Respiratory muscle and pulmonary function in polymyositis and other proximal myopathies

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ABSTRACT We studied 53 patients with proximal myopathy to determine at what level of muscle weakness hypercapnic respiratory failure is likely, and which tests of pulmonary function or respiratory muscle strength would best suggest this development. Respiratory muscle strength was determined from maximal static efforts and in half the patients, both inspiratory and expiratory muscle strengths were less than 50% of normal. In the 37 patients without lung disease respiratory muscle weakness was accompanied by significant decreases in vital capacity, total lung capacity, and maximum voluntary ventilation; by significant increases in residual volume and arterial carbon dioxide tension (Paco₂); and greater likelihood of dependence on ventilators, atelectasis, and pneumonia. Hypercapnia was particularly likely when respiratory muscle strength was less than 30% of normal in uncomplicated myopathy, and when vital capacity was less than 55% of the predicted value in any patient.

Myopathies which affect proximal limb muscles may also affect respiratory muscles; however, the relationships between the extent to which respiratory muscles are affected, abnormality of pulmonary function, and the onset of respiratory failure have not been defined. We addressed these questions in a group of 53 patients with various proximal myopathies.

Methods

The patients studied were adults with myopathy from the neurology and medicine services of Harlem Hospital Center and Columbia Presbyterian Medical Center in New York and the University of Virginia Hospital in Charlottesville. They were referred for evaluation of pulmonary or respiratory muscle function or both. They represent about one-third of all patients with a diagnosis of myopathy at these hospitals during the period of the study.

Myopathy was diagnosed by conventional criteria (table 1). Patients with neoplasm, diabetes mellitus, thyroid or parathyroid disease, chronic renal insufficiency, sarcoidosis, and granulomatous lung disease were excluded from the study.

Vital statistics are summarised in table 2. Patients with polymyositis or dermatomyositis (group I) had had symptoms for three months to two years; the symptoms of patients with other myopathies (group II) had lasted from several months to many years. Thirty-seven patients with no evidence of chronic lung disease (groups IA, IIA) were considered separately from the 16 patients with myopathy and

<table>
<thead>
<tr>
<th>Diagnosis</th>
<th>No</th>
<th>%</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total patients with polymyositis</td>
<td>31</td>
<td>58.5</td>
</tr>
<tr>
<td>Polymyositis alone</td>
<td>26</td>
<td>49.1</td>
</tr>
<tr>
<td>Polymyositis with dermatomyositis</td>
<td>3</td>
<td>5.7</td>
</tr>
<tr>
<td>Polymyositis with mixed connective tissue</td>
<td>2</td>
<td>3.8</td>
</tr>
<tr>
<td>disease</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total patients with chronic myopathies</td>
<td>22</td>
<td>41.5</td>
</tr>
<tr>
<td>Acid maltase deficiency</td>
<td>6</td>
<td>11.3</td>
</tr>
<tr>
<td>Idiopathic myopathy</td>
<td>6</td>
<td>11.3</td>
</tr>
<tr>
<td>Alcoholic myopathy</td>
<td>4</td>
<td>7.5</td>
</tr>
<tr>
<td>Carnitine deficiency</td>
<td>2</td>
<td>3.7</td>
</tr>
<tr>
<td>Glycogen-debrancher enzyme deficiency</td>
<td>1</td>
<td>1.9</td>
</tr>
<tr>
<td>Ragged red fibre disease</td>
<td>1</td>
<td>1.9</td>
</tr>
<tr>
<td>Lipid storage myopathy</td>
<td>1</td>
<td>1.9</td>
</tr>
<tr>
<td>Steroid myopathy</td>
<td>1</td>
<td>1.9</td>
</tr>
</tbody>
</table>

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chronic obstructive or interstitial lung disease (groups IB, IIB). Criteria for a diagnosis of obstructive lung disease included a history of cigarette smoking, chronic cough and sputum production, and a ratio of FEV₁ to vital capacity of less than 70%. The diagnosis of interstitial lung disease was made from an interstitial pattern on the chest radiograph; this was present in five patients with polymyositis and one with alcoholic myopathy.

