# Effect of azelastine on bronchoconstriction induced by histamine and leukotriene C4 in patients with extrinsic asthma

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ABSTRACT Azelastine, a new oral agent with antiallergic and antihistamine properties, has been shown to inhibit the effect of histamine and leukotriene (LT) in vitro, though not a specific leukotriene receptor antagonist. The effect of both a single dose (8·8 mg) and 14 days' treatment (8·8 mg twice ∠ daily) with azelastine on bronchoconstriction induced by LTC<sub>4</sub> and histamine has been examined in> 10 patients with mild asthma in a placebo controlled, double blind, crossover study. LTC<sub>4</sub> and = histamine were inhaled in doubling concentrations from a dosimeter and the results expressed as the cumulative dose (PD) producing a 20% fall in FEV<sub>1</sub> (PD<sub>20</sub>FEV<sub>1</sub>) and 35% fall in specific airways \( \tilde{S} \) conductance (PD<sub>35</sub>sGaw). The single dose of azelastine produced a significantly greater FEV<sub>1</sub> and  $\Box$ sGaw values than placebo at 3 hours, but this bronchodilator effect was not present after 14 days of \( \xi \) treatment. Azelastine was an effective H<sub>1</sub> antagonist; after a single dose and 14 days' treatment with placebo the geometric mean PD<sub>20</sub>FEV<sub>1</sub> histamine values (μmol) were 0.52 (95% confidence interval 2 0.14-1.83) and 0.54 (0.12-2.38), compared with 22.9 (11.5-38.3) and 15.2 (6.47-35.6) after azelastine 2.9(p < 0.01 for both). LTC<sub>4</sub> was on average 1000 times more potent than histamine in inducing  $\vec{z}$ bronchoconstriction. Azelastine did not inhibit the effect of inhaled LTC4; the geometric mean 3 PD<sub>20</sub>FEV<sub>1</sub> LTC<sub>4</sub> (nmol) after a single dose and 14 days' treatment was 0.60 and 0.59 with placebo compared with 0.65 and 0.75 with azelastine. The PD<sub>35</sub>SGaw LTC<sub>4</sub> was also unchanged at 0.66 and 0.73 for placebo compared with 0.83 and 0.74 for azelastine. Thus prolonged blockade of H<sub>1</sub> receptors did not attenuate the response to LTC<sub>4</sub>, suggesting that histamine and LTC<sub>4</sub> act on bronchial smooth muscle through different receptors. Four patients complained of drowsiness while taking azelastine but only one who was taking placebo and three patients complained of a bitter, metallic taste while taking azelastine.

Airway hyperresponsiveness to specific and nonspecific stimuli is characteristic of bronchial asthma, though the mechanisms are unclear. It has been suggested that the sulphidopeptide leukotrienes (LT), derived from membrane arachidonic acid, may play a part in airway hyperresponsiveness in asthmatic patients. <sup>1-5</sup> LTC<sub>4</sub> and LTD<sub>4</sub> are released in vitro and in vivo after allergen challenge<sup>67</sup> and both are extremely potent bronchoconstrictors in man. Inhaled LTE<sub>4</sub> has been reported to enhance airway responsiveness to inhaled histamine in patients with asthma. <sup>8</sup>

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Azelastine hydrochloride is a phthalazione derivative (4-(p-chlorobenzyl)-2-(hexahydro-1H-aze-pin-4yl)-(2H) phthalazione) with prolonged antigentation. It inhibits release of mediators from mast cells in response to antigen, calcium ionophore, concanavalin A, and compound 48/80 and in this respects is from 100 to 1000 times more potent than sodiume cromoglycate, theophylline, ketotifen, astemizole, and verapamil. It also inhibits the synthesis and release of leukotrienes from the rat peritoneal mast cell and is reported to modify leukotriene induced bronchoconstriction in guinea pigs. It is Recently azelastine has been shown to attenuate the early bronchoconstrictoruresponse to allergen in asthmatic patients. The effects of azelastine on leukotriene induced bronchoconstriction in patients with asthma has not been studied.

previously. We have examined the effect of a single dose (8·8 mg) and of two weeks' treatment (8·8 mg twice daily) with azelastine on the resting bronchomotor tone and histamine and LTC<sub>4</sub> induced bronchoconstriction in patients with mild extrinsic asthma in a double blind, placebo controlled, crossover study.

