Arterialised earlobe blood gas analysis: an underused technique

A D Pitkin, C M Roberts, J A Wedzicha

Abstract

**Background** – Techniques for sampling arterialized capillary blood from the finger pulp and the earlobe were first described over two decades ago but, although close agreement between arterial values and earlobe samples has been demonstrated in normal subjects, this technique is not in common usage.

**Methods** – Forty patients with chronic lung disease and a wide range of arterial blood gas values were studied. Simultaneous earlobe and arterial samples were drawn with the patient at rest and analysed in the same blood gas analyser. The respiratory function laboratory staff in 50 UK hospitals with a respiratory department were telephoned and asked whether the technique was used in their hospital and the reasons, if known, for not adopting it.

**Results** – Earlobe and arterial blood gas tensions agreed closely over a wide range of values of arterial pH, Pco2 (mean difference 0.21, 95% confidence intervals −0.24 to +0.67 kPa) and Po2 (mean difference −0.17, 95% confidence intervals −1.09 to +0.75 kPa), especially at arterial Po2 values lower than 8 kPa. Of 50 UK centres surveyed 18% used the arterialized earlobe technique and 4% had plans to introduce it. Reasons for not using it were lack of knowledge in 64%, no blood gas analyser in 6%, the technique was considered inaccurate in 4%, and insufficient staff in 4%.

**Conclusions** – Although earlobe blood gas analysis is sufficiently accurate to be reliably substituted for arterial sampling in routine clinical practice, most centres in the UK do not use the technique. The main reasons for this appear to be lack of knowledge of its existence and uncertainty over its accuracy.

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Measurement of arterial blood gas tensions is a routine part of the assessment of patients with acute and chronic respiratory disorders producing abnormalities of gas exchange. Blood sampling by direct arterial puncture is the accepted technique established in clinical practice. This method, however, requires qualified medical staff to perform it and may result in significant discomfort and morbidity for the patient. Alternative methods such as cutaneous pulse oximetry have also been used to estimate arterial oxygen tension but correlate poorly with arterial Po2 values on the steep phase of the oxyhaemoglobin dissociation curve. Transcutaneous carbon dioxide measurement is an even less accurate technique and it is relatively cumbersome and time consuming to obtain single estimations.

Techniques for sampling of arterialized capillary blood from the finger pulp and the earlobe were first described over two decades ago. Although close agreement with arterial values and earlobe samples has been shown, the technique is not in common usage. These studies used small numbers of normal volunteers, undergoing physiological exercise testing, who had a relatively limited range of arterial blood gas values. Patients with chronic lung disease have a much wider range of arterial blood gas tensions, and close agreement between arterial and earlobe blood gas values in this group has not yet been demonstrated.

We therefore undertook a study to assess the accuracy of this technique in measuring arterial blood gas tensions in patients with chronic respiratory disorders and with a wide range of values of arterial Po2 and Pco2. We also conducted a survey of other hospitals to find out how commonly the technique is used elsewhere, and the reasons, if any, for not adopting this method.

**Methods**

Forty patients with chronic respiratory disease were studied. Their diagnoses were chronic obstructive pulmonary disease and bronchiectasis (29 patients), restrictive disorders including kyphoscoliosis, ankylosing spondylitis, thoracoplasty (eight patients), sarcoidosis (two patients), and one patient being investigated for breathlessness with no identifiable cause. Median forced expiratory volume in one second (FEV1) was 0.77 (range 0.38–2.7) l and median forced vital capacity (FVC) was 1.7 (range 0.89–4.3) l.

Each patient had simultaneous capillary and arterial samples taken at rest which were measured by the same analyser (Radiometer ABL3, Copenhagen, Denmark) with an interval of no more than five minutes between samples. Earlobe samples were taken by one of four technicians experienced in the technique. The method of capillary sampling was as previously described and involves spreading an earlobe (usually the patient's left, with a right handed operator) with nicotinate paste (Algipan or similar) for at least 10 minutes to induce capillary vasodilatation. A stab incision is made in the inferolateral aspect of the pinna from which blood usually flows freely (this is
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Comparative data for pH, P0₂, and Pco₂ values of simultaneous arterial and capillary blood samples in 40 patients

<table>
<thead>
<tr>
<th></th>
<th>Range</th>
<th>Maximum difference</th>
<th>Mean difference</th>
<th>95% limits of agreement</th>
</tr>
</thead>
<tbody>
<tr>
<td>pH</td>
<td>7.32–7.50</td>
<td>0.034</td>
<td>+0.007</td>
<td>–0.008 to +0.022</td>
</tr>
<tr>
<td>Pco₂ (kPa)</td>
<td>3.87–10.03</td>
<td>0.78</td>
<td>+0.21</td>
<td>–0.24 to +0.67</td>
</tr>
<tr>
<td>P0₂ (kPa)</td>
<td>4.42–12.97</td>
<td>–1.6</td>
<td>–0.17</td>
<td>–1.09 to +0.75</td>
</tr>
</tbody>
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Arterialised blood (important) but may require a small amount of manual massage. The arterialised blood collects in a drop on the inferior aspect of the earlobe. It is drawn into a thin glass capillary tube by surface tension under the control of a gloved finger over the open end of the tube and then aspirated into the analyser.

Arterial samples were collected directly into a preheparinised 3 ml plastic syringe by direct puncture of the radial artery.