Spirometric volumes were measured by either a 13.5-litre water-filled spirometer with large-bore tubing (Warren E Collins, Braintree, Montana) or an Automated Pulmonary Functional Lab Mark I (System Research Laboratories Inc, Dayton, Ohio). Measurements included slow and forced vital capacity (VC, FVC), FEV₁, and the ratio FEV₁/FVC. The 12-second maximum voluntary ventilation (MVV) was measured with the water-filled spirometer in 40 patients. Functional residual capacity (FRC) was determined by helium dilution in 22 patients. Total lung capacity (TLC) was calculated as the sum of FRC and inspiratory capacity. In three additional patients, TLC was calculated using posteroanterior and lateral chest radiographs taken at full inspiration, a modification of the Barnhard ellipse method being used. Residual volume (RV) was calculated as the difference between TLC and VC. Vital and total lung capacities were expressed as percentage of predicted normal values (VC%, TLC%), and RV was expressed as percentage of predicted TLC (RV%).

Maximum static inspiratory and expiratory pressures at the mouth were measured in 47 patients by the technique of Black and Hyatt. Maximum inspiratory pressure (Pi-max) was measured at RV and maximum expiratory pressure (Pe-max) at TLC. These values were expressed as percentage of the predicted normal (Pi% and Pe%) for persons of the same sex and age group (table 3). Respiratory muscle strength (RMS%) was calculated as the average of Pi% and Pe%. The reported values of spirometric data and respiratory muscle pressures were the best of three efforts.

In the presence of respiratory muscle weakness the measured values of Pe-max and Pi-max may be affected by lung volume as well as by myopathy. If inspiratory muscle weakness prevents the patients from achieving a normal TLC, then Pe-max will be measured at a lung volume where the expiratory muscles are contracting from a resting length less than optimum. With expiratory muscle weakness the maximum inspiratory manoeuvre may be performed at a lung volume where the inspiratory muscles contract from a shorter-than-optimal length, reducing the observed Pi-max. To correct for these effects of lung volume on Pi-max and Pe-max, we used Ringqvist's regression equations predicting Pi% and Pe% from lung volume expressed as percentage of predicted TLC.²⁰

Arterial blood gas composition was determined in 45 patients, each patient having at least two samples taken. In eight patients end-tidal carbon dioxide tension (Pco₂) was measured with a carbon dioxide analyser (Mark I Capnograph, Godart) to estimate arterial carbon dioxide tension (Paco₂). The alveolar-arterial oxygen tension gradient (AaPo₂) was calculated from the alveolar air equation, a respiratory quotient of 0.8 being assumed.

Statistical analyses included χ² tests with Yates's correction, t tests for differences between two group means, and linear regression analysis. Data in text and tables are reported as means and standard deviations (SD).

Results

Normal values for Pi-max and Pe-max in men and women aged 14–49 and 50–70 years were obtained at the University of Virginia Hospital (table 3) and are similar to those reported elsewhere.⁹ ¹⁰ The largest coefficient of variation in table 3 is 30%, so that any value of Pi-max or Pe-max lower than 70% of the mean is more than one standard deviation below the mean normal value.

In the 47 subjects in whom measurements of maximum pressures were made Pi-max averaged 48 (19) cm H₂O and Pe-max 80 (41) cm H₂O, both significantly below normal (p < 0.001). Overall respiratory muscle strength (RMS) was less than 70% of normal (moderate weakness) in 37 patients and less than 50% of normal (severe weakness) in 24 of the 47 patients. The prevalence of moderate and severe respiratory muscle weakness was not significantly different in patients with polymyositis or dermatomyositis (group I) from that in patients with congenital and miscellaneous myopathies (group II). The ratio Pi-max/Pe-max averaged 0.57 (0.20), which is not significantly different from the normal values of 0.58 (0.12) for men and 0.65 (0.14) for women (table 3). This indicates that loss of strength was distributed evenly among inspiratory and expiratory muscles.

