In CF, however, inflammation is characterised by a severe and sustained neutrophil mediated response. It has been hypothesised that this persistent inflammatory response and abnormal accumulation of cells may arise from failure or inefficiency of the mechanisms responsible for resolution of inflammation and restitution of tissue homeostasis—that is, apoptosis and subsequent clearance by macrophages.⁹ Apoptosis is a granulocyte clearance mechanism which limits tissue injury;

in contrast to necrosis, granulocyte secretory processes are shut down and the intact senescent neutrophil is removed by macrophages using phagocytic recognition mechanisms and triggering release of anti-inflammatory mediators.⁹ Secondary necrosis of apoptotic cells is likely to occur if phagocytic clearance is disrupted resulting in amplification of inflammation.⁹ The apoptotic pathway is susceptible to modulation by cytokines, implying active control of cell death during the course of inflammation in vivo.¹⁰ Evidence also exists to suggest that pathogens may have the capability to modulate apoptosis.^{11–14} PA secretes a phenazine pigment exotoxin called pyocyanin which has pro-apoptotic effects on neutrophils in culture.¹² Hemolysin, a toxin produced by BC, is also capable of inducing apoptosis and degranulation of mammalian phagocytes.¹³

It has previously been shown that, in CF patients who are clinically stable, levels of inflammatory markers are higher than in a matched non-CF population.¹⁵ There is not such a clear pattern between levels of inflammation and various different infecting organisms. In this study we have examined the relationship between neutrophil cell death, activation, inflammatory mediator levels, and sputum microbiology in matched clinically stable CF patients.

METHODS

Patients

Thirty four clinically stable patients with CF participated in the study. A diagnosis of CF was confirmed by a typical phenotype, genotype and/or sweat test. A cross sectional

Abbreviations: BC, *Burkholderia cenocepacia*; CF, cystic fibrosis; CRP, C-reactive protein; FEV₁, forced expiratory volume in 1 second; FVC, forced vital capacity; NE, neutrophil elastase; PA, *Pseudomonas aeruginosa*; PS, phosphatidylserine

Table 1 Median (interquartile range) demographic data of study patients

	PA	NGN	BC	OGN
Age (years)	24 (20–26.5)	26 (22.5–29)	25.5 (19.5–31)	33 (no IQR)
Sex (F/M)	6/4	5/4	6/4	2/3
FEV ₁ (l)	2.6 (2.0–3.2)	2.6 (2.1–2.9)	2.5 (1.1–3.4)	1.9 (no IQR)
FEV ₁ (% predicted)	70.2 (65.1–89.7)	75.0 (65.1–94.0)	67.2 (40.6–86.5)	62.6 (no IQR)
C-reactive protein (mg/ml)	3.0 (1.0–4.0)	4.0 (1.5–8.0)	9.0 (2.0–15.0)	3.0 (no IQR)
Blood neutrophils ($\times 10^9$ cells/l)	4.4 (3.2–9.7)	4.5 (3.8–5.9)	6.2 (4.3–9.8)	3.4 (no IQR)
White cell count ($\times 10^9$ cells/l)	7.3 (6.4–9.9)	8.2 (7.5–8.8)	8.9 (7.4–13.8)	6.7 (no IQR)
Total cells ($\times 10^6$ /g sputum)	4.2 (3.2–14.4)	5.2 (0.7–7.4)	3.6 (0.9–12.2)	3.1 (no IQR)

FEV₁, forced expiratory volume in 1 second; PA, *Pseudomonas aeruginosa*; NGN, no Gram negative infection; BC, *Burkholderia cenocepacia*; OGN, other Gram negative infection.

sample of CF patients was recruited matched as closely as possible for lung function, age, sex, and genetics (table 1). No power calculation was done before the study. A blood sample was also taken for measurement of C-reactive protein (CRP) concentrations, white cell count, and neutrophil count. Informed written consent was obtained from all patients. The study was approved by the ethics committee of The Queen's University of Belfast and carried out in accordance with the Declaration of Helsinki (1989). Informed written consent was obtained from all patients.

Sputum microbiology

Patients were categorised as follows into four groups based on their microbiological status over the preceding 12 months and where the patients continued to culture the same organism during the study period as in the previous 12 week period: (1) *P. aeruginosa* (PA) group where patients had been chronically colonised with PA for at least 12 months before commencement of the study; (2) *B. cenocepacia* (BC) group where patients had been chronically colonised with *B. cenocepacia* for at least 12 months before commencement of the study; (3) no Gram negative growth (NGN) where patients had no significant bacteria noted for at least 12 months before commencement of the study; and (4) other Gram negative group (OGN) where patients had been colonised with other Gram negative organisms for at least 12 months before commencement of the study (predominantly *Stenotrophomonas maltophilia*) and in the absence of PA and BC.

