

Role of bronchoalveolar lavage in the evaluation of interstitial pneumonitis in recipients of bone marrow transplants

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ABSTRACT Forty episodes of pneumonitis in 30 recipients of allogeneic bone marrow transplant were investigated by fiberoptic bronchoscopy and bronchoalveolar lavage. A positive diagnosis was made in 32 episodes of pneumonitis (24 patients), giving a diagnostic yield of 80%. In 31 of these the diagnosis was made within 24 hours of bronchoscopy and this enabled the appropriate treatment to be instituted early. Eighteen patients recovered from their primary infection, although two died subsequently of respiratory failure due to postpneumonic lung destruction. Ten patients later developed a second episode of pneumonitis and a diagnosis was made in nine of these. Only three survived a second episode. Bronchoalveolar lavage was well tolerated by all patients and there was no morbidity or mortality that could be directly attributed to the procedure. Bronchoalveolar lavage is a safe and valuable early diagnostic procedure for the investigation of pulmonary complications in patients who have received bone marrow transplants.

Pulmonary complications are major causes of morbidity and mortality in patients who have received bone marrow transplantation.^{1,2} In some series^{1,3-5} as many as half of these patients have suffered from a major pulmonary problem in the six months immediately after transplantation and up to 70% of these (or 35% of all patients receiving transplants) have died from this complication.^{1,3-5} If results are to be improved, a diagnosis must be made early with a technique that has few important complications itself and results in the institution of successful treatment. Bronchoalveolar lavage has been found to be a safe and effective method of sampling peripheral lung tissue⁶ and has been used to aid diagnosis in heterogeneous populations of immunosuppressed patients,^{7,8} patients with renal transplants,⁹ and patients with the acquired immune deficiency syndrome (AIDS).^{10,11} Studies of heterogeneous groups of patients may be unhelpful in planning management within a homogeneous group. Furthermore, the value of bronchoalveolar lavage as the sole method of investigation of pneumonitis in a homogeneous population of patients who have received bone marrow

transplants has not previously been reported. For these reasons we have investigated 30 recipients of allogeneic bone marrow transplants who developed clinical evidence of pneumonitis.

Methods

PATIENTS

Forty consecutive episodes of suspected pneumonitis (defined as inflammation, as judged by clinical, radiological, or physiological signs, in the peripheral air spaces) were investigated in 30 recipients of allogeneic bone marrow transplants. Nineteen patients were male and ages ranged from 12 to 47 (median 27) years. The pretransplant haematological diagnoses are outlined in table 1. Treatment regimens used to induce remission varied between individuals; but the drugs commonly used were daunorubicin, cytosine arabinoside, and 6-thioguanine for acute myeloid leukaemia (AML); vincristine, prednisolone, daunorubicin, L-asparaginase, etoposide (VP16/213), thioguanine, methotrexate, and 6-mercaptopurine for acute lymphoblastic leukaemia (ALL); and busulphan or hydroxyurea, or both, for chronic granulocytic leukaemia (CGL). All patients received T cell depleted bone marrow from matched (n = 23) or partially matched (n = 7) sibling donors. All later developed clinical features of pneumonitis (table 2). Most

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Table 1 Pretransplant haematological diagnoses in the 30 patients

Disease	Stage	No of patients
Acute lymphoblastic leukaemia	Remission	6
	Relapse	1
Acute myeloid leukaemia	Remission	8
	Relapse	4
Chronic granulocytic leukaemia	Chronic phase	9
	Accelerated phase	1
Myelofibrosis	—	1

patients developed first episodes of pneumonitis 40–120 days after transplantation. Eight patients of those who developed second episodes did so in the first year after transplantation. One patient suffered a second episode during the second year and one during the third year after transplantation. In the three instances in which chest radiographs were normal, a diagnosis of pneumonitis was made on the basis of the development of cough, breathlessness, and lower transfer factor (TLCO) values than earlier after transplantation.

