Benign sclerosing pneumocytoma of lung (sclerosing haemangioma)

KEENG-WAI CHAN,* ALLEN R GIBBS,† WS LO,‡ GR NEWMAN**

From the *Kowloon Institute of Pathology, Kowloon Hospital, Hong Kong; †Department of Pathology, Welsh National School of Medicine, Cardiff; ‡Kowloon Hospital, Hong Kong; and **Department of Pathology, University Hospital of Wales, Cardiff

ABSTRACT The clinical and histopathological features of 14 cases of so-called sclerosing haemangioma of the lung are described. All developed in Hong Kong Chinese women. Histochemical and ultrastructural study of these tumours indicates an epithelial origin. The term benign sclerosing pneumocytoma is suggested as being more appropriate for this unusual tumour.

Liebow and Hubbell in 1956 described seven examples of a rare, benign pulmonary tumour under the term sclerosing haemangioma.1 Patients presented with haemoptysis or with a coin lesion. The tumour showed a striking preponderance in women and was characterised microscopically by “vascular channels,” sclerosis, papillae, sheets of cells, and haemorrhage in varying proportions. Considerable diagnostic confusion has occurred, however, since some of the reports have included plasma cell granuloma and histiocytic lesions inappropriately under the term sclerosing haemangioma and the reverse has also occurred.2-9 We review the clinicopathological features of 14 “sclerosing haemangiomas” that comply fully with the criteria of Liebow and Hubbell. This is the second largest series to be reported.10-18 We also report the electron microscopic and histochemical findings for several of these tumours. On the basis of these findings we proffer the term benign sclerosing pneumocytoma as being more appropriate for this unusual tumour.

Methods

The 14 cases of “sclerosing haemangioma” studied were taken from the file of the Institute of Pathology, Hong Kong, from 1974 to 1980. Ten of these cases came from the Cardiothoracic Unit, Kowloon Hospital, including four that presented in 1980. The five most recent cases were also examined by electron microscopy from formalin-fixed material.

All specimens were fixed in 10% buffered formalin. One to six blocks were taken from each tumour for paraffin embedding. Sections were cut and stained with haematoxylin and eosin, Victoria blue, Van Gieson’s, Gordon and Sweet’s reticulin method, Alcian blue/Unna Pappenheim technique (AB/UP), Periodic Acid-Schiff method with (PAS-D) and without (PAS) diastase digestion, and Perl’s method. Frozen sections were cut and stained with oil red O.

Case 13 was submitted fresh and frozen sections were cut for enzyme studies—acid phosphatase by Barka’s modified technique,19 non-specific esterase by the method of Davis and Ornstein,19 and alkaline phosphatase by Gomori’s method.20 Tissue, fixed for periods over two weeks in buffered formalin, was refixed in 2-8% glutaraldehyde in a 0-1 mol phosphate buffer at pH 7-4. After osmication in 2% veronal acetate buffered osmium tetroxide the tissue was treated en bloc with 3% aqueous uranyl acetate dehydrated and embedded in Epon 812. Thin sections were stained on the grid in lead citrate.

Results

CLINICAL FEATURES (table 1)

All cases occurred in females; the age ranged from 16 to 78 years (average 50-8, median 55 years). Seven were symptom free and the lesion was discovered by routine chest radiography; cough was present in five patients; four had haemoptysis; one had ill-defined chest pain. The size of the tumour varied from 13 mm to 35 mm in diameter (average 23-3 mm). Most of the tumours were located beneath the pleura, sometimes bulging out as a sessile polypoid growth. In some cases the lesions were
Benign sclerosing pneumocytoma of lung

Table 1  Clinical summary of 14 cases (all in women) of “sclerosing haemangiomas” of lung