## Methods

We studied 10 patients (five of them women), mean age 32 (range 22–40) years, with mild extrinsic asthma and positive responses to skinprick tests with common inhaled allergens. None of the patients was taking oral corticosteroids, theophyllines, sodium cromoglycate, or antihistamine or anticholinergic drugs. Inhaled  $\beta$  agonists were stopped at least 12 hours before the test. The study was approved by the hospital ethics committee and informed written consent was obtained from each subject.

Patients received either 8·8 mg of azelastine twice daily or identical placebo for 14 days with a washout period of 14 days between treatments. Treatment was given double blind and in random order. The full blood count and serum urea and electrolyte concentrations were determined and liver function tests were carried out on their entry to the study and at the end of each treatment period. Airway response was assessed by measuring FEV<sub>1</sub> with a dry wedge spirometer (Vitalograph, Buckingham) and specific airways conductance (sGaw) with a constant volume body plethysmograph (Fenyves and Gut, Basel, Switzerland). The best of three attempts was recorded for FEV<sub>1</sub> and the mean of eight satisfactory manoeuvres for sGaw by on line data acquisition. 14

Histamine inhalation challenge was carried out in seven patients three hours after medication on the first and the 14th day of each treatment period. After pretreatment and post-treatment baseline FEV, had been recorded patients inhaled 10 breaths of phosphate buffered saline (control) from a Mefar 120 nebuliser (Mefar, Elettromedicali, Brescia, Italy) with a dosimeter set at a constant delivery time (1.0 s) and pressure (25 lb/in<sup>2</sup>, 172 kPa). The patient breathed from functional residual capacity to total lung capacity with a breath hold time of three seconds between inhalations. Patients proceeded to histamine challenge if the change in FEV, after inhalation of buffered saline was less than 5%. Each subject inhaled 10 breaths of histamine diphosphate dissolved in phosphate buffered saline in doubling concentrations (from 0.018 to 39.4  $\mu$ mol) until the FEV<sub>1</sub> had fallen more than 20% below the lowest FEV, value after inhalation of buffered saline (control). The results, expressed as the cumulative dose producing a 20% fall in the FEV<sub>1</sub> (PD<sub>20</sub>FEV<sub>1</sub>), were obtained from the log

dose-response curves.

LTC<sub>4</sub> challenge was performed one hour after the histamine inhalation challenge (four hours after treatment) and when FEV<sub>1</sub> readings had returned to within 5% of post-treatment baseline values. LTC<sub>4</sub> (Miles Laboratories, Slough) was stored at  $-70^{\circ}$ C in sealed ampoules until it was used, and appropriate dilution was made freshly with phosphate buffered saline (pH 7·4). The dilutions were kept in ice until immediately before they were placed in the nebuliser. LTC<sub>4</sub> was inhaled in doubling concentrations (from 0·025 to 3·2 nmol), and FEV<sub>1</sub> and sGaw were measured five, seven, 10, 15, and 20 minutes later.

PD<sub>20</sub>FEV<sub>1</sub> and PD<sub>35</sub>sGaw (cumulative dose producing a 35% fall in sGaw) were obtained from log doseresponse curves. The changes in FEV<sub>1</sub> and sGaw at each time after placebo and after azelastine were compared by analysis of variance. Log PD<sub>20</sub>FEV<sub>1</sub> and PD<sub>35</sub>sGaw were compared by analysis of variance and Student's t test.