PRETRANSPLANT CONDITIONING AND TRANSPLANT

Patients with AML in the first complete remission, all those with CGL who received HLA matched transplants, and a patient with myelofibrosis were given two doses of cyclophosphamide 60 mg/kg body weight on days -4 and -3 and total body irradiation on day 0. The average dose to the lung was 670–830 cGy (rad). All patients with ALL and patients receiving mismatched transplants were given two doses of cyclophosphamide 45 mg/kg body weight on days -4 and -3 plus six doses of cytosine arabinoside 3 mg/m², with total body irradiation as above.

Immediately after transplantation all patients were isolated in reverse barrier nursing conditions and were given prophylactic acyclovir, ketoconazole, and amphotericin suspension together with gut decon-

tamination using neomycin and colistin. Co-trimoxazole, as prophylaxis against *Pneumocystis carinii* infection, was added when the neutrophil count rose above 1×10^9 /ml.

SAMPLE COLLECTION

Bronchoalveolar lavage was performed using an Olympus BF1T fiberoptic bronchoscope. The upper airways were anaesthetised with 10% lignocaine topical spray and 2 ml of 4% lignocaine were applied to the vocal cords under direct vision. Aliquots of 2% lignocaine were used to anaesthetise the lower respiratory tract. Supplemental oxygen was administered at a rate of 4 l/min throughout and for four hours after the procedure.

The bronchoscope was wedged into the orifice of a subsegmental bronchus of the middle lobe or lingula in patients with either normal chest radiographs or diffuse radiological changes. Where chest radiographs showed local changes the appropriate segment was lavaged. Bronchoalveolar lavage was performed with 3×60 ml aliquots of normal saline warmed to 37°C and buffered to pH 7.4 by the addition of 175 μEq (μmol) of sodium bicarbonate to 500 ml normal saline. Each aliquot was aspirated immediately after its instillation into silicon coated glass bottles maintained at 4°C.

PROCESSING LAVAGE SAMPLES

All lavage samples were investigated by routine Gram staining bacterial, culture and staining and culture for mycobacteria. Samples were also stained with Grocott's silver stain to detect fungi and *Pneumocystis carinii*. The lavaged cells were examined cytologically. For viral studies the fluid was inoculated into cultures of human lung embryo fibroblasts. Samples were also investigated for cytomegalovirus by using the detection of early antigen fluorescent foci (DEAFF) test. This technique has been fully described elsewhere.¹² In addition, samples of lavage fluid were centrifuged at 350 g for 10 minutes and aliquots of the cell pellet were stained directly with monoclonal antibodies raised against cytomegalovirus protein.¹²

Table 2 Clinical, radiological, and physiological features of 30 patients with 40 episodes of pneumonitis after bone marrow transplantation

Symptoms (n = 40)		Signs (n = 40)		Radiographic shadowing (n = 40)		Transfer factor (n = 29)	PaO ₂ (kPa) in room air (n = 13)
SOB	40	Fever	38	Localised	18	Reduction by >20% of pred value 27	Mean 7.9 Range 5.1–12.9
Cough	21	Wheeze	4	Diffuse	19		
Sputum	1	Crackles	26	Normal	3		
Haemoptysis	1	None in lung	10				

SOB—short of breath; PaO₂—arterial oxygen tension.

Results

A diagnosis was made in 32 of the 40 episodes of pneumonitis by lavage alone. No diagnosis was made from eight lavages, but in four cases the patients recovered rapidly with antimicrobial treatment that had already been started empirically before bronchoscopy and a presumptive diagnosis of a treated bacterial infection was made. One patient underwent open lung biopsy but this also failed to help with diagnosis. In the remaining three cases the patients died without a diagnosis of their pulmonary problem. Postmortem examination of two of these patients also failed to provide a diagnosis of their condition, which was finally termed idiopathic pneumonitis. Postmortem examination of the third patient produced samples from which a diagnosis of giant cell pneumonitis was made. The histological appearances were similar to those seen in measles pneumonitis, but the virus was never isolated. The diagnostic sensitivity of bronchoalveolar lavage in this series was therefore 86% (32/37).