Sputum induction

Sputum induction was performed using the Sonix 2000 nebuliser (Clement Clarke International Ltd, Harlow, UK), adapting methods described by Pavord *et al.*^{16–17} In brief, 3% saline was nebulised for 20 minutes. Forced expiratory volume in 1 second (FEV₁) and forced vital capacity (FVC) (Vitalograph) and their reversibility to salbutamol were determined by spirometry according to ATS standards. Oxygen saturation and pulse rate were recorded throughout (Sat-Trak pulse oximeter; SensorMedics, Bilthoven, The Netherlands). Nebulisation was stopped if FEV₁ fell by >20% at any stage. If this occurred, salbutamol (200 µg) was administered as necessary. Sputum was collected in a sterile plastic pot placed on ice.

Sputum processing

Sputum samples were processed within 2 hours. The total weight of the sample was recorded. Sputum plugs were separated and processed, adapting methods described by Pavord *et al.*^{16–17} The supernatant was removed from the cell pellet and stored at –70°C for future analysis. Cell viability was determined by the trypan blue exclusion method. Cytospin slides were made and stained with Diff Quik (Clin-Tech Ltd, Essex, UK). Concentrations of interleukin (IL)-6, IL-8 and IL-10 in the supernatant were measured

using a quantitative sandwich immunoassay (Pelikine Compact ELISA kits). Neutrophil elastase (NE) was measured using a quantitative continuous (kinetic) assay.¹⁸ Sputum was cultured for identification of major pathogens using routine microbiological methods as outlined by the United Kingdom Cystic Fibrosis Trust.¹⁹ C-reactive protein (CRP) was measured by nephelometry.

Flow cytometry

Cell suspensions (1×10^6 cells/ml) were washed twice with phosphate buffered saline/1% bovine serum albumin (PBS/1% BSA) and resuspended in the binding buffer supplied with the annexin V-FITC (fluorescein isothiocyanate) (Pharmingen). Cell suspensions were then labelled with annexin V-FITC (AV) and propidium iodide (PI) or anti-human monoclonal CD66b-FITC (BD Pharmingen). Sputum cells were gated on a granulocyte population using a forward scatter and side scatter plot. Negative controls consisted of omission of AV and PI, as positive controls are not readily accessible for apoptosis detection. Mouse IgM κ isotype control was used in flow cytometric assessment of CD66b. To eliminate cellular debris from the analysis, the discrimination level was set at 100. Analysis of samples was performed using the Immuno 4 software programme which involved subtraction of negative controls from positive samples. Flow cytometry was performed as previously described.²⁰

Morphology

Cytospin slides were also prepared from sputum and evaluated by light microscopy for the presence of apoptotic cells. In order to evaluate the proportion of apoptotic cells in neutrophils recovered from sputum, at least 500 neutrophils were graded for apoptosis on each cytospin slide. Cells showing typical features of apoptosis such as nuclear chromatin condensation, nuclear coalescence, and cell shrinkage were considered apoptotic.²¹

Statistical analysis

Results are reported as median values with interquartile range. Data were analysed using a Kruskal-Wallis one-way ANOVA which, if significant, was followed by non-parametric Mann-Whitney two sample (non-matched) tests. A p value of <0.05 was considered statistically significant. When the measured mediator concentration was below the detection limit of the assay, a value equal to the detection limit of the assay was substituted in the statistical analysis. Statistical analysis was performed using SPSS Version 11.5 for Windows.

RESULTS

Thirty four patients with CF completed the study (10 with PA infection, 10 with BC infection, nine NGN, and five OGN); their demographic data are shown in table 1. There were no significant differences in lung function (% predicted p = 0.4 or litres, p = 0.8) or age (p = 0.1) between any of the groups

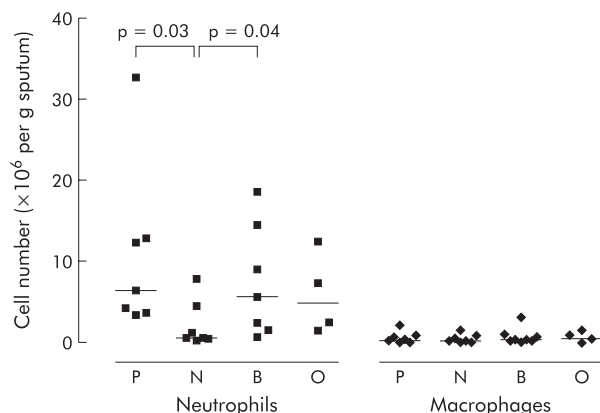


Figure 1 Neutrophil ($\times 10^6/\text{g}$ sputum) and macrophage ($\times 10^6/\text{g}$ sputum) cellular data for cystic fibrosis patients colonised with *Pseudomonas aeruginosa* (P), no Gram negative infection (N), *Burkholderia cenocepacia* (B), and other Gram negative infections (O). Data are shown as individual points; the horizontal lines represent median values. p values reaching statistical significance are shown.

studied when analysed using a Kruskal-Wallis one way ANOVA.