Respiratory function laboratories in 50 hospitals around the UK were contacted by telephone. They consisted primarily of teaching hospitals and centres with an interest in respiratory medicine. The senior laboratory technician was asked how routine blood gas measurements were performed in that hospital. If the earlobe blood gas technique was known they were asked the reasons, if any, for not using it.

Simple descriptive statistics for the pH, P0₂, and Pco₂ values obtained with each technique were calculated, consisting of mean differences and 95% confidence intervals for arterial and earlobe blood gas tensions. The data are shown graphically using simple scatter plots of earlobe P0₂, arterial P0₂, and earlobe Pco₂ vs arterial Pco₂. They were also plotted as the difference between arterial and earlobe values vs the mean value as recommended by Bland and Altman.

Results

The range, mean, standard deviation, and 95% limits of agreement for pH and blood gas values obtained by simultaneous sampling of earlobe and arterial blood in 40 patients are shown in the table. The concordance between arterial and earlobe blood gas tensions was good throughout a wide range of values of arterial P0₂ (fig 1) and Pco₂ (fig 2). A particularly good correlation between samples was observed at arterial P0₂ values lower than 8 kPa. Above this level earlobe P0₂ values tended to be slightly lower than arterial P0₂ values. Nearly all values, however, lie within 0.5 kPa or less and underestimation of true arterial P0₂ at higher levels of oxygenation is unlikely to be of clinical significance. The correlation between Pco₂ measurements derived from the two methods is even better with the earlobe technique, tending towards slightly higher values. The mean difference between samples was, however, only 0.21 kPa.

Fifty hospitals were surveyed by telephone. Of these, nine (18%) used the arterialised earlobe technique and two (4%) had plans to introduce it. In 32 (64%) of the 59 remaining hospitals the main reason for not using earlobe blood gases was that the laboratory staff were unaware of the technique. In three hospitals (6%) a blood gas analyser was not available in the laboratory. In two centres (4%) the technique was thought to be inaccurate, and in a further two (4%) there were insufficient technical staff to carry out the procedure.

Discussion

The close agreement between arterial and earlobe blood gas values found in our study confirms the findings of earlier work in normal subjects where earlobe blood gas values have been found to be sufficiently accurate for use in exercise testing to calculate cardiac output, venous admixture, and dead space. Our findings indicate that this accuracy extends throughout the much wider ranges of arterial P0₂ and Pco₂ found in patients with respiratory disease, and suggest that earlobe blood gas analysis can reliably be extended from a research technique into routine clinical practice.

Earlobe blood gas measurement could have a valuable role in the assessment of patients for long term oxygen therapy in accordance with published guidelines. Although pulse oximetry is useful for screening, it is not sufficiently precise to be substituted for direct arterial P0₂ measurement. Earlobe blood gas analysis, however, is particularly accurate at arterial P0₂ values less than 8 kPa, can determine arterial Pco₂, and is painless enough to allow several samples to be taken on a single occasion with the patient breathing oxygen at varying flow rates to ensure adequate correc-
tion of their hypoxaemia. Thus, provision of this technique on an outpatient basis would facilitate appropriate prescription of long term oxygen therapy (LTOT) by general practitioners. Although this may necessitate additional technician time in the lung function laboratory, it would eliminate the need for qualified medical staff to perform arterial puncture. Additional savings might be made by reducing inappropriate prescription of LTOT. At this hospital over 3000 earlobe blood gas measurements are performed each year in inpatients and outpatients. Many of them spontaneously indicate their preference for earlobe sampling.

Collection of arterialized earlobe samples does not occur within a closed system and therefore the technique poses a theoretical risk of infection to the operator from exposure to the patient’s blood. The amount involved is small (about 1–2 ml), but appropriate measures to eliminate any possible hazard of infection should be used. In our unit the operator wears gloves and an absorbent disposable towel is placed underneath the earlobe. The tiny incision bleeds for a few seconds only and is then dressed with a small waterproof plaster. In contrast, arterial puncture exposes the operator to the risk of a needlestick injury and, furthermore, inoculation of blood is more likely with the low friction syringes often used for blood gas analysis. Spillage of the contents of the syringe may also occur during injection of the sample into some analysers; this is less likely to occur during aspiration of an earlobe sample from its glass capillary tube.

Our survey included most of the teaching hospitals and specialist centres for respiratory medicine within the UK, and this sample may be biased towards units more likely to employ the technique. The proportion of all UK hospitals using earlobe blood gas analysis is unlikely to be higher than the 18% found in our survey, which also suggests that the main reason for not using it is lack of knowledge of the technique’s existence or its accuracy. Other reasons identified in our survey for not using earlobe blood gas analysis, such as availability of staff and blood gas analysers, are logistical and unique to each centre. Concern that delayed analysis of the sample renders the result inaccurate applies more to conventional plastic syringes than to glass capillary tubes which have been shown to preserve blood gas tensions better than glass or plastic syringes at room temperature.

In summary, based on the results of this study we conclude that earlobe blood gas analysis is a valuable technique in the management of patients with all forms of respiratory disease producing abnormalities of gas exchange, and that only a minority of centres use it. The principal reasons for its underuse are a lack of awareness of its existence and concern over its accuracy, neither of which should preclude its adoption in many more hospitals in the UK.

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