## Results

The subjects' mean (SEM) FEV, was 87% (3.3%) predicted on entry to the study. There was no significant difference in the mean pretreatment baseline FEV, and sGaw values on the four study days. FEV, and sGaw were unchanged after a single dose and after 14 days' treatment with placebo (table). After the single dose of azelastine mean FEV, was 9.6% greater than after placebo (2.91 v 2.61 l) and mean sGaw 20% greater (1.4 v 0.95 s<sup>-1</sup> kPa<sup>-1</sup>) at 3 hours, both changes being significant (p < 0.05). After 14 days' azelastine, however, there was no significant difference in mean FEV, and sGaw values before and after azelastine. The drug was a potent H<sub>1</sub> receptor antagonist in the airways (table, fig 1). After a single dose and 14 days' treatment with placebo the geometric mean PD<sub>20</sub>FEV<sub>1</sub> (95% confidence interval) for histamine was 0.52 (0.14-1.83) and 0.54 (0.12-2.38)  $\mu$ mol. After a single dose of azelastine the geometric mean PD<sub>20</sub>FEV, was 22.9 (11.55–38.30)  $\mu$ mol, a 45 fold increase over placebo values (p < 0.01). After 14 days' treatment with azelastine the geometric mean PD<sub>20</sub>FEV<sub>1</sub> was 15.2  $\mu$ mol (6.47–35.6), a 28 fold increase over placebo values (p < 0.01). The difference in inhibition after a single dose and 14 days' treatment was not significant.

In seven patients who underwent LTC<sub>4</sub> and histamine challenges, LTC<sub>4</sub> was about 1000 times more potent than histamine. Azelastine had no effect on LTC<sub>4</sub> induced bronchoconstriction (table, fig 2). The geometric mean PD<sub>20</sub>FEV<sub>1</sub> for LTC<sub>4</sub> was 0.60 (95% confidence interval 0.19–1.97) and 0.59 (0.22–1.54) nmol after a single dose and 14 days' treatment with placebo respectively, compared with 0.65 (0.25–1.70) and 0.75 (0.46–1.22) nmol after a single dose and 14

Effect of a single dose and of 14 days' treatment with azelastine and placebo on baseline respiratory function and the responses to inhalation challenge in patients with asthma

	Placebo		Azelastine			
	Day 1	Day 14	Day 1		Day 14	
Baseline respiratory	function (mean (SEM))					
$FEV_1(1)$ : n = 10						
Before drug	2.79 (0.16)	2.70 (0.16)	2.61 (0	)·18)	2.71 (0.16)	
After drug	2·75 (0·17)	2·76 (0·14)	2.91 (0	)·21)	2·79 (0·14)	
p value	NS	NS	< 0.05	,	NS	
P						
sGaw (s-1 kPa-1): n	= 10				2·71 (0·16) 2·79 (0·14) NS 0·93 (0·13) 0·97 (0·16) NS	
Before drug	0.81 (0.13)	0.84 (0.10)	0.95 (0	0.16)	0.93 (0.13)	
After drug	0.85 (0.16)	0.86 (0.14)	1.14 (0		0.97 (0.16)	
p value	NS (C 15)	NS	< 0.05	,	NS	
p varac	115	110	10 05		145	
Responses to inhalat	ion challenges (geometric med	in (05% confidence interval))				
$LTC_4$ (nmol): $n = 1$		ar (>5>0 confluence ance var)				
PD <sub>m</sub> FEV	0.60 (0.19 to 1.97)	0·59 (0·22 to 1·54)	0.65 ((	0·25 to 1·70)	0.75 (0.46 to 1	.221
PD <sub>15</sub> sGaw	0.66 (0.27 to 1.65)	0.73 (0.39 to 1.40)		0.32 to 2.14)	0·74 (0·42 to 1	.33
p value*	0.00 (0.27 to 1.03)	0.73 (0.33 to 1.40)	NS (C	3.32 (0 2.14)	NS	33)
	_ 7.		149		143	
Histamine (µmol): n		0.54 (0.10 + 0.20)	22.0.(11	CC + 20 21)	150/6/71 25	_
PD <sub>20</sub> FEV	0·52 (0·14 to 1·83)	0·54 (0·12 to 2·38)		55 to 38·31)	15·2 (6·47 to 35·	5)
p value*			< 0.01		< 0.01	

