Quantitative study of bronchial mucous gland enlargement

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ABSTRACT Histological sections from 30 lower lobe bronchi, taken from coalminers' lungs collected for the British National Coal Board's Pneumoconiosis Field Research, were selected according to the proportion of mucous gland area occupying the non-cartilaginous part of the bronchial wall. The total gland area expressed as a percentage of the non-cartilaginous wall was called the gland index. Estimations were made of the total number of gland cells and acini on a section and of their numbers per unit area of gland. These estimations were compared with the gland index. The total numbers of gland cells and acini were found to be directly related to the gland index (r=0.84 and 0.86), whereas no relationship was found between the number of gland cells or acini per unit area of gland and the gland index (r=0.08 and 0.02). This indicates that bronchial mucous gland enlargement is primarily a hyperplastic change. A direct relationship between the numbers of gland cells per unit area of gland and acini per unit area gland was found (r=0.69), and this suggests that acinar enlargement is likely to be caused by accumulation of mucus in the acinar lumen.

It has been recognised for many years that the enlargement of bronchial mucous glands is related to excessive production of mucus in chronic bronchitis.1-4 This enlargement has been loosely described as "hypertrophy" on many occasions but recently the term "hyperplasia" has been used.5

It is of interest to know whether the process of bronchial gland enlargement seen in chronic bronchitis is caused primarily by an increase in the size of the gland cells or by an increase in their number. These processes are called hypertrophy and hyperplasia respectively. Reid6 concluded that hypertrophy was the dominant process but later results have not confirmed this. De Haller and Reid,6 for example, found no evidence that the mean acinar diameter increased in chronic bronchitis as might have been expected, and Bedrossian et al7 found little association between acinar density and gland size. This gap in the knowledge of the pathology of chronic bronchitis was commented on by Thurlbeck.8

This paper contains the results of some work undertaken to clarify this aspect of bronchial gland enlargement.

Methods

The bronchi examined formed part of a series obtained from the lungs of coalminers included in the British National Coal Board's Pneumoconiosis Field Research. Three groups of 10 lower lobe bronchi were selected according to the proportion of mucous gland area occupying the non-cartilaginous part of the bronchial wall. The ratio of gland area: non-cartilaginous wall area expressed as a percentage was called the gland index. The three groups of bronchi had gland indices of ≤15, 16—25, and >25.

The sections were cut at 3 μm to minimise nuclear overlap, thereby facilitating the counting of nuclei. The sections were stained with Hae-matoxylin with Van Gieson counterstain. Before examining the sections, their order was randomised to minimise bias.

The following measurements were obtained from each section. (1) The area of the non-cartilaginous wall was measured from a drawing of the projected image of the whole bronchus, and will be called the "soft" wall in the text. (2) The total area of the bronchial glands was measured from drawings made with the aid of a "camera lucida" microscope attachment. The drawings in both these two measurements were made on tracing paper and the relevant areas...
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were measured using a point count grid placed under the paper. (3) The mean number of acini per unit area of gland was determined by the following method. A number of gland fields from each section were photographed and printed to give a magnification of $\times 240$. As many non-overlapping fields as possible were photographed, the number being governed by the area of gland on the section. A sampling square ($50 \times 50$ mm) drawn on a transparent sheet was placed over each photograph at two non-overlapping locations. To minimise selection bias the square was aligned to cover the same positions on each photograph. The number of acini falling within the square was counted and the average reading used to calculate the number of acini per mm$^2$ of gland. Acini crossed by the north and west lines of the square were counted to lie in the square while those crossed by the south and east lines were not. (4) The mean number of mucous and serous gland cells per unit area of gland was determined by a similar method to that used for determining mean number of acini per unit area of gland except that a total photographic magnification of $\times 650$ was used. The same sampling grid ($50 \times 50$ mm) and sampling technique was used. Two bronchial sections were later omitted as the nuclei were indistinct because of autolysis.

Measurements (1) and (2) were combined as follows to give the gland index or proportion of wall occupied by glands:

$$\text{Gland index} = \frac{\text{gland area} \times 100}{\text{non-cartilaginous wall area}}$$

An estimation of the number of cells in the complete section was calculated as follows: cell density (cells mm$^{-2}$ gland) $\times$ total gland area $=$ total gland cells in bronchial section.

To allow for variations in bronchial dimensions between individuals the gland areas were adjusted to a “soft” wall area of 30 mm.$^2$. This figure was chosen as it approached the average value (33.1 mm$^2$) for the cases in the study.

**Results**

A summary of the results is given in Table 1. As the gland index increased, the “total cell number” increased but the acinar densities remained the same.

Figure 1 shows the linear relationship between the total gland cell number on the bronchial section and the gland index ($r = 0.84$). A similar relationship was found between “total acini” and gland index ($r = 0.86$).