<table>
<thead>
<tr>
<th>Case</th>
<th>Age</th>
<th>Presenting symptoms</th>
<th>Clinical diagnosis</th>
<th>Anatomical location</th>
<th>Diameter (mm)</th>
<th>Operation</th>
<th>Follow-up Nov 1980</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>56</td>
<td>None</td>
<td>Tuberculoma, haemangioma</td>
<td>Left lung, oblique fissure</td>
<td>23</td>
<td>Excision, 1974</td>
<td>—</td>
</tr>
<tr>
<td>2</td>
<td>55</td>
<td>None</td>
<td>Carcinoma</td>
<td>RML</td>
<td>30</td>
<td>RM lobectomy, 1975</td>
<td>—</td>
</tr>
<tr>
<td>3</td>
<td>16</td>
<td>Cough, 6 months</td>
<td>Bronchial adenoma</td>
<td>RLL</td>
<td>13</td>
<td>Excisional biopsy, 1977</td>
<td>—</td>
</tr>
<tr>
<td>4</td>
<td>55</td>
<td>None</td>
<td>Carcinoma</td>
<td>RLL</td>
<td>20</td>
<td>RL lobectomy, 1978</td>
<td>—</td>
</tr>
<tr>
<td>5</td>
<td>58</td>
<td>RL chest pain</td>
<td>Carcinoma</td>
<td>LUL</td>
<td>30</td>
<td>LU lobectomy, 1974</td>
<td>—</td>
</tr>
<tr>
<td>6</td>
<td>46</td>
<td>Haemoptysis</td>
<td>Tuberculoma</td>
<td>RLL</td>
<td>18</td>
<td>Apical segmentectomy, 1976</td>
<td>—</td>
</tr>
<tr>
<td>7</td>
<td>78</td>
<td>None</td>
<td>Carcinoma</td>
<td>LUL</td>
<td>24</td>
<td>LU lobectomy, 1977</td>
<td>Died 1 year later of cerebrovascular accident</td>
</tr>
<tr>
<td>8</td>
<td>47</td>
<td>None</td>
<td>Carcinoma</td>
<td>RML</td>
<td>28</td>
<td>RM lobectomy, 1978</td>
<td>Alive and well</td>
</tr>
<tr>
<td>9</td>
<td>46</td>
<td>Haemoptysis, cough, coin lesions since 1968</td>
<td>Hamartoma</td>
<td>RLL</td>
<td>35</td>
<td>RL lobectomy, 1978</td>
<td>Alive and well</td>
</tr>
<tr>
<td>10</td>
<td>49</td>
<td>Cough and blood-stained sputum</td>
<td>Carcinoma</td>
<td>LUL</td>
<td>22</td>
<td>LU lobectomy, 1979</td>
<td>Alive and well</td>
</tr>
<tr>
<td>11</td>
<td>63</td>
<td>Cough</td>
<td>Tuberculoma</td>
<td>LUL</td>
<td>20</td>
<td>Enucleation, Feb 1980</td>
<td>Alive and well</td>
</tr>
<tr>
<td>12</td>
<td>60</td>
<td>Cough, 6 months (haemoptysis twice 10 and 20 years ago)</td>
<td>Carcinoma</td>
<td>LLL</td>
<td>13</td>
<td>LI lobectomy, May 1980</td>
<td>Alive and well</td>
</tr>
<tr>
<td>13</td>
<td>20</td>
<td>None</td>
<td>Hamartoma</td>
<td>Lingula</td>
<td>22</td>
<td>Enucleation, June 1980</td>
<td>Alive and well</td>
</tr>
<tr>
<td>14</td>
<td>61</td>
<td>None</td>
<td>Carcinoma</td>
<td>RLL</td>
<td>28</td>
<td>RL lobectomy</td>
<td>Alive and well</td>
</tr>
</tbody>
</table>

LUL = left upper lobe  RML = right middle lobe  LLL = left lower lobe  RLL = right lower lobe.

situated near a segmental bronchus (fig 1). In case 1 it was situated over the oblique fissure of the left lung.

