Cardiovascular depressant effect of protamine sulphate: experimental study and clinical implications

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Fadali, M. A., Papacostas, C. A., Duke, J. J., Ledbetter, M., and Osbakken, Mary (1976). Thorax, 31, 320–323. Cardiovascular depressant effect of protamine sulphate: experimental study and clinical implications. The mechanisms underlying protamine-induced hypotension and bradycardia were the subject of this investigation. Six groups of dogs with intact circulation were tested in controlled circumstances with various drugs. The following parameters were observed: femoral arterial pressure, central venous pressure, left ventricular pressure and its rate of rise, left ventricular contractile element velocity of shortening, maximal Vce, and cardiac output. The six groups were studied under these pharmacological conditions: ganglionic and adrenal medullary block with hexamethonium chloride, postganglionic parasympathetic blockade by atropine sulphate, alpha and beta adrenergic receptor block by phenoxybenzamine and propranolol respectively, and depletion of endogenous histamine by compound 48/80 (a condensation product of p-methoxyphenethyl methylamine with formaldehyde). The last group was put on extracorporeal circulation to isolate the vascular tree from the heart. The effect of the drug on this isolated vasculature was observed by recording the femoral arterial pressure. Our findings show that the hypotension and bradycardia are produced by a direct effect of protamine on the myocardium and peripheral vascular system.

Heparin is necessary for the performances of extracorporeal circulation. Protamine sulphate is used to neutralize heparin at the completion of cardiopulmonary bypass (Hurt et al., 1956). Hypotension and bradycardia, the common side effects of protamine, can be detrimental to postcardiotomy patients (Jaques, 1949; Egerton and Robinson, 1961). The mechanism of action of protamine on the intact cardiovascular system and isolated peripheral vasculature has been investigated and is the subject of this paper.

METHODS
A total of 40 adult mongrel dogs weighing 14 to 23 kg were divided into seven groups. Six groups (A–F) of six dogs each had intact circulations, and the seventh group (G) of four dogs was put on total cardiopulmonary bypass. All dogs were anaesthetized with thiopental sodium, intubated and ventilated by a Harvard mechanical respirator.

DOGS WITH INTACT CIRCULATIONS In groups A–F a transsternal incision through the left fifth and right fourth interspaces was used to expose the heart. An electromagnetic flow probe (Statham) was placed around the ascending aorta to measure cardiac output (CO). Polyethylene catheters were introduced into the left ventricular cavity, left femoral artery, and left femoral vein to measure left ventricular pressure (LVP), its rate of rise (dp/dt), the femoral arterial pressure (FAP), and central venous pressure (CVP) respectively. The left ventricular contractile element velocity (Vce) was determined from isovolumic pressure (IP) and
dp/dt; maximal Vce (V max) was obtained by construction of an IP to Vce curve and extrapolation to zero load (Mason, Spann, and Zelis, 1970). Electrocardiograms were always recorded.

Each animal in groups A–F received intravenous heparin, 5 mg/kg, followed by protamine sulphate, 10 mg/kg. Group A animals had both carotid arteries exposed and clamped proximal to the sinus region to test the carotid sinus reflex after heparin-protamine administration. Groups B–F were pretreated intravenously with these drugs: group B was given hexamethonium chloride, 10 mg/kg, to block the autonomic ganglia and epinephrine release by the adrenal medulla. Group C received atropine sulphate, 0.5 mg/kg, to block postganglionic parasympathetic receptors (vagal block). Norepinephrine, 2 μg/kg, was given after 10 minutes to test the vagal block. Ordinarily, an increase in blood pressure would be followed by reflex bradycardia; however, an atropinized dog would have no reflex bradycardia following an increased blood pressure. Group D dogs were given phenoxybenzamine (Dibenzyline) 5 mg/kg and a 30-minute waiting period to achieve complete alpha blockade. In group E, propranolol (Inderal), 1 mg/kg, was given with a 10-minute waiting period for complete sympathetic beta blockade. Group F dogs were given compound 48/80 (a condensation product of p-methoxyphenethyl methyleamine with formaldehyde), 0.1 mg/kg, slowly over a period of 20 minutes to deplete endogenous histamine (Paton, 1951; Papacostas, Loew, and West, 1959). Before and after heparin-protamine administration the blood pH was tested in one animal of each group.