<sup>\*</sup>In the comparison with placebo.

sGaw, specific airways conductance; LTC<sub>4</sub>, leukotriene C<sub>4</sub>; PD<sub>20</sub>FEV<sub>1</sub>, provocative dose causing a 20% fall in FEV<sub>1</sub>; PD<sub>35</sub>, provocative dose causing a 35% fall in sGaw.

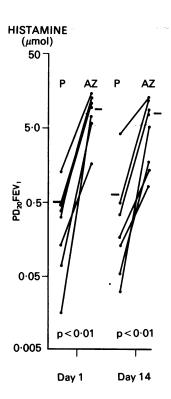


Fig 1 Individual and geometric mean (—)  $PD_{20}FEV_1$  histamine values after treatment with placebo (P) and azelastine (AZ).  $PD_{20}FEV_1$ —provocative dose causing a 20% fall in  $FEV_1$ .

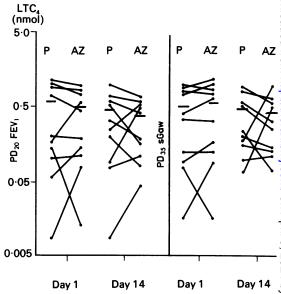


Fig 2 Individual and geometric mean (—) PD<sub>20</sub>FEV<sub>1</sub> and PD<sub>35</sub>sGaw leukotriene C<sub>4</sub> values after treatment with placebo (P) and azelastine (AZ). PD<sub>20</sub>FEV<sub>1</sub>—provocative dose causing a 20% fall in FEV<sub>1</sub>; PD<sub>35</sub>sGaw—provocative dose causing a 35% fall in specific airways conductance.

days' treatment with azelastine. There was no significant period effect (analysis of variance).

Four patients taking azelastine complained of g drowsiness and three patients complained of a bitter g

metallic taste. One patient taking placebo noted drowsiness. There were no significant changes in haematological and biochemical indices after azelastine treatment.

#### Discussion

Azelastine in a single dose produced a small but significant increase in FEV, and sGaw in the patients in this study. After 14 days' of treatment, however, this bronchodilator effect was attenuated and no significant difference between azelastine and placebo was observed. Our results contrast with the observations of Ollier et al,13 who failed to show any change in mean FEV, after single or multiple dose treatment with azelastine; but these workers did show a significant increase in mean sGaw after a single dose of azelastine, and this increase in sGaw was present at three weeks. The difference between our results and those of Ollier et al<sup>13</sup> may be related to differences in doses (2.2 mg and 4.4 mg compared with 8.8 mg) and also patient selection. Azelastine is a potent H<sub>1</sub> receptor antagonist and its bronchodilator effect is likely to be due to its airway H, receptor blockade. A similar degree of bronchoconstriction has been observed with other H. receptor antagonists, such as clemastine, chlorpheniramine, and terfenadine. 15-17 In addition, ketotifen, an antiallergic compound with potent H, receptor blocking activity, also produces a small but important amount of bronchodilatation when inhaled. 18 Azelastine shifted the histamine dose-response (PD<sub>20</sub>FEV<sub>1</sub>) curve 45 fold to the right after a single dose and 28 fold after 14 days of treatment. Although the mean inhibition of histamine induced bronchoconstriction by azelastine was higher after a single dose than after 14 days of treatment, the difference was not significant. This large effect of azelastine on histamine induced bronchoconstriction confirms that azelastine is a very effective H<sub>1</sub> receptor in blocking activity in human airways.