Seven of the 10 patients operated on at Kowloon Hospital were interviewed during November 1980 by one of us (WSL) and found to be well. One patient had died of cerebral haemorrhage a year after operation. No follow-up information was available for the two remaining cases or the four cases not operated on at Kowloon Hospital. No patient was known to have shown recurrence or dissemination of the tumour.

PATHOLOGY

Gross appearance

The tumours were well circumscribed, soft to firm, and surrounded by a pseudocapsule. The cut surface usually had a variegated appearance, yellowish to grey with scattered reddish-brown areas. Sometimes necrosis and calcification were present. Occasional tumours were more uniform in appearance and minute cleft-like spaces were sometimes seen (fig 1).

Light-microscope appearance

Four major histological patterns—solid, papillary, angiomatoid, and sclerotic—were encountered in varying proportions within these tumours (table 2). Solid, papillary, and sclerotic areas (figs 2 and 3) were evident in every tumour, although they sometimes had to be searched for. Angiomatoid areas (figs 2 and 4), however, were absent in four of the tumours.

(1) Solid—These areas consisted of sheets and irregular masses of plump fusiform or polygonal cells with moderate amounts of eosinophilic cytoplasm and round, ovoid, or slightly convoluted nuclei with a fine chromatin pattern and small discrete nucleolus. Occasional nuclei were grooved. Mitoses were rare. Sometimes large cells with clear cytoplasm were present. Silver impregnation showed reticulin surrounding groups of cells.

Fig 1  Case 14: centrally located tumour close to a segmental bronchus.
Table 2  Summary of histological features of 14 ‘sclerosing haemangiomatous’

<table>
<thead>
<tr>
<th>Case</th>
<th>Solid</th>
<th>Papillary</th>
<th>Angiomatoid</th>
<th>Sclerosis</th>
<th>Clear cells</th>
<th>Haemorrhage</th>
<th>Haemosiderin</th>
<th>Foam cells</th>
<th>Mast cells</th>
<th>Fat</th>
<th>Glycogen</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>++</td>
<td>+</td>
<td>++</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>(Calcification)</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>2</td>
<td>+</td>
<td>+</td>
<td>++</td>
<td>+</td>
<td>0</td>
<td>++</td>
<td>+</td>
<td>±</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>3</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>0</td>
<td>++</td>
<td>Siderotic nodule</td>
<td>+</td>
<td>±</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>4</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>5</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>0</td>
<td>++</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>6</td>
<td>+</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>Siderotic nodule</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>7</td>
<td>++</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>(Calcification)</td>
<td>Siderotic nodule</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>8</td>
<td>++</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>9</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>(Haematoxin +)</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>10</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>(Haematoxin +)</td>
<td>Siderotic nodule</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>11</td>
<td>±</td>
<td>±</td>
<td>0</td>
<td>±</td>
<td>0</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>12</td>
<td>++</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
<td>(Haematoxin +)</td>
<td>Siderotic nodule</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>13</td>
<td>+</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>±</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>14</td>
<td>+</td>
<td>+</td>
<td>±</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>(Calcification)</td>
<td>Siderotic nodule</td>
<td>+</td>
<td>±</td>
<td>+</td>
</tr>
</tbody>
</table>

0 = absent; + = present; ± = scanty; ++ = predominant or abundant.

(2) Papillary—This pattern consisted of irregular anastomosing, cleft-like spaces with projections of papillary structures covered by a layer of cuboidal to low columnar cells (figs 2 and 3), some with a hobnail appearance (fig 6). The cores were composed of a central capillary surrounded by collagenous fibrous tissue with varying degrees of hyalinisation admixed with variable numbers of rounded tumour cells (figs 2 and 3). Semi-thin sections showed the cells lining the papillae to be a mixture of light and dark cells. Several appeared vacuolated (fig 7). Some of the vacuoles appeared clear and others contained a dense, dark core; these corresponded to lamellar bodies seen on electron microscopy.

(3) Angiomatoid—This pattern consisted of clusters of variable-sized honeycomb spaces containing red blood cells (figs 2 and 4), some without apparent lining cells, others lined by flattened or cuboidal cells. Elastic staining showed a complete absence of elastic tissue.