INFECTIVE AGENTS ISOLATED

The types of infection in the 32 positive lavage samples are shown in tables 3 and 4. More than one pathogen was isolated from the lavage fluid from four patients. The most common pathogen found was cytomegalovirus, which was present in 20 lavage samples (15 patients); in four cases it was found in association with other microorganisms. Of these 20, 19 were positive by DEAFF and two also on direct staining of the lavage cells. The twentieth was negative by DEAFF and direct staining but was positive in conventional cell culture, and this was the only result that was not available either on the day of bronchoscopy or the following day.

DEAFF was also performed on the saliva of 29 patients on the day of bronchoscopy. Only two patients out of the 14 with cytomegalovirus present in the lavage fluid also had cytomegalovirus in the saliva; it was also present in the saliva of two out of 15

patients in whom no cytomegalovirus was detected in the lavage fluid.

All samples were put up in cultures of human lung embryo fibroblasts. No other viruses were identified by this method in this group of patients and several problems were encountered with the technique. Four of the cultures became contaminated with bacteria and fungi during their three week incubation and three others were also of no diagnostic use owing to toxicity of the fibroblasts. This latter problem is particularly troublesome with bronchoalveolar lavage fluid, although it may also occur to a lesser extent with samples of blood, urine, and saliva. In addition, cultures had to be incubated for 5–21 days for detection of cytomegalovirus.

Pneumocystis carinii was isolated from four lavage samples, in two in association with cytomegalovirus. The bacteria found were *Staphylococcus aureus* and *Pseudomonas aeruginosa*. Two patients had repeated infections with these organisms despite appearing to recover from the first episode. Fungi were isolated from a total of five lavage samples. In one case *Aspergillus fumigatus* was found in association with cytomegalovirus, and in another cryptosporidial oocytes were found concurrently with cytomegalovirus. Leukaemic relapse was diagnosed by lavage in one patient.

There was no relationship between type of radiological abnormality and pathogen isolated. Patients with cytomegalovirus in particular presented with diffuse, focal, and normal chest radiographs.

RESPONSE TO TREATMENT

The results from lavage were available on either the same or the following day in all except one case, and this meant that appropriate changes could be made to treatment within 24 hours of bronchoscopy. Eleven of the 12 patients with a first episode of cytomegalovirus pneumonitis were diagnosed by the DEAFF test within 24 hours; 10 of these were treated with cytomegalovirus hyperimmune globulin and the eleventh with Sandoglobulin and the antiviral agent Gan

Table 3 Results and outcome after first episode of pneumonitis in 30 patients

Organism(s)	No of patients	% BAL diagnoses	No of survivors	No developing second infection
CMV	12	40	7	6
CMV and <i>Pneumocystis carinii</i>	2	7	0	—
CMV and <i>Aspergillus fumigatus</i>	1	3	0	—
<i>Pneumocystis carinii</i>	2	7	1	1
Bacteria: <i>Staphylococcus aureus</i>	2	7	2	1
<i>Pseudomonas aeruginosa</i>	1	3	1	1
Fungi: <i>Candida albicans</i>	1	3	1	—
<i>Aspergillus fumigatus</i>	1	3	0	—
Leukaemic relapse	1	3	0	—
No diagnosis	7	23	5	1

CMV—cytomegalovirus; BAL—bronchoalveolar lavage.

Table 4 Results and outcome of second episode of pneumonitis in 10 patients

Organism	No of patients	No of survivors	Original organism
CMV	4	3	CMV 4
CMV and <i>Cryptosporidium</i>	1	0	CMV
Bacteria: <i>Staphylococcus aureus</i>	1	0	<i>S aureus</i>
<i>Pseudomonas aeruginosa</i>	1	0	<i>P aeruginosa</i>
Fungi: <i>Candida albicans</i>	1	0	<i>Pneumocystis carinii</i>
<i>Aspergillus fumigatus</i>	1	0	None found
No diagnosis	1	0	CMV

CMV—cytomegalovirus.

ciclovir (BW759U, DHPG). Seven patients survived this first episode but six of them went on to develop a second episode (table 3). The patient who was not diagnosed within 24 hours was not treated for cytomegalovirus pneumonitis and died. Two patients had cytomegalovirus and *Pneumocystis carinii* infections and one recovered from the infection, but he sustained considerable lung damage and later developed bilateral pneumothoraces resistant to treatment; and he subsequently died. One patient recovered from *P carinii* alone, and one from *Candida albicans*. Three patients with bacterial infections recovered from a first episode.