### Table 2 Mean (SD) age and height and sex distribution of the four groups of patients

<table>
<thead>
<tr>
<th>Groups*</th>
<th>IA</th>
<th>IIA</th>
<th>IB</th>
<th>IIB</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (y)</td>
<td>41 (15)</td>
<td>40 (13)</td>
<td>54 (15)</td>
<td>41 (14)</td>
</tr>
<tr>
<td>Height (cm)</td>
<td>164 (8)</td>
<td>174 (12)</td>
<td>174 (11)</td>
<td>177 (6)</td>
</tr>
<tr>
<td>Sex</td>
<td>5M</td>
<td>10M</td>
<td>7M</td>
<td>6M</td>
</tr>
<tr>
<td></td>
<td>15F</td>
<td>6F</td>
<td>3F</td>
<td></td>
</tr>
</tbody>
</table>

*Group I comprises patients with polymyositis or dermatomyositis and group II patients with other proximal myopathies; groups IA and IIA have no chronic lung disease and IB and IIB have interstitial or obstructive lung disease.

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Table 3  Normal mean (SD) values of maximum static inspiratory and expiratory pressures measured at the mouth (Pimax, Pemin) and the ratio Pimax/Pemin (coefficient of variation (%) in square brackets)

<table>
<thead>
<tr>
<th></th>
<th>Males</th>
<th>Females</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (y)</td>
<td>14-49</td>
<td>14-49</td>
</tr>
<tr>
<td>Number</td>
<td>80</td>
<td>27</td>
</tr>
<tr>
<td>Pimax (cm H2O)</td>
<td>127 (28) [22]</td>
<td>91 (25) [27]</td>
</tr>
<tr>
<td>Pemin (cm H2O)</td>
<td>216 (45) [22]</td>
<td>138 (39) [28]</td>
</tr>
<tr>
<td>Pimax/Pemin</td>
<td>0.59 (0-12)</td>
<td>0.67 (0-16)</td>
</tr>
</tbody>
</table>

When Pimax and Pemin were plotted against each other, the regression line was not significantly different from the line of identity (fig 1).

Total lung capacity (TLC) and residual volume (RV) were measured in 22 of the patients whose Pimax and Pemin were measured. The mean corrections for the effect of lung volume and Pimax and Pemin were 9% (3%) and 12% (4%) of the observed value respectively. As a result, for this subset of patients Pimax increased from 49% (28%) predicted to 54% (32%) predicted, and Pemin increased from 45% (22%) to 51% (25%) predicted. The distribution of weakness between inspiratory and expiratory muscles was not significantly affected by the lung volume corrections.

The mean values for respiratory muscle strength, pulmonary function results, and arterial blood gas composition in each of the groups are given in table 4. All patients with myopathy uncomplicated by chronic lung disease (groups IA and IIA) had normal values of FEV1/VC, and there were significant reductions from normal in Pimax, Pemin, RMS%, TLC%, VC%, and MVV%; AaPo2 was greater than normal. The reductions in Pimax, Pemin, RMS%, and MVV% were greater in group IIA, which includes patients with congenital myopathies, than in group IA (patients with polymyositis or dermatomyositis) (p < 0.05).

To assess the effect of coexistent chronic lung disease we compared groups IA and IIA with groups IB and IIB. The patients with chronic lung disease (groups IB, IIB) had significantly lower values of MVV% and arterial oxygen tension (Pao2) and higher values of AaPo2 than did the patients without chronic lung disease (groups IA, IIA) (p < 0.05). There were no significant differences between those with and without chronic lung disease with respect to Pimax, Pemin, RMS%, VC%, or Paco2.