(4) Sclerotic—These areas were irregular in size and shape and composed of dense fibrous tissue with varying amounts of hyalinisation. Elastic tissue was sparse (fig 3).

Associated features—(1) Haemorrhage: this was irregular in distribution, with extravasation of red blood cells adjacent to the angiomatoid areas, within solid areas and within the lung parenchyma surrounding the tumour. (2) Haemosiderin: in some cases siderotic nodules and haematoxidin pigments were also seen. (3) Foamy histiocite-like cells, mast cells, and other chronic inflammatory cells varied from sparse to conspicuous in different tumours. (4) Fat droplets were evident in cells lining channels and forming solid areas; oil-red-O-positive material was also present in sclerotic areas. (5) Necrosis, cholesterol clefts, and calcification were occasionally present.

Histochemical findings
Enzyme studies performed on fresh tissue from one of the tumours showed slight-to-moderate non-specific esterase activity and slight acid phosphatase activity but no alkaline phosphatase activity in the tumour cells.

Electron-microscope findings
All five tumours showed similar ultrastructural features so they will be described together. The predominant cellular component, regardless of whether it formed sheets, lined papillae or vascular spaces, or was enmeshed in collagen, was considered to be epithelial and to show three forms. There were “dark” cells, “light” cells, and cells transitional between light and dark (fig 8), the variation depending on the quantity of cytoplasmic organelles. Dark cells manifested fairly large amounts of rough endoplasmic reticulum, glycogen, and free ribosomes. These were relatively scanty in light cells and intermediate in amount in transitional cells. Otherwise the three categories of cells showed similar characteristics. They were rectangular, cuboidal, or polygonal with an irregular indented nucleus and peripheral, irregularly condensed chromatin. Most
Benign sclerosing pneumocytoma of lung

Fig 2  Case 1: this shows papillary (left and top), solid (centre right), and angiomatoid (right) areas (H and E × 60).

Fig 3  Case 1: this shows solid area and characteristic papillae formation with sclerotic areas (H and E × 33).

Fig 4  Case 12: angiomatoid areas; irregular spaces, some lined by flattened cells, containing red blood cells (H and E × 110).

Fig 5  Case 7: clear cells with grooved nuclei (H and E × 445).

of these cells contained a single prominent central or eccentric nucleolus but occasionally two nucleoli were observed (fig 9). The Golgi apparatus was usually rudimentary. Moderate numbers of well-formed elongated mitochondria were present. Variable numbers of desmosome-like attachments and tight junctions were observed between the cells. The cells possessed numerous microvillous processes, which in many areas formed complex interdigitating folds (fig 9) but no cilia. Several of the cells contained electron-dense bodies. These varied from one or two per cell within a given section to considerable numbers packing out the cytoplasm. They also varied considerably in size, shape, and complexity. They ranged from small, rounded bodies with a dense central area and light peripheral rim or small rounded bodies with a light centre and dense rim to large complex lamellar structures (fig 10). Multi-vesicular bodies that appeared to be developing within the lamellar structures were also observed. All these were considered to represent stages in development of lamellar bodies as seen in normal type 2 pneumocytes. Many of the cells which appeared foamy on light microscopy, and thus macrophage-like in appearance, were found to be vacuolated epithelial cells. While many of the vacuoles appeared to be empty, some showed residual lamellar structures (fig 11). This appearance was seen in cells lying free within spaces and also within cells lining channels. Basement membrane material was present at the base of cells which lined channels and papillae, between some of the cells forming solid sheets, and within the stroma of the sclerotic areas. Occasional cells lining haemorrhagic spaces or papillae contained numerous mucous vacuoles and were interpreted as mucous cells (fig 10).
Fig 6 (left)  Case 8: abundant glycogen granules in the solid areas (PAS × 470).