A diagnosis of an initial infection was made by bronchoalveolar lavage in 22 patients (table 3) and 12 of these survived (55%). A further five in whom no organism was isolated also survived. Ten of these 17 survivors, however, later relapsed after successful treatment of their first infection. The time taken to relapse varied from three weeks (two patients) to two years three months but most patients remained well for from two to eight months between episodes. The two patients who relapsed after only three weeks had initially responded to treatment with clearing of the chest radiograph, reduced temperature, and improvement in symptoms and signs; but this response was short lived, and when they did relapse deterioration was swift and both died. The other eight patients recovered completely between episodes, with gradual improvement in pulmonary function in addition to the other indices.

Three patients recovered completely from a second episode of interstitial pneumonitis and in all three both episodes were due to infection with cytomegalovirus (table 4).

In the 30 patients studied there were 40 episodes of pneumonitis and recovery followed 20 of these (50%). Thirty one of these 40 episodes were due to infection diagnosed by bronchoalveolar lavage and recovery followed 15 of these individual episodes of infection (48%). At the end of the study a total of 10 of the 30 patients (33%) had survived episodes of pneumonitis and three of these had overcome two infections with cytomegalovirus.

COMPLICATIONS OF THE PROCEDURE

Bronchoalveolar lavage was well tolerated by all patients, even those who were already extremely unwell. There was occasional contact bleeding in those whose platelet counts were below $20 \times 10^9/l$ but this was minimised by the infusion of platelet concentrate both before and during the procedure. Two children had their bronchoscopies under general anaesthesia and a further four patients had them while being ventilated for respiratory failure. Bronchoscopy and lavage were performed easily via the endotracheal tube without appreciably compromising ventilation. No morbidity or mortality could be directly attributed to the lavages.

Discussion

This study describes pulmonary infections diagnosed by bronchoalveolar lavage alone that were encountered in a homogeneous group of recipients of bone marrow transplants and the subsequent outcome of these patients after prompt diagnosis and treatment. The success of bone marrow transplant programmes has been hampered by diverse and serious pulmonary complications. These problems are common^{1 5 13 14} and result in considerable morbidity and mortality. The distinction between infective and non-infective processes is rarely clear on the basis of clinical criteria alone. In addition to infection, lung damage can result from graft versus host disease, irradiation, and previous drug treatment used to induce remission. Furthermore, leukaemic relapse may itself present as pneumonitis. Since patients have received T cell depleted bone marrow, graft versus host disease is no longer a major problem in our unit¹⁵ and we have found that infection is the greatest cause of morbidity and mortality. The infections seen include those caused by bacteria, especially the Gram negative group, fungi, protozoa, and viruses. The prophylactic use of ketoconazole and co-trimoxazole has reduced the incidence of some of these infections, and acyclovir has significantly reduced the problems due to herpes simplex and varicella zoster viruses but has had no impact on the incidence of cytomegalovirus infection. Cytomegalovirus remains the most com-

mon pathogen encountered in our unit.

It is important to make an early diagnosis of pulmonary infection in the immunocompromised patient as rapid diagnosis allows the early institution of appropriate treatment and changes to treatment which has been started empirically. Drugs that may be toxic (including those with known toxicity to the bone marrow) can be added with confidence once a positive diagnosis is made, and in our study four patients could begin treatment with co-trimoxazole and two with amphotericin. Three of these patients survived their infection. Infections with cytomegalovirus are associated with >90% mortality⁵ and, until recently, a diagnosis of cytomegalovirus pneumonitis was not particularly helpful as no effective therapeutic agents were available. Recent reports, however, indicate that cytomegalovirus hyperimmune globulin and some of the new antiviral agents have been used successfully in some patients^{16,17}; so it is now just as important to diagnose cytomegalovirus early in the course of the infection as it is to diagnose infections due to bacteria, *P carinii*, and fungi. Early diagnosis also means that in the future new antiviral agents can be assessed.