CRP, blood neutrophils, and white cell counts

There were no significant differences in CRP levels, blood neutrophils ($\times 10^9$ cells/l), and total white cell counts between any of the four patient groups ($p = 0.21$, $p = 0.439$ and $p = 0.213$, respectively, Kruskal-Wallis test; table 1).

Sputum cell counts

The total number of cells isolated per gram of sputum did not differ significantly between patients with CF colonised with different micro-organisms ($p = 0.8$, Kruskal-Wallis test; table 1). The number of neutrophils ($\times 10^6/\text{g}$ sputum) in the sputum of patients with PA infection (Kruskal-Wallis $p = 0.05$, median (IQR) 6.3 (3.5–12.7), $p = 0.03$) and BC infection (median (IQR) 5.7 (1.5–14.5), $p = 0.04$) was significantly higher than in the NGN group (median (IQR)

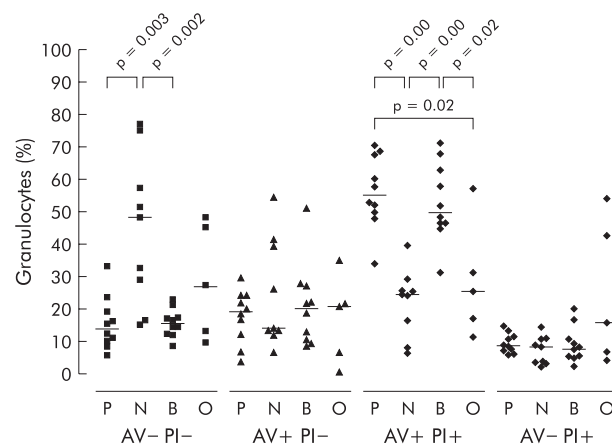


Figure 2 Flow cytometry data for sputum cells labelled with either annexin V (AV) or propidium iodide (PI) for cystic fibrosis patients colonised with *Pseudomonas aeruginosa* (P), no Gram negative infection (N), *Burkholderia cenocepacia* (B), and other Gram negative infections (O). Cells labelled with AV only (AV+/PI-) are apoptotic, cells labelled with PI only (AV-/PI+) are necrotic, cells labelled with neither AV nor PI (AV-/PI-) are neither apoptotic nor necrotic, and cells labelled with AV and PI (AV+/PI+) have secondary necrosis. Data are shown as individual points; the horizontal lines represent median values. p values reaching statistical significance are shown.

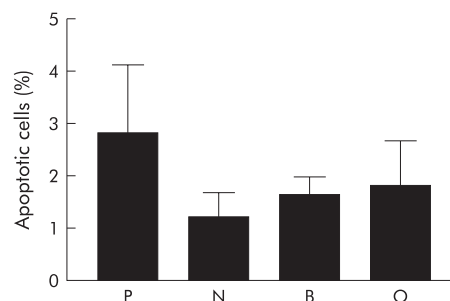


Figure 3 Percentage of apoptotic cells in sputum as assessed by morphology for cystic fibrosis patients colonised with *Pseudomonas aeruginosa* (P), no Gram negative infection (N), *Burkholderia cenocepacia* (B), and other Gram negative infections (O). Data are shown as mean (SE).

0.5 (0.5–4.3); fig 1). The number of macrophages ($\times 10^6/\text{g}$ sputum) did not differ significantly between each of the four groups (Kruskal-Wallis $p = 0.9$; fig 1). The percentage of macrophages (data not shown) was negatively correlated with the percentage of cells with secondary necrosis ($R = -0.555$, $p = 0.004$).

Apoptosis and activation

There were no significant differences in the percentage of apoptotic (AV+PI-) granulocyte numbers as measured by Annexin V staining or morphology in each of the four patient groups studied (Kruskal-Wallis $p = 0.8$ and $p = 0.4$, respectively; figs 2 and 3). As previously reported,²⁰ a lower proportion of apoptotic cells was detected by light microscopy than by flow cytometry. This is probably due to the different stages of apoptosis measured by the two methods.^{22–23} There were no significant differences in necrotic (AV-PI+) granulocyte numbers in each of the patient groups studied (Kruskal-Wallis, $p = 0.4$; fig 2). Patients infected with PA (Kruskal-Wallis $p = 0.01$, median (IQR) 14.2% (9.4–21.6)) or BC (median (IQR) 15.8% (12.3–19.5)) had a significantly lower percentage of viable (AV-PI-) granulocytes in sputum than patients in the NGN group (median (IQR) 48.4% (23.0–66.4), $p = 0.003$ and $p = 0.002$, respectively). Conversely, patients with PA (Kruskal-Wallis $p < 0.0001$, median (IQR) 55.5% (49.1–68.5)) or BC (median (IQR) 50.4% (46.0–65.8)) infection had significantly higher levels of secondary necrotic granulocytes (AV+PI+) in their sputum than patients in the NGN (median (IQR) 24.8% (12.5–27.8), $p < 0.0001$ and $p < 0.0001$, respectively) or OGN groups