Fig 6 (right)  Case 11: hobnail cells covering the papillary projections with cleft-like spaces in between; note a mitotic figure and cells with prominent nucleoli (H and E × 470).

Fig 7  A semi-thin section of a papillary area showing vacuolation of several of the cells lining the papillae; some of the vacuoles are clear areas, others contain a dense homogenous core (toluidine blue × 310).

Fig 8  The edge of a solid area showing light and dark cells with microvillous and interdigitating borders and tight junctions; the dark cells show many complex lamellar structures and smaller simpler dense bodies; a capillary is present at the bottom right (× 4610).
Benign sclerosing pneumocytoma of lung

Fig 9 An "angiomatoid" area showing rectangular and cuboidal cells lining a blood-containing channel. The cells show many microvilli, have irregular indented nuclei, and contain occasional dense bodies. At the bottom right there is a "desquamated" cell that appears similar to the lining cells; and at the top of the picture a cell containing mucous vacuoles can also be seen lining the channel (× 2140).

Fig 10 In this area the lining cells contain numerous complex lamellar bodies; on the left a mucous cell is evident (× 8050).
Within spaces the majority of the cells were red blood cells or epithelial cells (fig 9), many of the latter containing complex lamellar electron-dense bodies. There were occasional macrophages containing similar lamellar structures, presumably phagocytosed, and many fragments of lamellar osmiophilic material (figs 9 and 10). The sclerosed areas contained large amounts of collagen, elastic, basement membrane material, fibroblasts, portions of epithelial cells, and osmiophilic (sometimes lamellated) debris within the stroma.

Occasional mast cells, plasma cells and lymphocytes were seen within the tumour. Occasional capillaries were identified in the tumour but they showed a normal morphology and were not considered to represent a neoplastic component of the tumour. "Weibel-Palade" bodies were not identified in tumour cells despite intensive search.

**Discussion**

All 14 tumours occurred in women, which is in agreement with the striking female preponderance in previously reported series. The age range was wide (16–78 years), which compares with the series reported by Katzenstein et al (15–69 years). No particular predilection for any lobe was noted but most were located near the pleura. All were solitary, but multiple tumours have been reported.

Histologically, the tumour shows a variety of features, which are distinctive but generally not well recognised. Whereas Kennedy emphasised the papillary pattern of the tumour this was not the most prominent feature in most of our cases. If sufficient blocks are examined, a spectrum of histological features will be observed—solid, papillary, angiomatoid, and sclerotic patterns and variable quantities of haemorrhage, haemosiderin, foamy cells, mast cells, lymphoid cells, fat, and glycogen. It is this wide range of patterns that has caused confusion with other tumours. Katzenstein et al emphasised the presence of distinctive round tumour cells and stated that they were a constant feature in the stroma of all microscopic patterns. They were also consistently present in our cases.

Enzyme studies performed on fresh tissue from one of our tumours showed slight-to-moderate non-specific esterase activity, slight acid phosphatase activity, and no alkaline phosphatase activity, which, together with the presence of glycogen, suggest that the tumour cells are epithelial rather than endothelial in type since (1) pneumocytes may contain acid phosphatase and non-specific esterase, whereas endothelium does not; (2) pneumocytes may contain glycogen granules, whereas endothelial cells usually lack it; (3) endothelial cells and pneumocytes may contain alkaline phosphatase. Only one other histochemical study of
sclerosing haemangioma has been reported. Here the tumour cells were negative with ATP-ase and focally positive for alkaline phosphatase in some of the cells lining the tumour spaces.

The results of electron-microscope examination of nine cases of sclerosing haemangioma have been reported; some favoured an epithelial origin, whereas others concluded that it was endothelial. There are three possible reasons for these contradicting conclusions. (1) Not all the tumours studied complied with the original light-microscope description of sclerosing haemangioma. Our perusal of the illustrations shown by each of the above authors makes this explanation very unlikely. (2) Electron-microscope examination of tumours has inherent sampling problems and possibly the areas examined in some cases were not truly representative of the tumour. (3) There may have been interpretative differences. This is a variegated tumour and one element may be interpreted as “entrapped” by one author or “neoplastic” by another.