Six of the patients with myopathy had interstitial lung disease; their FEV1/VC averaged 84% (5%), not significantly different from the values in patients without chronic lung disease. As expected, the 10 patients with obstructive lung disease had an FEV1/VC averaging 54% (15%). Otherwise, there were no differences between patients with interstitial and obstructive disease with respect to Pimax, Pemin, RMS%, TLC%, MVV%, arterial blood gas composition, or AaPo2.

To illustrate the relationships between respiratory muscle weakness, pulmonary function, and gas exchange, we plotted several ventilatory and blood gas variables as functions of respiratory muscle strength (figs 2–5). In these figures regression lines in the figures represent data from groups IA and IIA only.

Residual volume (RV%) was inversely related to Pemin (fig 2). The values from groups IB and IIB were within the 95% confidence limits of the regression based on groups IA and IIA. There were no correlations between TLC% and Pemin, Pimax, or RMS%.

Vital capacity (VC%) showed a significant linear correlation with RMS%: VC% = 0.523 RMS% ± 38.3 (N = 33, r = 0.642, p < 0.001), but the relationship appears curvilinear. Hence we used a log-log relationship and found that the correlation

![Fig 1  Relationship between maximum static inspiratory and expiratory pressures (Pimax, Pemin) in 33 patients with myopathy uncomplicated by chronic lung disease (closed circles, regression line) and 14 with myopathy plus chronic interstitial or obstructive lung disease (open circles). The solid regression line (r = 0.841, p < 0.001) is not significantly different from the dashed line of identity.](http://thorax.bmj.com/)

Fig 1  Relationship between maximum static inspiratory and expiratory pressures (Pimax, Pemin) in 33 patients with myopathy uncomplicated by chronic lung disease (closed circles, regression line) and 14 with myopathy plus chronic interstitial or obstructive lung disease (open circles). The solid regression line (r = 0.841, p < 0.001) is not significantly different from the dashed line of identity.
Respiratory muscle and pulmonary function in polymyositis and other proximal myopathies

Table 4  Mean (SD) maximum respiratory pressures, lung volumes, ventilatory capacity, and arterial blood gas values in the four groups of patients (number of observations for each test in square brackets)

<table>
<thead>
<tr>
<th>Group</th>
<th>IA</th>
<th>IIA</th>
<th>IB</th>
<th>IIB</th>
</tr>
</thead>
<tbody>
<tr>
<td>(P_{\text{max}}) (% predicted)</td>
<td>60 (22) [18]</td>
<td>40 (24) [15]</td>
<td>44 (12) [8]</td>
<td>39 (9) [6]</td>
</tr>
<tr>
<td>(P_{\text{e}_{\text{max}}}) (% predicted)</td>
<td>57 (24) [18]</td>
<td>42 (21) [15]</td>
<td>51 (21) [8]</td>
<td>46 (18) [6]</td>
</tr>
<tr>
<td>(\text{Paco}_2) (mmHg)</td>
<td>40 (8) [21]</td>
<td>44 (9) [16]</td>
<td>43 (9) [10]</td>
<td>42 (9) [6]</td>
</tr>
<tr>
<td>(\text{Pao}_2) (mmHg)</td>
<td>77 (14) [15]</td>
<td>79 (13) [14]</td>
<td>65 (13) [9]</td>
<td>73 (9) [5]</td>
</tr>
<tr>
<td>AaPo2 (mmHg)</td>
<td>22 (9) [15]</td>
<td>16 (11) [14]</td>
<td>31 (17) [9]</td>
<td>23 (10) [5]</td>
</tr>
</tbody>
</table>

*See footnote to table 2 for definition of groups.