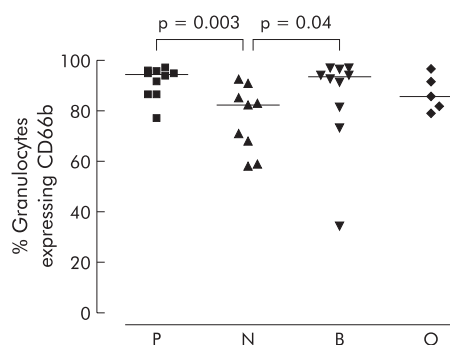


Figure 4 Flow cytometry data for sputum cells labelled with anti-CD66b for cystic fibrosis patients colonised with *Pseudomonas aeruginosa* (P), no Gram negative infection (N), *Burkholderia cenocepacia* (B), and other Gram negative infections (O). Data are shown as individual points; the horizontal lines represent median values. p values reaching statistical significance are shown.

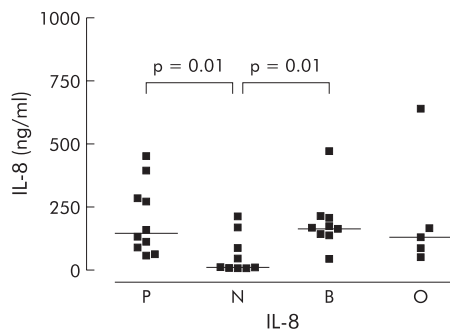


Figure 5 Concentrations of interleukin 8 (IL-8) in sputum sol from cystic fibrosis patients colonised with *Pseudomonas aeruginosa* (P), no Gram negative infection (N), *Burkholderia cenocepacia* (B), and other Gram negative infections (O). Data are shown as individual points. Horizontal lines represent the median values. p values reaching statistical significance are shown.

(median 26.0% (no IQR), $p = 0.02$ and $p = 0.02$, respectively; fig 2).

Sputum neutrophils obtained from all four patient groups were found to be highly activated as indicated by surface expression of the granulocyte activation marker CD66b (fig 4). Sputum neutrophils from patients infected with PA (Kruskal-Wallis $p = 0.04$, median (IQR) 94.2% (86.4–95.4)) or BC (median (IQR) 93.4% (77.4–96.8)) had a significantly higher percentage of granulocytes expressing CD66b than patients in the NGN group (median (IQR) 82.1% (63.3–87.9), $p = 0.003$ and $p = 0.043$, respectively).

Supernatant mediators

The concentrations of IL-8 (ng/ml) in sputum sol were significantly higher in patients infected with PA (Kruskal-Wallis, $p = 0.03$, median (IQR) 145.4 ng/ml (75.3–337.3)) and BC (median (IQR) 166.3 ng/ml (7–209.9)) than those in the NGN group (median (IQR) 13.2 ng/ml (7.3–128.4), $p = 0.01$ and $p = 0.01$; fig 5). There were no significant differences between the groups in levels of IL-6 (pg/ml) or IL-10 (pg/ml) (Kruskal-Wallis ANOVA, $p = 0.58$ and $p = 0.12$, respectively; fig 6). Three anomalous points have been omitted from the IL-6 figure to allow easier comparison of levels between groups; the omissions are as follows: P, 1927.4 ng/ml; N, 3043.09 ng/ml; O, 3366.15 ng/ml. Removal of the anomalous points did not alter the statistical findings. The concentration of NE (mg/ml) in sputum sol was significantly increased in patients infected with PA

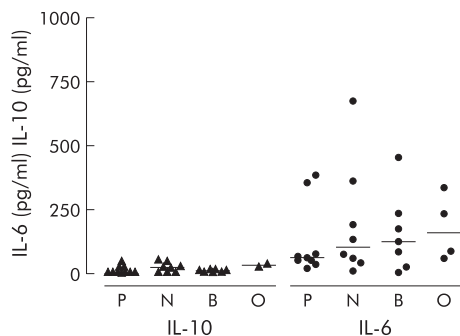


Figure 6 Concentrations of interleukins 6 and 10 (IL-6 and IL-10) in sputum sol from cystic fibrosis patients colonised with *Pseudomonas aeruginosa* (P), no Gram negative infection (N), *Burkholderia cenocepacia* (B), and other Gram negative infections (O). Data are shown as individual points. Horizontal lines represent median values. p values reaching statistical significance are shown.

(Kruskal-Wallis, $p = 0.02$, median (IQR) 35.2 mg/ml (14.4–96.0)), BC (median (IQR) 62.4 mg/ml (16.6–96.8)), or OGN (median 49.4 mg/ml, no IQR) compared with the NGN group (median (IQR) 9.6 mg/ml (3.5–29.2), $p = 0.02$, $p = 0.01$, $p = 0.03$ respectively; fig 7). Sputum sol levels of IL-8 were positively correlated with NE levels ($R = 0.60$, $p < 0.0001$).

