

Penetration of amoxicillin/clavulanic acid into bronchial mucosa with different dosing regimens

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Abstract

Background – The efficacy of an antibiotic is related to its concentration at the site of infection. Previous studies of the concentrations of amoxicillin and clavulanic acid (co-amoxiclav) in respiratory secretions or whole lung tissue have suffered from methodological problems. The concentration of amoxicillin and clavulanic acid was determined in bronchial mucosal biopsy samples obtained at bronchoscopy following five different dosing regimens.

Methods – Bronchial biopsy and serum samples were obtained from 50 patients undergoing diagnostic bronchoscopy. Ten patients each received 375 mg, 625 mg, 750 mg, and 3.25 g oral, and 1.2 g intravenous co-amoxiclav 1–3 hours before bronchoscopy. The concentrations of clavulanic acid and amoxicillin were determined by high performance liquid chromatography using a microbore column, solid phase extraction, and pre-concentration to improve sensitivity tenfold over previous methods.

Results – Concentrations of both clavulanic acid and amoxicillin in bronchial mucosa were dose related and were well above the MIC₉₀ of co-amoxiclav for the common bacterial respiratory pathogens including *Haemophilus influenzae*, *Micrococcus catarrhalis* and *Streptococcus pneumoniae* for all dosing regimens. Mean mucosal levels were 200% and 118% of the corresponding serum levels for amoxicillin and clavulanic acid respectively.

Conclusions – Amoxicillin and clavulanic acid are concentrated in bronchial mucosa and, even at the lowest dose of 375 mg orally, are likely to produce tissue levels in the lung sufficient to inhibit all the common community acquired respiratory pathogens.

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Several studies have attempted to assess the concentrations of amoxicillin and clavulanic acid in respiratory secretions¹⁻⁵ and whole lung tissue.⁶ Antibiotic concentrations in sputum and bronchial mucus vary greatly because of difficulty in sampling, drug instability, and contamination by blood and saliva.⁷⁻⁸ Bronchial washings also suffer from the same problems and are further compounded by the dilutional

effect of lavage. Lung tissue obtained at thoracotomy is non-homogenous and so represents an average of concentrations found in different tissues. The concentration of antibiotic in bronchial mucosa provides a reliable guide to bronchial penetration^{9,10} and may be a better predictor of clinical efficacy than the serum level¹¹ for treatment of bronchitis and bronchopneumonia. Although one study has investigated the concentration of amoxicillin in bronchial mucosa,⁹ there have only been provisional reports on the bronchial mucosal levels of clavulanic acid.^{12,13} We have assessed levels of amoxicillin and clavulanic acid in the bronchial mucosa following the administration of co-amoxiclav by five different dosing regimens in patients undergoing diagnostic bronchoscopy.

Methods

The study was approved by the hospital ethical committee and informed consent was obtained. Patients were excluded if they were pregnant, suffering from severe cardiovascular or renal disease, hypersensitive to β-lactams, undergoing bronchoscopy for diagnosis of infection, or had received systemic antibacterial chemotherapy in the previous three days. Fifty patients, 10 for each dosing regimen, undergoing diagnostic bronchoscopy were enrolled. No patient had clinical signs of bronchial infection during the study. Patients received 375 mg, 625 mg, 750 mg, or 3.25 g oral co-amoxiclav or 1.2 g intravenously 1–3 hours before bronchoscopy.

Fibreoptic bronchoscopy was performed under general anaesthetic and serum samples were obtained prior to mucosal biopsy. Two biopsy specimens were taken from secondary carinae in both lungs. Grossly blood contaminated biopsies were discarded. Specimens were transported to the laboratory on ice, weighed and buffered to pH 6 in 200 μl NaH₂PO₄, and stored at –70°C within 90 minutes of collection. Weighing the biopsy samples before storage avoided any risk of artificial concentration of the drug in the event of desiccation of the sample. Before assay mucosal samples were homogenised by ultrasonication on ice to prevent overheating during processing. A tapered microtip attached to the horn of an ultrasonicator (Branson Sonic Power, Danbury, Connecticut, USA) was used and each biopsy sample was subjected to 20–30 seconds of ultrasonication to give a uniform suspension ready for high performance liquid

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Table 1 Mean (SD) and coefficient of variation (CV) in serum and bronchial mucosal levels of amoxicillin following administration of co-amoxiclav

Dose of amoxicillin (co-amoxiclav)	Serum levels (mg/l)			Bronchial mucosa levels (mg/kg)		
	Mean	(SD)	CV	Mean	(SD)	CV
250 mg (375 mg)	2.6	(1.37)	0.53	4.1	(1.58)	0.38
500 mg (625 mg)	5.6	(1.87)	0.33	10.1	(1.75)	0.17
500 mg (750 mg)	5.1	(1.59)	0.31	7.2	(2.58)	0.35
1 g (1.2 g)	8.4	(2.34)	0.27	20.8	(5.71)	0.27
3 g (3.25 g)	10.9	(2.43)	0.22	23.1	(5.1)	0.22

Table 2 Mean (SD) and coefficient of variation (CV) in serum and bronchial mucosal levels of clavulanic acid