\(P_{\text{max}}, P_{\text{e}_{\text{max}}}\) — maximum static inspiratory and expiratory pressure; RMS — respiratory muscle strength; TLC — total lung capacity; RV — residual volume; VC — vital capacity; FEV1 — one-second forced expiratory volume; MVV — maximum voluntary ventilation; \(\text{Paco}_2\), \(\text{Pao}_2\) — arterial carbon dioxide and oxygen tensions; AaPo2 — alveolar-arterial oxygen tension gradient.

Conversion: Traditional to SI units — Blood gases: 1 mmHg = 0.133 kPa.

coefficient and the explained variance in VC% were higher (fig 3). Only three patients from groups IB and IIB had values of VC% which lay below the 95% confidence limits of the regression. Maximum voluntary ventilation (MVV%) was linearly related to RMS% (fig 4). Six patients from groups IB and IIB had values of MVV% which lay below the 95% confidence limits of the regression calculated from group IA and IIA data. The group mean values (table 4) obscure any relationship between carbon dioxide retention and respiratory muscle weakness. When data from the 33 patients without lung disease were plotted, however, \(\text{Paco}_2\) appeared to be a discontinuous function of RMS% (fig 5). Below an RMS value of 50% predicted \(\text{Paco}_2\) increased linearly as RMS fell (\(p < 0.01\)). When RMS exceeded 50% of the predicted value \(\text{Paco}_2\) was normal and independent of RMS%. Five of the 14 patients in groups IB and IIB had \(\text{Paco}_2\) values that exceeded the 95% confidence limits of the regressions calculated from groups I A and IIA data. The results were virtually identical when \(P_{\text{max}}\) and \(P_{\text{e}_{\text{max}}}\) were substituted for RMS%.

In contrast to the relationship between \(\text{Paco}_2\) and RMS%, the relationship between \(\text{Paco}_2\) and \(\text{VC}\) appeared to be linear throughout the range of VC%.

Fig 2  Relationship between residual volume (RV) and expiratory pressure (\(P_{\text{e}_{\text{max}}}\)) in 15 patients with uncomplicated myopathy (closed circles, regression line) and seven with myopathy plus chronic lung disease (open circles). The equation of the line is: RV% = 48.9 - 0.361 \(P_{\text{e}_{\text{max}}}\%\) (\(r = -0.651\), \(p < 0.01\)).

Fig 3  Relationship between vital capacity (VC) and respiratory muscle strength (RMS) in 33 patients with uncomplicated myopathy (closed circles, regression line) and 17 with myopathy plus chronic lung disease (open circles). The equation of the line is: \(\log \text{VC}\% = 1.051 + 0.499 \log \text{RMS}\%\) (SEE 0.089, \(r = 0.794\), \(p < 0.001\)).
Fig 4  Relationship between maximum voluntary ventilation (MVV) and respiratory muscle strength (RMS) in 26 patients with uncomplicated myopathy (closed circles, regression line) and 12 with myopathy plus chronic lung disease (open circles). The equation of the line is: MVV% = 33.2 + 0.574 RMS% (r = 0.708, p < 0.001).

Fig 5 Relationship between arterial carbon dioxide tension (Paco₂) (mm Hg) and respiratory muscle strength (RMS) (%) in 33 patients with uncomplicated myopathy (closed circles, regression line) and 14 patients with myopathy plus chronic lung disease (open circles). The equation of the line when RMS <50% predicted is: Paco₂ = 55 - 0.37 RMS (r = -0.587, p < 0.02). When RMS >50% predicted the equation is: Paco₂ = 40 - 0.045 RMS (NS). Conversion: Traditional to SI units—Paco₂: 1 mm Hg = 0.133 kPa.

in patients from groups IA and IIA (fig 6). The plot includes three patients with carbon dioxide retention and low values of VC in whom RMS was not measured. The data points from groups IB and IIB patients lay within the 95% confidence limits of the regression calculated from the groups IA and IIA data.