Several techniques have been advocated for diagnosing interstitial pneumonitis in immunocompromised patients.¹⁸⁻²³ Diagnostic yields are variable and serious complications have been reported with transtracheal aspiration,²³⁻²⁵ transbronchial biopsy,²⁶⁻²⁸ percutaneous needle biopsy,^{29,30} and open lung biopsy.¹⁹ The problems associated with these techniques are both more frequent and more severe in patients with refractory thrombocytopenia and in those being mechanically ventilated. Most of our patients had platelet counts below $40 \times 10^9/l$; four were being mechanically ventilated and two had to have bronchoscopy under general anaesthesia. For these reasons we considered that biopsy techniques were inappropriate and confined our invasive investigations to bronchoalveolar lavage. A positive diagnosis was made in 32 out of the 40 episodes of pneumonitis (80%), a yield that compares favourably with the results from other centres where lavage has been used to investigate mixed groups of immunosuppressed patients,^{8,10} patients with acquired immune deficiency syndrome (AIDS),^{10,11} and recipients of renal transplants.⁹ The results also compare favourably with those of other studies where the yield from bronchoalveolar lavage has been compared with that from transbronchial biopsies.^{10,11,31} In our series, although eight of the 40 lavages had a negative result, tissue investigation by open lung biopsy (one patient) and postmortem examination (two patients) also failed to give a diagnosis either histologically or by culture. The negative results in these three patients should therefore be

accepted as true negatives. Necropsy in a fourth patient did, however, allow a histological diagnosis to be made, but again no causative organism was isolated. Four of the patients with negative results from lavage recovered rapidly after bronchoscopy and presumptive diagnosis of a treated bacterial infection was made. This seems a reasonable assumption as they had all been started on empirical broad spectrum antibiotic treatment at the start of their illness and did not have bronchoscopy immediately.

Within the infective group the identification of viral cause can be particularly difficult. The use of monoclonal antibodies in a double layer immunofluorescence technique has improved the speed of diagnosis of cytomegalovirus infections. In our institution the DEAFF test has been found to have a specificity of 100% and a sensitivity of 80%. DEAFF has also been performed on lavage material from patients with cryptogenic fibrosing alveolitis and sarcoidosis and there have been no false positives. These data, with the negative results obtained with saliva, support the contention that when cytomegalovirus is found in bronchial lavage fluid it may be considered a true pathogen. Cytomegalovirus takes from five to 12 days to grow in conventional cell culture and this is too long to be of practical use. The long incubation period also means that fungal or bacterial contamination of the culture is more likely. Cultures were also lost owing to the toxic effect of the bronchoalveolar lavage material on the fibroblasts. For many reasons therefore cell culture is an unsatisfactory method for diagnosing cytomegalovirus pulmonary infections in immunocompromised patients. Two of the DEAFF positive samples were also positive by direct staining. These were from patients with advanced disease and extensive chest radiographic changes. Although the sensitivity of direct staining was much lower than that found with the DEAFF technique, it has the advantage of even greater speed as the results are available within five hours of bronchoscopy.

It is sometimes questioned whether the identification of fungi, such as *Candida albicans*, in bronchoalveolar lavage fluid is a reliable indicator of their presence in the lung; but, in an immunocompromised patient where no other pathogen has been isolated, we believe that a reasonable growth of candida from lavage fluid should be regarded as significant, provided that the oropharynx is clear.

Not all patients had signs in the chest or abnormal radiographs at presentation, but important symptoms and signs leading to a clinical suspicion of pneumonitis were shortness of breath, cough, fever, and reduction in gas transfer. In most of these patients it was still possible to recover a significant pathogen from the lower respiratory tract. A normal chest

radiograph should not be a deterrent to investigation as those patients who are diagnosed and treated early will have a greater chance of recovery. We found that the prognosis for patients presenting with widespread diffuse changes and an arterial oxygen tension of <8.0 kPa while breathing air was extremely poor.