**Table 1 Summary of results**

<table>
<thead>
<tr>
<th>Gland index</th>
<th>Area of gland on bronchial section (a)*</th>
<th>Acinar density</th>
<th>Number of gland cells in (a)</th>
<th>Number of cases</th>
</tr>
</thead>
<tbody>
<tr>
<td>$\leq 15$</td>
<td>3-18</td>
<td>201</td>
<td>4204</td>
<td>10</td>
</tr>
<tr>
<td>$15 \leq 25$</td>
<td>5-34</td>
<td>211</td>
<td>8690</td>
<td>8</td>
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<tr>
<td>$&gt; 25$</td>
<td>9-02</td>
<td>212</td>
<td>14084</td>
<td>10</td>
</tr>
<tr>
<td></td>
<td>SD</td>
<td>SD</td>
<td>SD</td>
<td>SD</td>
</tr>
<tr>
<td></td>
<td>0-94</td>
<td>51</td>
<td>1349</td>
<td>5</td>
</tr>
<tr>
<td></td>
<td>0-40</td>
<td>34</td>
<td>1836</td>
<td>3</td>
</tr>
<tr>
<td></td>
<td>1-15</td>
<td>37</td>
<td>3146</td>
<td>6</td>
</tr>
</tbody>
</table>

*Corrected to soft wall area of 30 mm.$^2$.

Differences in the number of gland cells between gland index groups are statistically significant ($p < 0.001$).

**Fig 1 Total number of gland cells on a section plotted against the gland index ($r=0.84$, $p<0.001$).**

**Fig 2 Mean number of acini per mm$^2$ gland plotted against the gland index ($r=0.02$, NS).**
Fig 3  Relationship between the mean number of gland cells per mm$^2$ gland and the mean number of acini per mm$^2$ gland ($r=0.69$, $p<0.001$).

Figure 3 shows that a decrease in acinar density was associated with a decrease in cell density ($r=0.69$), suggesting hypertrophic changes. Hypertrophy, where present, however, was not associated with an increase in gland index (fig 2). Correlation coefficients were calculated for some of the pairs of variables and these are shown in table 2.

Discussion

The results show that the dominant process in bronchial mucous gland enlargement is hyperplasia. The proportion of total gland area in the bronchial wall has been used as an index of gland enlargement previously, but in each case total bronchial wall was used. Restrepo and Heard found that the proportion of cartilage in the bronchial wall of large airways was unaffected in bronchitics, and so the use of the total wall area minus the area of cartilage has been used here in the calculation of the gland index to make it more sensitive to changes in the glands.

<table>
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<tr>
<th></th>
<th>Gland index</th>
<th>p</th>
<th>Cell number</th>
<th>p</th>
<th>Acinar number</th>
<th>p</th>
<th>Cell density</th>
<th>p</th>
<th>Acinar density</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cell number</td>
<td>0.84</td>
<td>&lt; 0.001</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Acinar number</td>
<td>0.86</td>
<td>&lt; 0.001</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cell density</td>
<td>0.08</td>
<td>NS</td>
<td></td>
<td></td>
<td></td>
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<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Acinar density</td>
<td>0.02</td>
<td>NS</td>
<td>0.69</td>
<td>&lt; 0.001</td>
<td></td>
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</table>

It is unfortunate that the clinical histories of the cases examined are not known in sufficient detail to classify them according to their bronchitic symptoms. It has previously been shown, however, that the proportion of gland in the bronchial wall increases in bronchitics, and it seems likely from the evidence presented here that this increase is caused by a hyperplastic change of the glands. It is also possible that this type of change might be partly responsible for the maintenance of bronchitic symptoms since a hyperplastic change is less likely to revert to the original state than a hypertrophic change.

The results show that hypertrophic changes do occur but are not obviously related to an increase in gland index. Although the number of cases examined was small, it appears that the relationship between cell density and acinar density is linear, the slope of the line representing the mean number of cells associated with an acinus on the section. This suggests that acinar enlargement may not be caused by an increase in gland cell size. If that was the case then it might be expected that the number of gland nuclei associated with an acinus would decrease as the acinar density decreased. This is because compared to the thickness of the section, an acinus is a large structure, and acinar density will be unaffected by section thickness. The density of nuclei, however,
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may be affected as the diameter of a nucleus is similar to the thickness of a section. If a hypertrophic change is present then as the volume of cell cytoplasm increases the probability of nuclei appearing in the section, and therefore the number of nuclei associated with any acinus, should decrease (fig 4). The data presented suggest that such reduction does not occur and thus that cellular hypertrophy is not the primary mechanism of acinar enlargement.

It remains a possibility that acinar enlargement may be caused by swelling of the acinar lumen as a result of mucus accumulation (fig 4). Where the glands are greatly enlarged the amount of mucus to be expelled through the gland duct is likely to be considerably increased and this may result in a back pressure which could swell the acinus. Alternatively a back pressure might result from alterations in the rheological properties of the mucus. Such changes in the properties of mucus have been reported but variation in the properties of mucus among bronchitics is considerable. Such changes in the properties of mucus have not been reported as being associated with gland size and could account for the variations in cell and acinar densities with gland index.

These results show that a major factor contributing to bronchial gland enlargement is an increase in cell number, and where acinar enlargement is seen this is likely to be caused by engorgement with mucus.

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References

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