We are quite certain that the five cases we examined ultrastructurally conform to the original description of Liebow and Hubbell in every way. From our own study we firmly conclude that sclerosing haemangioma is misnamed and has an epithelial derivation.

Occasional capillaries are seen within the tumours but most blood-containing spaces which appear on light microscopy to be vascular channels are lined by cells with epithelial characteristics. The round cells interpreted as neoplastic cells by Katzenstein et al., which are characterised by uniform, bland-appearing, round nuclei with abundant pale cytoplasm and situated within the stroma, also appear to have epithelial characteristics on ultrastructural examination.

Many of the cells, whether lining channels or papillae, forming sheets, or lying within sclerotic areas, had features of type 2 pneumocytes—micronvillous borders, specialised junctions, and complex lamellated bodies. Other cells retained the first two characteristics but had only what we interpreted to be rudimentary “osmiophilic bodies.” We base this interpretation on the fact that similar dense bodies were present in the same cells as more complex lamellated inclusion bodies typical of type 2 pneumocytes and forms intermediate between the two. Moreover, the “rudimentary bodies” show features similar to those described in the developing human fetus. Furthermore, the additional presence of mucous cells supports an epithelial derivation.

Many published reports of sclerosing haemangioma have been of isolated cases. Our series of 14 cases, occurring during 1974–80, seems unusually large. Whether this is fortuitous or due to some environmental factor in Hong Kong remains conjectural. The striking preponderance in women makes it tempting to compare this tumour to the liver cell adenoma occurring in women taking contraceptive pills. Interestingly, many Chinese women ingest Chinese herbs for various conditions—for example, Angelica sinensis (Tan-kuei) after menstruation. Perhaps studies of the effects of these herbs on animals would prove fruitful. That the tumour could represent an unusual pulmonary response to an insult during pregnancy is unlikely since occasional cases have been recorded in men and a very few in children.

The true nature of this tumour is debatable. We do not believe that it represents an inflammatory process, since it is usually a discrete, circumscribed lesion; inflammatory cells are not always evident; and the pneumocytes composing the tumour appear immature ultrastructurally and histochemically. It is unlikely to be hamartomatous since it usually presents, and may be seen to expand, in adult life. We believe it to be a benign neoplasm; and the suggestion made by Hill and Eggleston, that it develops from primitive, incompletely differentiated cells derived from respiratory bronchioles, is the one we find most attractive. This would explain the variation in appearance of the epithelial cells. Furthermore several studies have demonstrated that alveolar epithelium differentiates from respiratory epithelium. Perhaps some environmental factor or sex hormone plays a part in its pathogenesis.

As our observations show that the major cell type within these lesions manifests the characteristics of pneumocytes, we consider the name sclerosing haemangioma inappropriate and likely to perpetuate confusion. The conspicuous light-microscope features of haemorrhage and fibrosis are accompanied by a cellular proliferation of pneumocytes, and we consider that the most appropriate name for this lesion is “benign sclerosing pneumocytoma.” We would emphasise the value of electron-microscope examination, even of formalin-fixed material, wherever there is any difficulty in establishing this diagnosis.

We wish to thank Dr TB Teoh for permission to publish cases 1, 2, 3, and 4; Mr LH Ma for technical assistance; Mrs A Kwan, Mrs V Hamilton, and Miss J Stiffall for secretarial assistance; and Professor ED Williams for guidance in the preparation of this manuscript.
References

Benign sclerosing pneumocytoma of lung (sclerosing haemangioma)
Keeng-Wai Chan, Allen R Gibbs, WS Lo and GR Newman

Thorax 1982 37: 404-412
doi: 10.1136/thx.37.6.404

Updated information and services can be found at:
http://thorax.bmj.com/content/37/6/404

These include:

Email alerting service
Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

Notes

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/