Dose of clavulanic acid (co-amoxiclav)	Serum levels (mg/l)			Bronchial mucosa levels (mg/kg)		
	Mean	(SD)	CV	Mean	(SD)	CV
125 mg (375 mg)	1.4	(0.82)	0.58	1.9	(0.72)	0.37
125 mg (625 mg)	2.3	(0.85)	0.37	2.4	(0.94)	0.38
250 mg (750 mg)	2.3	(1.15)	0.50	1.6	(0.47)	0.30
200 mg (1.2 g)	2.7	(0.77)	0.28	3.6	(1.35)	0.37
250 mg (3.25 g)	2.9	(1.4)	0.48	4.1	(1.65)	0.40

chromatography with a microbore column, solid phase extraction, and preconcentration.¹⁴ Spiked standards were prepared in water from pure powder. Fresh antibiotic free biopsy tissue was ultrasonicated with buffer to produce a control biopsy suspension. Standards and biopsy suspension or serum were mixed 1:1 to give a range of known biopsy standards of 500, 250, 125, 64, 32, 16 and 8 µg/kg for clavulanic acid and 1000, 500, 250, 125, 64 and 32 µg/kg for amoxicillin. Blank standards were included. Serum standards were 4, 2, 1, 0.5 and 0.25 mg/l for clavulanic acid and 8, 4, 2, 1 and 0.5 mg/l for amoxicillin. The biopsy and serum standards were treated in the same way as the biopsy and serum samples. For clavulanic acid internal standard (Salicylamide) was added, derivatised with imidazole, and concentrated using a C18 Bond Elut cartridge as previously prescribed. Amoxicillin was assayed by HPLC.^{15,16} Plots of the standards were used to obtain the values of the samples after HPLC analysis. Lower limits of the assay were 8 µg/l or µg/kg for clavulanic acid and 32 µg/l or µg/kg for amoxicillin. Samples were assayed in batches of 6–10 with standards stored under the same conditions. Within and between assay precision was 96–99%.

Results

Mean mucosal biopsy weights were 1.69 mg, 1.4 mg, 1.25 mg, 1.79 mg, and 1.34 mg for the five dosage groups respectively. All samples had detectable levels of amoxicillin and clavulanic acid in the mucosal biopsy and serum samples, with the exception of two patients on the 750 mg dose of co-amoxiclav who had undetectable levels of clavulanic acid (tables 1 and 2). These patients had the lowest serum levels of clavulanic acid in their dosing group (0.875 and 0.91 mg/l), although they both had high mucosal and serum levels (11.15 and 8.8 mg/kg) of amoxicillin. The time of sampling after dosing was one hour and 1.75 hour respectively for these two patients. Forty one samples were taken 1–2 hours after the drug was administered and the remaining nine samples 2–3 hours after, so serum levels ap-

proximate to peak levels.¹⁷ Percentage penetration into the bronchial mucosa was 200% for amoxicillin (range 158–248%) with concentrations increasing with dose. Penetration for clavulanic acid was 118% (range 70–136%) and again concentrations increased with dose, except for the 750 mg oral dose where levels were lower than expected for no obvious reason.

Discussion

The concentrations of amoxicillin and clavulanic acid found in bronchial mucosa in this study are up to 20 times higher than in bronchial mucus and sputum.^{1–5} Higher concentrations of β-lactams occur in bronchial mucosa than in mucus¹⁸ and we have noticed a similar difference between bronchial mucus and mucosal levels for aztreonam (unpublished). The mucosal levels of amoxicillin are approximately double those where mean mucosal concentrations of amoxicillin were 75% of simultaneous serum levels.⁹ Preliminary results from a further study by the same group have shown similar levels to our own for both amoxicillin and clavulanic acid.¹³ The difference in results might be due to the use of a bioassay,⁹ different times between drug dosing and biopsy sampling, and different biopsy weights, suggesting different tissue site samples.

The mucosal amoxicillin levels are well above the MIC₉₀ of all common β-lactamase negative respiratory pathogens, even at the 250 mg dose of amoxicillin. Similarly, the mucosal concentrations of clavulanic acid, even at the 125 mg dose, may be sufficient to maximally inhibit the β-lactamases of all the common community acquired respiratory pathogens although this remains to be proven. For β-lactamase positive organisms such as *Staphylococcus aureus*, *Haemophilus influenzae*, and *Micrococcus cattarhalis* the MIC₉₀ for amoxicillin/clavulanic acid is easily achieved by the 375 mg dose. This may be important as there is evidence that tissue levels several times greater than the in vitro MIC level are necessary for optimal antimicrobial activity at certain sites of

infection.¹⁹ This degree of concentration in mucosa is similar to that seen with ciprofloxacin²⁰ and cefuroxime, but greater than that seen with other β -lactams.^{10 11 18 21 22}

The tissue concentrations reported provide a valid basis for the use of co-amoxiclav in the treatment of pneumonia and bronchitis. Ideal dosing regimens may be designed when the duration of therapeutic tissue levels, post antibiotic effects, and the relevance of in vitro studies on β -lactamase inhibition and antibacterial activity to the in vivo situation are understood. It appears that, for β -lactams, good bronchial mucosal levels are easily achieved. Poor clinical response or relapse, reported in some patients infected with sensitive organisms, may be due to a reservoir of organisms in the bronchial secretions where therapeutic levels are not so easily attained.

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