Serious complications potentially attributable to respiratory, laryngeal, or pharyngeal muscle weakness occurred in 20 of the 53 patients with myopathy. There were five deaths; seven patients remain dependent on ventilators; and 10 have had recurrent aspiration pneumonia and atelectasis (one of the pneumonia group was also ventilator dependent). For the entire group there was no relationship between muscle weakness and the development of complications. In groups IA and IIA, however, nine of the 17 (53%) with RMS less than 50% of predicted had complications, but only one of the 16 patients (6%) whose RMS exceeded 50% of predicted (p < 0.02). In two patients with polymyositis the severity of the respiratory muscle weakness was not initially appreciated, and respiratory muscle strength was not measured. One of these died of acute and the other of chronic hypercapnic respiratory failure.

Discussion

The clinical and physiological expressions of myopathy in these patients were similar to those in patients described by others. No previous study, however, has related the development of hypercapnia to the degree of respiratory muscle dysfunction. Congenital myopathies and polymyositis are well known to affect the respiratory muscles, and corticosteroid treatment has been implicated as a
Respiratory muscle and pulmonary function in polymyositis and other proximal myopathies

cause of respiratory muscle weakness in obese asthmatic patients. We also found respiratory muscle weakness in alcoholic myopathy (table 1).

Since lung disease might be expected to affect adversely the respiratory muscles, we included 16 patients with chronic pulmonary disease; our patients with polymyositis and interstitial lung disease had values of total lung capacity, vital capacity, and PaO₂ that were virtually identical to those reported by Salmeron et al. Ten others had chronic obstructive lung disease not related to the myopathy. The functional abnormalities were greater in the patients with additional obstructive or interstitial lung disease; the influence of lung disease was, however, small and this may have been related to the fact that airflow obstruction was mild or only moderately severe in the patients studied.

Thirty-eight percent of our patients had complications attributable to muscle weakness, including carbon dioxide retention and ventilator dependence; there were five deaths. These complications were significantly related to respiratory muscle weakness.

We found that the deficit in respiratory muscle strength was evenly distributed between inspiratory and expiratory muscles in patients with myopathy (fig 1). This conclusion was not altered when PiO₂max and PeO₂max were corrected for lung volume, as has also been shown for the distribution of muscle weakness in undernourished patients. In contrast, patients with amyotrophic lateral sclerosis, Duchenne's dystrophy, myotonic dystrophy, myasthenia gravis, and curarised normal volunteers have more weakness of the expiratory muscles than of the inspiratory muscles. The reason for the even distribution of weakness in myopathy and undernutrition and the predominance of expiratory muscle weakness in the other diseases is not known.

The increase in residual volume with progressive expiratory muscle weakness (fig 2) resulted from inability of weakened expiratory muscles to force thoracic volume below functional residual capacity (FRC), which is the resting volume of the respiratory system. Kreitzer et al found a similar relationship between residual volume and expiratory muscle weakness with amyotrophic lateral sclerosis.

Both total lung capacity and vital capacity are determined by the balance between respiratory system recoil and the force generated by contraction of the respiratory muscles. Above FRC most of the respiratory system recoil is contributed by the lungs. Respiratory system recoil could be altered because of changes in lung or chest wall compliance. A reduction in lung compliance has been found but its mechanism is not entirely clear. One reason may be that alterations in the configuration of surfactant and microatelectasis attend low tidal volumes. The increase in AaPo₂ in our patients of groups IA and IIA (table 4) is consistent with this hypothesis. A further reason may be that inspiratory muscle weakness results in the inability to develop a large inspiration volume history, which reduces the slope of the pulmonary volume-pressure curve. Thus we would expect that the weaker the inspiratory muscles the lower would be TLC and VC. In fact, for TLC no such relation was observed. Others have found TLC to be normal or reduced in patients with muscle weakness of various causes.