DISCUSSION

The main aim of this study was to compare levels of sputum neutrophil apoptosis, necrosis, and activation in four groups of clinically stable CF patients matched as closely as possible for age, sex, and lung function who were colonised with micro-organisms commonly found in the CF lung. A power calculation was not performed before commencement and this is a recognised limitation of this study. We also determined levels of the inflammatory mediators IL-6, IL-8, IL-10, NE, and cell numbers in sputum, our hypothesis being that levels of apoptosis and necrosis may be modulated by the dominant micro-organism colonising the lung and hence influence the inflammatory milieu of the lung. We found significant differences in levels of cell death, cytokine production, and cell counts in sputum between patients infected with Gram negative organisms (PA and BC) and those without Gram negative infection. We did not find significant differences in these inflammatory markers between patients infected with PA and those infected with BC, which suggests that the inflammatory reaction provoked by these two micro-organisms is similar.

This is the first study to compare levels of neutrophil cell death and inflammatory markers in sputum samples from clinically stable patients with CF colonised with different lung pathogens. Levels of necrotic (AV+PI+) neutrophils were very similar between the groups studied, with no significant variations observed. Patients infected with PA or BC had a significantly lower number of viable (AV–PI–) granulocytes in sputum than those with no Gram negative infection. When neutrophils are recruited to a site of infection, their life span is extended by the actions of pro-inflammatory cytokines and death is then induced as a consequence of phagocytosis of microbes and activation of microbicidal mechanisms.²⁴ This induction of death as a result of microbe ingestion provides a probable explanation for the lower numbers of viable neutrophils observed in the infected groups. Induction of neutrophil death following phagocytosis may be limited in patients infected with organisms such as PA, which can successfully resist neutrophil phagocytosis by switching from a non-mucoid variant to a mucoid phenotype by forming large exopolysaccharide coated microcolonies after onset of

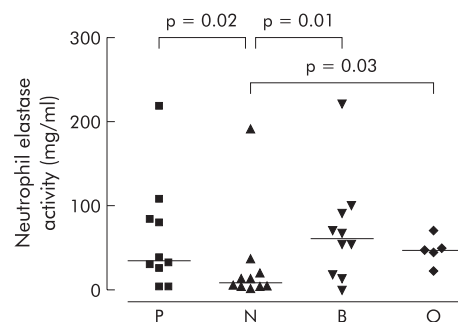


Figure 7 Concentrations of neutrophil elastase in sputum sol from cystic fibrosis patients colonised with *Pseudomonas aeruginosa* (P), no Gram negative infection (N), *Burkholderia cenocepacia* (B), and other Gram negative infections (O). Data are shown as individual points. Horizontal lines represent the median values. p values reaching statistical significance are shown.

infection.²⁵ This biofilm formation then results in frustrated neutrophil phagocytosis.²⁵

The mean level of apoptosis (AV+PI-) observed in the sputum of patients with CF was around 20% when analysed by annexin V staining. This is in agreement with levels observed in CF sputum by Vandivier *et al.*²⁶ There were no significant differences in levels of apoptotic granulocytes between the groups studied. A trend for higher median levels of apoptosis was observed in the PA and BC groups than in the NGN group when apoptosis was assessed by flow cytometry and morphology, although this did not reach statistical significance.

The percentage of secondary necrotic granulocytes was significantly higher in patients colonised with PA and BC than in the NGN group. Secondary necrosis of apoptotic cells is likely to occur if phagocytic clearance is rendered ineffective. Phagocytic clearance may be ineffective for a number of reasons including a lack of mature inflammatory macrophages, overwhelming of the clearance system by massive waves of apoptosis, or failure of phagocytic recognition systems.⁹ If clearance is slower than rates of transition of apoptotic to secondary necrotic, the increase in rates of apoptosis might be translating into the significant increase observed in secondary necrosis. Secondary necrosis of neutrophils exacerbates the inflammatory response and results in further tissue damage. BC and PA have both been shown to induce neutrophil apoptosis through the production of the toxins hemolysin and pyocyanin, respectively.¹²⁻¹³ Induction of non-inflammatory apoptosis may prove advantageous to the infecting microbe; however, extensive tissue damage exists in patients infected with PA or BC, which suggests that excessive apoptosis overwhelms phagocytic clearance resulting in secondary necrosis. Levels of pyocyanin or hemolysin were not measured in this study.