Overall survival in this group of patients was 33%, a figure that compares favourably with results from other centres. Meyers *et al*⁵ reported a mortality rate varying from 63% for idiopathic pneumonia to 91% for cytomegalovirus pneumonia—that is, a survival rate under 10% among patients with a first episode of cytomegalovirus pneumonitis. In our series four out of 12 patients recovered from cytomegalovirus pneumonitis (33%). This figure represents a threefold increase over the survival figures of Meyer *et al*, which is particularly notable since three of these patients survived two episodes. Survival after all episodes of pneumonitis is 50% in our series. Unfortunately, some patients developed a second episode almost immediately after appearing to recover from the first and subsequently died, but others developed a second episode only after a considerable interval, and for these patients recovery from a first episode represented an important gain. One patient was able to live a normal life for two years three months between episodes. Other studies of recipients of bone marrow transplants do not give details of second episodes on pneumonitis.

In conclusion, we would recommend bronchoalveolar lavage as a first line investigation in all recipients of bone marrow transplants developing evidence of pneumonitis.

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References

- Buckner CD, Meyers JD, Springmeyer SC, *et al*. Pulmonary complications of marrow transplantation: review of the Seattle experience. *Exp Haematol* 1984;**12**(suppl 15):1–5.
- Meyers JD. Viral infections in marrow transplant recipients. In: Gale RP, ed. *Recent advances in bone marrow transplantation*. New York: Alan R Liss, 1986:214.
- Cardozo BL, Hagenbeek A. Interstitial pneumonitis following bone marrow transplantation: pathogenesis and therapeutic considerations. *Eur J Cancer Clin Oncol* 1985;**21**:43–51.
- Neiman PE, Reeves W, Ray G, *et al*. A prospective analysis of interstitial pneumonia and opportunistic viral infection among recipients of allogeneic bone marrow grafts. *J Infect Dis* 1977;**136**:754–67.
- Meyers JD, Fluornoy N, Thomas ED. Non bacterial pneumonia after allogeneic marrow transplantation: a review of ten years experience. *Rev Infect Dis* 1982;**4**:1119–32.
- Daniele RP, Elias JA, Epstein PE, Rossman MD. Bronchoalveolar lavage: role in the pathogenesis, diagnosis and management of interstitial lung disease. *Ann Intern Med* 1985;**102**:92–108.
- Stover DE, Saman MB, Hajdu SI, Lange M, Gold J, Armstrong D. Bronchoalveolar lavage in the diagnosis of diffuse pulmonary infiltrates in the immunosuppressed host. *Ann Intern Med* 1984;**101**:1–7.
- Kelley J, Landis JN, Davis GS, Trainer TD, Jakab GJ, Green GM. Diagnosis of pneumonia due to pneumocystis by subsegmental pulmonary lavage via the fiberoptic bronchoscope. *Chest* 1978;**74**:24–8.
- Hopkin JM, Young JA, Turner JH, Abu D, Michael J. Rapid diagnosis of obscure pneumonia in immunosuppressed renal patients by cytology of alveolar lavage fluid. *Lancet* 1983;ii:299–301.
- Stover DE, White DA, Romano PA, Gellene RA. Diagnosis of pulmonary disease in acquired immune deficiency syndrome (AIDS): role of bronchoscopy and bronchoalveolar lavage. *Am Rev Respir Dis* 1984;**130**:659–62.
- Broadus C, Dake MD, Stullberg MS, *et al*. Bronchoalveolar lavage and transbronchial biopsy for the diagnosis of pulmonary infections in the acquired immunodeficiency syndrome. *Ann Intern Med* 1985;**102**:747–52.
- Griffiths PG, Panjwani DD, Stirk PD, *et al*. Rapid diagnosis of cytomegalovirus infection in immunocompromised patients by detection of early antigen fluorescent foci. *Lancet* 1984;ii:1242–5.
- Sloane JP, Depledge MH, Powles RL, Morgenstern GR, Trickey BS, Dady PJ. Histopathology of the lung after bone marrow transplantation. *J Clin Pathol* 1983;**36**:546–54.
- Winston DJ, Gale RP, Meyer DV, Young LS. Infectious complications of human bone marrow transplantation. *Medicine (Baltimore)* 1979;**58**:1–31.
- Prentice HG, Blacklock HA, Janossy G, *et al*. Depletion of T lymphocytes in donor marrow prevents significant graft-versus-host disease in matched allogeneic leukaemic marrow transplant recipients. *Lancet* 1984;i:472–6.
- Blacklock HA, Griffiths PG, Stirk P, Prentice HG. Specific hyperimmune globulin for cytomegalovirus pneumonitis (letter). *Lancet* 1985;ii:152–3.
- Shepp DH, Dandliker PS, De Miranda P, *et al*. Activity of 9-[2-hydroxy-1-(hydroxymethyl)ethoxymethyl] guanine in the treatment of cytomegalovirus pneumonia. *Ann Intern Med* 1985;**103**:368–73.
- Puksa S, Hutcheon MA, Hyland RH. Usefulness of transbronchial biopsy in immunosuppressed patients with pulmonary infiltrates. *Thorax* 1983;**38**:146–50.
- Leight GS, Michaelis LL. Open lung biopsy for the diagnosis of acute, diffuse pulmonary infiltrates in the immunosuppressed patient. *Chest* 1978;**73**:477–82.
- Cunningham JH, Savala DC, Corry RJ, Keim LW. Trephine air drill, bronchial brush, and fiberoptic transbronchial lung biopsies in immunosuppressed