The relationship between vital capacity and respiratory muscle strength is curvilinear. Because the static recoil curve is sigmoid, reducing respiratory strength to 50% of normal would theoretically reduce VC by only about 15%, whereas further muscle weakness should produce a greater reduction in VC. Our results (fig 3) generally agree with those of others in that VC was lower than predicted from theoretical considerations. The discrepancy between the theoretical and observed behaviour of the vital capacity as a function of respiratory muscle strength could be explained by alterations either in the force-length relation of the weakened muscles or in the static recoil curve of the respiratory system. To our knowledge, the force-length curve of myopathic muscles has not been characterised.

In curarised normal volunteers, however, trans-diaphragmatic pressure during maximum static inspiratory efforts was reduced proportionately at all lung volumes. This suggests that, while force at any given diaphragm muscle length was reduced, the intrinsic shape of the force-length curve was preserved. Thus the disproportionate reduction of vital capacity in neuromuscular disease is probably not explained by abnormal muscle mechanics alone.

The magnitude of the maximum voluntary ventilation has been shown to be a function of respiratory muscle strength in normal subjects, undernourished patients, patients with obstructive and interstitial lung diseases, and patients with other neuromuscular diseases, and now in patients with myopathy (fig 4). In contrast to the relationships between RV or VC and RMS, the MVV–RMS relation was relatively sensitive to the presence of coexistent lung disease (fig 4). The dependence of MVV on respiratory muscle strength is attributable to the relationship between pressure and airflow. In maximum inspiratory efforts airflow depends on pressure throughout inspiration, but in maximal expiratory efforts airflow is pressure dependent only in early expiration. During maximum voluntary ventilation both inspiratory and expiratory muscle pressures contribute to airflow and minute volume. Because the effort is sustained for only 12–15 seconds, the strength of the respiratory muscles, rather than their endurance
characteristics, appears to be of paramount importance in determining MVV.

Respiratory failure with carbon dioxide retention is a serious complication of generalised myopathy. Nine of our patients without lung disease had hypercapnia; this was clearly related to respiratory muscle weakness in six (fig 5), and undoubtedly attributable to respiratory muscle weakness in two of the other three, whose VC was less than 20% of predicted. There was a discontinuous relationship between Paco₂ and respiratory muscle strength (fig 5), such that hypercapnia did not appear until RMS was less than 40% of normal, and it was not severe until RMS had dropped to 30% of normal.

The vital capacity has long been used to reflect the severity of respiratory muscle weakness and to predict respiratory failure in neuromuscular diseases and an inverse relationship between arterial carbon dioxide tension and vital capacity has been noted in previous studies. We found a similar relationship (fig 6) but severe hypercapnia was present in some patients with VC as high as 55% of predicted. The difference in the shape of the Paco₂-RMS and Paco₂-VC relationships reflects the curvilinear nature of the VC-RMS relationship (fig 3), but the advantage of relating Paco₂ to RMS instead of to VC is that the Paco₂-RMS graph more readily identifies the patients in whom respiratory muscle weakness is the primary cause of hypercapnia (fig 5).

In several reported cases of neuromuscular disease hypercapnia appears to have resulted primarily from reduced ventilatory drive, even when respiratory muscle weakness was minimal. Ventilatory drive, however, is usually well preserved in neuromuscular disease, as judged from normal mouth occlusion pressures despite substantial respiratory muscles weakness. We have found the same to be true in four of our patients with acid maltase deficiency, two of whom had RMS values of 7% and 14% of predicted, and Paco₂ of 61 and 49 mm Hg (8.1 and 6.5 kPa).

We conclude that respiratory muscle weakness alone is sufficient to account for hypercapnia when respiratory muscle strength is less than 30% of normal. Vital capacity is a useful index, and a reduction below 55% of predicted in patients with neuromuscular diseases is likely to be associated with hypercapnia. If, however, carbon dioxide retention is found with a VC above this level or with RMS greater than 30% normal, coexistent lung disease or abnormality of ventilatory control, or both, are probably also contributory.

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