In contrast to the findings with histamine, the bronchoconstrictor response to LTC<sub>4</sub> was not altered by either a single dose or 14 days' treatment with azelastine. In the present study LTC<sub>4</sub> was about 1000 times more potent than histamine, and this observation is consistent with previous reports.<sup>38</sup>

The mechanism of histamine hyperresponsiveness is unclear. Histamine acts on bronchial smooth muscle by interaction with at least two distinct receptors, H<sub>1</sub> and H<sub>2</sub> receptors, and it also increases the rate of firing of bronchial irritant receptors, an effect that can be blocked by atropine. <sup>19</sup> Human airway smooth muscle contracts in vitro in response to histamine, but when H<sub>1</sub> receptors are blocked histamine produces relaxation, an effect attributed to H<sub>2</sub> receptor stimulation as it can be blocked by the H<sub>2</sub> antagonist metiamide. <sup>20 21</sup>

Terfenadine and astemizole are specific H<sub>1</sub> receptor antagonists and lack anticholinergic and antiserotonin activity. These drugs have been shown to modify exercise<sup>22 23</sup> and allergen induced bronchoconstriction<sup>24</sup> in patients with asthma. In addition to H<sub>1</sub> receptor antagonism, many antihistamines at high concentrations have the capacity in vitro to stabilise mast cells and ketotifen falls into this class.<sup>24</sup> Ketotifen, however, offers no greater protection against the immediate response to inhaled antigen than can be attributed to its capacity to block histamine.<sup>24</sup>

Astemizole has also been reported to attenuate the early component (2–15 min) of the bronchoconstrictor response to antigen challenge.<sup>25</sup> The protective effect of azelastine in the immediate asthmatic response to allergen inhalation reported by Ollier *et al*<sup>13</sup> can also be explained by its potent H<sub>1</sub> receptor blocking activity.

The time course of the bronchoconstrictor response to leukotrienes and histamine differ in vivo and in vitro in man: leukotrienes have a slow onset of action, which is more prolonged and persistent than that of histamine. After histamine the peak response is reached within 4-8 minutes of inhalation whereas with leukotrienes the response is slower, reaching a peak at 20 minutes.22627 Recently Arm et ale have shown that inhaled LTE, can enhance histamine responsiveness in asthmatic patients but not in normal subjects. Holroyde et al 27 and Barnes et al,28 using the specific leukotriene antagonists FPL 55712, FPL 59257, and L 49923, have shown that the drugs will effectively inhibit LTC4 and LTD4 mediated airway responses without modifying histamine responsiveness in normal subjects. H<sub>1</sub> receptor blocking drug<sup>11</sup> did not inhibit leukotriene induced bronchoconstriction and our results with azelastine in this respect are consistent with these observations. Leukotrienes and histamine act independently on the bronchial smooth muscle through specific receptors and studies in animal lung tissues have identified a site specific for LTC4 and LTD<sub>4</sub>.<sup>29</sup> It has been suggested that there may be heterogeneity of leukotriene receptors in view of the very different molar ratios of LTC4, LTD4 and LTE4 required to elicit identical biological effects in different tissues, 30 31 and because the rank order of potency for the leukotrienes in contracting guinea pig tracheal spirals differs from that for contraction of parenchymal strips. 32 33 Drugs may vary in their ability to block responses according to their different receptor affinities.34 FPL 55712 was found to have a higher affinity for the LTD<sub>4</sub> receptor, which is consistent with its more effective antagonism of the LTD, induced contractile response of lung parenchymal strips.35 The differences between the effects in animals and in patients with asthma of azelastine.11 sodium cromoglycate, and the calcium channel blocker

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verapamil<sup>36</sup> on leukotriene induced bronchoconstriction may be related to species differences and to the lack of a good animal model that can mimic human asthma. Further studies are required to elucidate the role of azelastine and similar compounds in asthma.