- patients. *Am Rev Respir Dis* 1977;**115**:213–20.
- 21 Davis GS, Kelley J. Invasive techniques for the diagnosis of respiratory infections. *Clinics Lab Med* 1982;**2**: 269–83.
 - 22 Coleman DL, Dodek PM, Luce JM, Golden JA, Gold WM, Murray JF. Diagnostic utility of fiberoptic bronchoscopy in patients with pneumocystis carinii pneumonia and the acquired immune deficiency syndrome. *Am Rev Respir Dis* 1983;**128**:795–9.
 - 23 Kalinske RW, Parker RH, Brandt D, Hoeprick PD. Diagnostic usefulness and safety of transtracheal aspiration. *N Engl J Med* 1967;**276**:604–8.
 - 24 Spencer DC, Beaty HN. Complications of transtracheal aspiration. *N Engl J Med* 1972;**286**:304–5.
 - 25 Davidson M, Tempest B, Palmer DL. Bacteriologic diagnosis of acute pneumonia: comparison of sputum, transtracheal aspiration and lung aspirates. *JAMA* 1976;**235**:158–63.
 - 26 Anderson HA, Fontana RS. Transbronchoscopic lung biopsy for diffuse pulmonary diseases: technique and results in 450 cases. *Chest* 1972;**62**:125–8.
 - 27 Flick MR, Wasson K, Dunn LJK, Block AJ. Fatal pulmonary haemorrhage after transbronchial biopsy through the fiberoptic bronchoscope. *Am Rev Respir Dis* 1975;**111**:853–6.
 - 28 Zavala DC. Pulmonary haemorrhage in fiberoptic transbronchial biopsy. *Chest* 1976;**70**:584–8.
 - 29 Pearce JG, Patt NL. Fatal pulmonary haemorrhage after percutaneous aspiration lung biopsy. *Am Rev Respir Dis* 1974;**110**:345–9.
 - 30 Wallace JM, Batra P, Gone H jun, Ovenfors CO. Percutaneous needle aspiration for diagnosing pneumonitis in the patient with acquired immunodeficiency syndrome (AIDS). *Am Rev Respir Dis* 1985;**131**: 389–92.
 - 31 Williams D, Yungbluth M, Adams G, *et al.* The role of fiberoptic bronchoscopy in the evaluation of immunocompromised hosts with diffuse pulmonary infiltrates. *Am Rev Respir Dis* 1985;**131**:880–5.