Apoptosis is an intrinsic process, however; it can also be modulated by cytokines such as IL-8 which has been shown to delay spontaneous apoptosis of neutrophils in a dose dependent manner.²⁷ Levels of IL-8 were significantly higher in patients infected with PA and BC than in NGN patients, although this was not coupled with significantly lower levels of apoptosis in these patients. However, IL-8 also acts as a major neutrophil chemoattractant and, indeed, increased neutrophil numbers were observed in the PA and BC patient groups. The number of neutrophils ($\times 10^6$ /g sputum) was significantly higher in the PA and BC groups than in the NGN group. However, the number of macrophages ($\times 10^6$ /g sputum) was not significantly different between the four groups studied. An increase in neutrophil numbers without an accompanying increase in macrophage numbers may have an adverse effect on the efficiency of clearance of apoptotic cells. This may contribute to the significantly higher levels of secondary necrotic cells observed in the PA and BC groups. The total number of cells/g sputum isolated did not differ significantly between any of the CF groups recruited. It is interesting to note that, even though patients with no Gram negative infection remain relatively well, neutrophil infiltration of the lung is still observed at levels comparable with patients infected with major lung pathogens.

As previously mentioned, clearance is a critical feature of the apoptotic process, providing deletion of large numbers of neutrophils while preserving the structure and function of surrounding tissue.²⁸ Exposure of phosphatidylserine (PS) on the outer leaflet of the plasma membrane is a well recognised surface change which occurs early in the apoptotic process.²⁹⁻³⁰ PS expression coupled with the dominant role of the PS receptor suggests that interaction between the two plays a crucial role in uptake of apoptotic cells.²⁶⁻³¹ Cleavage of the PS receptor by NE specifically disrupts phagocytosis of apoptotic cells.²⁶ Our data show significantly increased levels of NE in

the sputum sol of patients infected with PA and BC, although we found no correlation with the percentage of secondary necrotic cells present in the sputum. Decreased neutrophil clearance has also been observed as a consequence of the induction of macrophage apoptosis in the presence of tumour necrosis factor α and increased levels of sodium chloride, resulting in conditions of hyperosmolarity.³² This may partially explain the persistence of inflammation in CF through impairment of macrophage clearance of neutrophils.

CD66b is a membrane bound glycoprotein involved in neutrophil migration and activation; it is present in the intracellular membrane of different subsets of cytoplasmic granules which, during cellular activation, become mobilised resulting in increased expression on the cell surface.³³ Kinhult *et al.*³⁴ have demonstrated a marked increase in the expression of CD66b on the cell surface of neutrophils following their migration from the bloodstream to the nasal mucosa. We used CD66b as a marker to assess neutrophil activation in each of the study groups. All of the CF patients studied had high levels of CD66b expression as assessed by flow cytometry. Neutrophils from patients infected with PA or BC expressed significantly higher levels of CD66b, probably as a direct result of the presence of bacterial products in the lung.

Activated neutrophils release large quantities of NE which directly damages the airways through the digestion of elastin and other structural proteins in addition to cleaving opsonins and receptors necessary for phagocytosis.³⁵ As a result, opsonophagocytosis and killing of PA and other CF related pathogens such as *S aureus* and *H influenzae* has been found to be markedly impaired.²⁵ NE also stimulates the release of pro-inflammatory cytokines such as IL-8 and IL-6, increased levels of which are seen in the CF lung.³⁶⁻³⁸ Once again the significantly higher levels of IL-8 and NE observed in the PA and BC groups compared with the NGN group may be related to higher levels of neutrophil activation in these groups, or the higher levels of cytokines and proteases observed may result from secondary necrosis. In contrast to this, the levels of IL-10 measured in all the groups were very low, and this is hypothesised to contribute to the development of chronic airways inflammation.³⁹

PA and BC infection produced very similar results in terms of levels of cell death and inflammatory mediator production during clinical stability. This study does not provide any obvious explanation for the rapid deterioration seen in patients with BC. How similar these levels would remain during an infective exacerbation remains to be investigated. The role of the epithelium in driving this deterioration may be important. It has been shown that extracellular products of BC stimulate the respiratory epithelium to produce IL-6, IL-8 and prostaglandin E₂, suggesting that the interaction between the respiratory epithelium and secreted bacterial products is important.⁴⁰

The results of this study suggest that cell death pathways in patients infected with PA and BC may be impaired. Whether this occurs as a result of altered apoptotic rate or impaired macrophage clearance requires further investigation. Patients with no Gram negative infection were shown to have high levels of neutrophil infiltration, neutrophil activation, and inflammatory mediators in their induced sputum, indicating that, even in the absence of major lung pathogens, these CF patients have an exaggerated inflammatory response. It remains controversial whether excessive lung inflammation is an intrinsic property of the CFTR defect or whether it is secondary to the unique environment of the CF lung.⁴¹

A vicious cycle of infection and inflammation occurs in CF which makes it difficult to assess the dynamic sequence of events which is cell death and clearance, leaving a number of

questions unanswered. Which cells cause more damage to the lung—activated viable cells or necrotic and secondary necrotic cells? Where does the equilibrium between neutrophil apoptosis and macrophage clearance become dysregulated? The use of induced sputum gives the advantage of providing physiologically true samples of the inflammatory milieu of the CF lung; however, it is very difficult to measure the dynamics of cell clearance within these samples. It is therefore problematic to assess whether apoptosis is inhibited or accelerated in patients infected with PA and BC compared with NGN patients or whether phagocytic clearance mechanisms have been impaired.