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### References

- Dahlen SE, Hedqvist P, Hammarstrom S, Samuelsson B. Leukotrienes are potent constrictors of human bronchi. Nature 1980;288:484-6.
- 2 Creese BR, Bach MK. Hyperreactivity of airways smooth muscle produced in vitro by leukotrienes. *Prostaglan-dins and Leukotrienes in Medicine* 1983;11:161-9.
- 3 Griffen M, Weiss JW, Leitch AG, et al. Effects of leukotriene D on the airways in asthma. N Engl J Med 1983;308:436-9.
- 4 Smith LJ, Greenberger PA, Patterson R, Krell RD, Bernstein PR. The effect of inhaled leukotriene D<sub>4</sub> in humans. Am Rev Respir Dis 1985;131:368-72.
- 5 Lee TH, Austen KF, Corey EJ, Drazen JM. Leukotriene E<sub>4</sub>-induced airway hyperresponsiveness of guinea pig tracheal smooth muscle to histamine and evidence for three separate sulphidopeptide leukotriene receptors. Proc Natl Acad Sci USA 1984;81:4922-5.
- 6 Dahlen SE, Hansson G, Hedqvist P, Bjorck T, Granstrom E, Dahlen B. Allergen challenge of lung tissue from asthmatics elicits bronchial contraction that correlates with the release of leukotrienes C<sub>4</sub>, D<sub>4</sub> and E<sub>4</sub>. Proc Natl Acad Sci USA 1983;80:1712-6.
- 7 Creticos PS, Peters SP, Adkinson NF, et al. Peptide leukotriene release after antigen challenge in patients sensitive to rag weed. N Engl J Med 1984;310:1626-30.
- 8 Arm JP, Spur BW, Lee TH. Leukotriene E<sub>4</sub> (LTE<sub>4</sub>) enhances airway histamine responsiveness in asthmatic subjects [abstract]. *Thorax* 1987;42:220.
- 9 Katayama S, Akimoto N, Shionoya H, Morimoto T, Katoh Y. Antiallergic effect of azelastine hydrochloride on immediate type hypersensitivity reaction in vivo and in vitro. Arzneimittelforschung 1981;31: 1196-203.
- 10 Chand N, Pillar J, Diamantis W, Perhach JL, Sofia RD. Inhibition of calcium ionophore (A23187)—stimulated histamine release from rat peritoneal mast cells by azelastine: implications for its mode of action. Eur J Pharmacol 1983;96:227-33.
- 11 Chand N, Nolan K, Diamantis W, Perhach JL, Sofia RD. Inhibition of leukotriene (SRS-A) mediated bronchospasm by azelastine, a novel orally effective antiasthmatic drug [abstract]. J Allergy Clin Immunol 1983;71:149.

12 Chand N, Pillar J, Nolan K, Diamantis W, Sofia RD. Inhibition of 5-HETE, LTB<sub>4</sub> and LTC<sub>4</sub> synthesis by azelastine and its d- and 1- isomers in rat mixed peritoneal mast cells [abstract]. Am Rev Respir Dis 1987;135:318.

- 13 Ollier S, Gould CAL, Davis RJ. The effect of single and multiple dose therapy with azelastine on the immediate asthmatic response to allergen provocation testing. J Allergy Clin Immunol 1986;78:358-64.
- 14 Roberts JA, Pugh JR, Thomson NC. A new adaptable computerised system for measurement of specific airways conductance. Br J Dis Chest 1986;80:218-28.
- Popa VT. Bronchodilating activity of an H<sub>1</sub> blocker chlorpheniramine. J Allergy Clin Immunol 1977;59: 54-63.
- 16 Nogrady SG, Bevan C. Inhaled antihistamines—bronchodilatation and effects on histamine and methacholine induced bronchoconstriction. *Thorax* 1978;33:700-4.
- 17 Patel KR. Effect of terfenadine on methacholine induced bronchoconstriction in asthma. J Allergy Clin Immunol 1987;79:35-8.
- 18 Dorward AJ, Patel KR. Inhaled ketotifen in exerciseinduced asthma—a negative report. Eur J Respir Dis 1985;67:378-80.
- 19 Drazen JM, Austen KF. Atropine modification of the pulmonary effects of chemical mediators in the guinea pig. J Appl Physiol 1975;38:834-8.
- 20 Dunlop LS, Smith AP, Piper PJ. The effect of histamine antagonists on antigen induced contractions of sensitised human bronchus in vitro proceedings. Br J Pharmacol 1977;59:475P.
- 21 Nathan RA, Segali N, Schocket AL. A comparison of the actions of H<sub>1</sub> and H<sub>2</sub> antihistamines on histamine induced bronchoconstriction and cutaneous wheal response in asthmatic patients. J Allergy Clin Immunol 1981;67:171-7.
- 22 Patel KR. Terfenadine in exercise induced asthma. Br Med J 1984;288:1496-7.
- 23 Clee MD, Ingram CG, Reid PC, Robertson AS. Theeffect of astemizole on exercise induced asthma. Br J Dis Chest 1984;78:180-3.
- 24 Holgate ST, Emanuel MB, Howarth PH. Astemizole and other H<sub>1</sub> antihistaminic drug treatment of asthma. J Allergy Clin Immunol 1985;76:375-80.
- 25 Church ML, Gradidge CF. Inhibition of histamine = release from human lung in vitro by antihistamines and related drugs. Br J Pharmacol 1980;69:663-7. ⋈
- 26 Barnes N, Piper PJ, Costello J. Comparative effects of inhaled leukotriene C<sub>4</sub>, Leukotriene D<sub>4</sub> and histamine in normal human subjects. *Thorax* 1984;39:500-4.
- 27 Holroyde MC, Altounyan RE, Cole M, Dixon M, Elliott C EV. Bronchoconstriction produced in man by leukotrienes C and D. Lancet 1981;ii:17-8.
- 28 Barnes N, Piper PJ, Costello J. The effect of an oral leukotriene antagonist L-649923 on histamine and leukotriene D₄ induced bronchoconstriction in normal man. J Allergy Clin Immunol 1987;79:816-21.
- 29 Kuehl FA, De Haven RN, Pong SS. Lung tissue receptors for sulfidopeptide leukotrienes. J Allergy Clin Immunol 1984;74:378-81.
- 30 Lewis RA, Drazen JM, Austen KF, Clark DA, Corey EJ

- Identification of the C(6)-S-conjugate of leukotriene A with cysteine as a naturally occurring slow reacting substance of anaphylaxis (SRS-A). Importance of the 11-cis-geometry for biological activity. *Biochem Biophys Res Commun* 1980;96:271-7.
- 31 Burke JA, Levi R, Guo ZG, Corey EJ. Leukotrienes C<sub>4</sub>, D<sub>4</sub> and E<sub>4</sub>: effects on human and guinea pig cardiac preparations in vitro. J Pharmacol Exp Ther 1982;221:235-41.
- 32 Drazen JM, Austen KF, Lewis RA, et al. Comparative airway and vascular activities of leukotrienes C-1 and D in vivo and in vitro. Proc Natl Acad Sci USA 1980;77:4354-8.
- 33 Lee TH, Austen KF, Corey EJ, Drazen JM. Leukotriene

- E<sub>4</sub> induced airway hyperresponsiveness of guinea pig tracheal smooth muscle to histamine and evidence for three separate sulphidopeptide leukotriene receptors. *Proc Natl Acad Sci USA* 1984:81:4922-5.
- 34 Lewis RA, Austen KF. The biologically active leukotrienes. Biosynthesis, metabolism, receptors, functions and pharmacology. J Clin Invest 1984;73:889-97.
- 35 Pong SS, De Haven RN. Characterization of a leukotriene D<sub>4</sub> receptor in guinea pig lung. *Proc Natl Acad Sci USA* 1983;80:7415-9.
- 36 Advenier C, Cerrina J, Duroux P, Floch A, Pradel J, Renier A. Sodium cromoglycate, verapamil and nicardipine antagonism to leukotriene D<sub>4</sub> bronchoconstriction. Br J Pharmacol 1983;78:301-6.