Inflammation in the CF lung—like other chronic lung diseases—involves a number of pro-inflammatory pathways and an apparent dampening of the anti-inflammatory pathway. In the CF lung, inflammatory markers are apparent even in the absence of major pathogens such as PA and BC. In the presence of infection this response is heightened further, as shown by increased neutrophil activation and higher levels of inflammatory mediators, which suggests that CF neutrophils may have limited capability in restricting their response to infection. These results suggest that cell death and clearance may be altered in patients with CF who are colonised with PA and BC compared with those with no Gram negative infection.

ACKNOWLEDGEMENTS

The authors thank Dr L Martin, School of Pharmacy, The Queen's University of Belfast, for supplying the protease inhibitor Z-Phenylphosphonate and Ms K Moffit, School of Pharmacy, The Queen's University of Belfast for measurement of neutrophil elastase.

Authors' affiliations

A P Watt, J Courtney, M Ennis, J S Elborn, Respiratory Research Group, The Queen's University of Belfast, Belfast, UK
J Moore, Department of Bacteriology, Belfast City Hospital, Belfast, UK

This study was funded by an Ireland-Northern Ireland Co-operation Research Project Grant from the Research & Development Office, Northern Ireland Health & Social Services Central Services Agency.

There are no competing interests to declare.

REFERENCES

- Tomlin KL, Coll OP, Ceri H. Interspecies biofilms of *Pseudomonas aeruginosa* and *Burkholderia cepacia*. *Can J Microbiol* 2001;**47**:949–54.
- Lyczak JB, Cannon CL, Pier GB. Lung infections associated with cystic fibrosis. *Clin Microbiol Rev* 2002;**15**:194–222.
- Frangolias DD, Mahenthiralingam E, Rae S, et al. *Burkholderia cepacia* in cystic fibrosis. Variable disease course. *Am J Respir Crit Care Med* 1999;**160**:1572–7.
- Nelson JW, Butler SL, Krieg D, et al. Virulence factors of *Burkholderia cepacia*. *FEMS Immunol Med Microbiol* 1994;**8**:89–97.
- Straus DC, Lonon MK, Woods DE, et al. Production of an extracellular toxic complex by various strains of *Pseudomonas cepacia*. *J Med Microbiol* 1989;**30**:17–22.
- Stover GB, Drake DR, Montie TC. Virulence of different *Pseudomonas* species in a burned mouse model: tissue colonization by *Pseudomonas cepacia*. *Infect Immun* 1983;**41**:1099–104.
- Hammerle AF, Tatschl C, Jatzko S, et al. The lung as an inflammatory organ. *Acta Anaesthesiol Scand Suppl* 1997;**111**:59–64.
- Henson PM. Possible roles for apoptosis and apoptotic cell recognition in inflammation and fibrosis. *Am J Respir Cell Mol Biol* 2003;**29**(3 Suppl):S70–6.
- Haslett C. Granulocyte apoptosis and its role in the resolution and control of lung inflammation. *Am J Respir Crit Care Med* 1999;**160**:S5–11.
- Lee A, Whyte MK, Haslett C. Inhibition of apoptosis and prolongation of neutrophil functional longevity by inflammatory mediators. *J Leukoc Biol* 1993;**54**:283–8.
- Zychlinsky A, Sansonetti PJ. Apoptosis as a proinflammatory event: what can we learn from bacteria-induced cell death? *Trends Microbiol* 1997;**5**:201–4.
- Usher LR, Lawson RA, Geary I, et al. Induction of neutrophil apoptosis by the *Pseudomonas aeruginosa* exotoxin pyocyanin: a potential mechanism of persistent infection. *J Immunol* 2002;**168**:1861–8.
- Hutchison ML, Poxton IR, Govan JR. *Burkholderia cepacia* produces a hemolysin that is capable of inducing apoptosis and degranulation of mammalian phagocytes. *Infect Immun* 1998;**66**:2033–9.
- Dacheux D, Toussaint B, Richard M, et al. *Pseudomonas aeruginosa* cystic fibrosis isolates induce rapid, type III secretion-dependent, but ExoU-independent, oncosis of macrophages and polymorphonuclear neutrophils. *Infect Immun* 2000;**68**:2916–24.
- Hendry J, Elborn JS, Nixon L, et al. Cystic fibrosis: inflammatory response to infection with *Burkholderia cepacia* and *Pseudomonas aeruginosa*. *Eur Respir J* 1999;**14**:435–8.
- Pavord ID, Pizzichini MM, Pizzichini E, et al. The use of induced sputum to investigate airway inflammation. *Thorax* 1997;**52**:498–501.
- Kelly MG, Brown V, Martin SL, et al. Comparison of sputum induction using high-output and low-output ultrasonic nebulizers in normal subjects and patients with COPD. *Chest* 2002;**122**:955–9.
- Castillo MJ, Nakajima K, Zimmerman M, et al. Sensitive substrates for human leukocyte and porcine pancreatic elastase: a study of the merits of various chromophoric and fluorogenic leaving groups in assays for serine proteases. *Anal Biochem* 1979;**99**:53–64.
- United Kingdom Cystic Fibrosis Trust. Antibiotic treatment for cystic fibrosis. Bromley, Kent, 2000.
- Watt AP, Brown V, Courtney J, et al. Neutrophil apoptosis, proinflammatory mediators and cell counts in bronchiectasis. *Thorax* 2004;**59**:231–6.
- Zhang X, Moilanen E, Kankaanranta H, Bedomethason, budesonide and fluticasone propionate inhibit human neutrophil apoptosis. *Eur J Pharmacol* 2001;**431**:365–71.
- Chilvers ER, Cadwallader KA, Reed BJ, et al. The function and fate of neutrophils at the inflamed site: prospects for therapeutic intervention. *J R Coll Physicians Lond* 2000;**34**:68–74.
- Willingham MC. Cytochemical methods for the detection of apoptosis. *J Histochem Cytol* 1999;**47**:1101–9.
- Steensma DP, Timm M, Witzig TE. Flow cytometric methods for detection and quantification of apoptosis. *Methods Mol Med* 2003;**85**:323–32.
- Doring G. Cystic fibrosis respiratory infections: interactions between bacteria and host defence. *Monaldi Arch Chest Dis* 1997;**52**:363–6.
- Vandivier RW, Fadok VA, Hoffmann PR, et al. Elastase-mediated phosphatidylserine receptor cleavage impairs apoptotic cell clearance in cystic fibrosis and bronchiectasis. *J Clin Invest* 2002;**109**:661–70.
- Ketritz R, Gaido ML, Haller H, et al. Interleukin-8 delays spontaneous and tumor necrosis factor- α -mediated apoptosis of human neutrophils. *Kidney Int* 1998;**53**:84–91.
- Fadok VA. Clearance: the last and often forgotten stage of apoptosis. *J Mammary Gland Biol Neoplasia* 1999;**4**:203–11.
- Martin SJ, Reutelingsperger CP, McGohon AJ, et al. Early redistribution of plasma membrane phosphatidylserine is a general feature of apoptosis regardless of the initiating stimulus: inhibition by overexpression of Bcl-2 and Abl. *J Exp Med* 1995;**182**:1545–56.
- Verhoven B, Schlegel RA, Williamson P. Mechanisms of phosphatidylserine exposure, a phagocyte recognition signal, on apoptotic T lymphocytes. *J Exp Med* 1995;**182**:1597–601.
- Fadok VA, Bratton DL, Rose DM, et al. A receptor for phosphatidylserine-specific clearance of apoptotic cells. *Nature* 2000;**405**:85–90.
- Kerby GS, Cottin V, Accurso FJ, et al. Impairment of macrophage survival by NaCl: implications for early pulmonary inflammation in cystic fibrosis. *Am J Physiol Lung Cell Mol Physiol* 2002;**283**:L188–97.
- Skubitz KM, Campbell KD, Skubitz AP. CD66a, CD66b, CD66c, and CD66d each independently stimulate neutrophils. *Leukoc Biol* 1996;**60**:106–17.
- Kinhult J, Egesten A, Benson M, et al. Increased expression of surface activation markers on neutrophils following migration into the nasal lumen. *Clin Exp Allergy* 2003;**33**:1141–6.
- Konstan MW, Berger M. Current understanding of the inflammatory process in cystic fibrosis: onset and etiology. *Pediatr Pulmonol* 1997;**24**:137–42.
- Bonfield TL, Panuska JR, Konstan MW, et al. Inflammatory cytokines in cystic fibrosis lungs. *Am J Respir Crit Care Med* 1995;**152**:2111–8.
- Bonfield TL, Konstan MW, Berger M. Altered respiratory epithelial cell cytokine production in cystic fibrosis. *J Allergy Clin Immunol* 1999;**104**:72–8.
- Pukhalsky AL, Kapranov NI, Kalashnikova EA, et al. Inflammatory markers in cystic fibrosis patients with lung *Pseudomonas aeruginosa* infection. *Mediators Inflamm* 1999;**8**:159–67.
- Takanashi S, Hasegawa Y, Kanehira Y, et al. Interleukin-10 level in sputum is reduced in bronchial asthma, COPD and in smokers. *Eur Respir J* 1999;**14**:309–14.
- Fink J, Steer JH, Joyce DA, et al. Pro-inflammatory effects of *Burkholderia cepacia* on cystic fibrosis respiratory epithelium. *FEMS Immunol Med Microbiol* 2003;**38**:273–82.
- Becker MN, Sauer MS, Muhlebach MS, et al. Cytokine secretion by cystic fibrosis airway epithelial cells. *Am J Respir Crit Care Med* 2004;**169**:645–53.