The four dogs in group G were cannulated and put on extracorporeal circulation. The heart was isolated by clamping the ascending aorta and pulmonary artery. The effect of protamine on the vascular tree was measured by observing the FAP before and after injection of 10 mg/kg of protamine into the arterial inflow of the extracorporeal pump.

**TABLE**

**MEAN PERCENTAGE CHANGE IN DIFFERENT PARAMETERS FOLLOWING HEPARIN-PROTAMINE ADMINISTRATION IN SEVEN ANIMAL GROUPS**

<table>
<thead>
<tr>
<th>Premedication</th>
<th>Group A</th>
<th>Group B</th>
<th>Group C</th>
<th>Group D</th>
<th>Group E</th>
<th>Group F</th>
<th>Group G</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean FAP (mmHg)</td>
<td>None</td>
<td>Hexamethonium</td>
<td>Atropine</td>
<td>Phenoxybenzamine</td>
<td>Propranolol</td>
<td>Compound 48/80</td>
<td>Extracorporeal circulation</td>
</tr>
<tr>
<td>39</td>
<td>24</td>
<td>29</td>
<td>29</td>
<td>31</td>
<td>25</td>
<td>24</td>
<td></td>
</tr>
<tr>
<td>IP (mmHg)</td>
<td>44</td>
<td>33</td>
<td>40</td>
<td>38</td>
<td>41</td>
<td>41</td>
<td></td>
</tr>
<tr>
<td>dp/dt</td>
<td>38</td>
<td>32</td>
<td>40</td>
<td>36</td>
<td>41</td>
<td>37</td>
<td></td>
</tr>
<tr>
<td>V max (ml/sec)</td>
<td>31</td>
<td>27</td>
<td>33</td>
<td>27</td>
<td>30</td>
<td>27</td>
<td></td>
</tr>
<tr>
<td>CO (ml/min)</td>
<td>33</td>
<td>26</td>
<td>29</td>
<td>28</td>
<td>32</td>
<td>32</td>
<td></td>
</tr>
</tbody>
</table>
tamine. Group A animals tested the concept of neurogenic reflex by some authors (Egerton and Robinson, 1961; Minker and Koltai, 1964). Proximal carotid artery clamping reversed the hypotension and bradycardia produced by protamine which attests to the integrity of the carotid sinus reflex mediated by autonomic pathways. To test further the neurogenic reflex concept different steps of the autonomic pathways were blocked through premedication. In group B, the autonomic ganglia and the adrenal medulla were blocked by hexamethonium chloride. The femoral arterial pressure, left ventricular pressure, its rate of rise, isovolumic pressure, maximal left ventricular contractile element velocity, and cardiac output were substantially reduced following protamine administration, a response similar to that in dogs with intact ganglia (group A). Thus protamine does not produce its cardiovascular effects through blocking the autonomic ganglia. Atropine sulphate was used to block postganglionic parasympathetic receptors in group C dogs to eliminate possible reflex cholinergic effect of protamine. Again, as in group A, the animals showed hypotension and depression of myocardial contractility when protamine was given. Alpha blockade by phenoxybenzamine in group D and beta blockade by propranolol in group E both failed to avert the hypotension and bradycardia after protamine injection. The findings in groups A–E do not support the involvement of neurogenic reflexes or adrenergic receptors in protamine-induced hypotension and bradycardia.

Sensitivity and histamine release due to protamine was proposed by Shelley, Hodgkins, and Visscher (1942). Kern and Langner (1939) could not produce protamine sensitivity in guinea-pigs despite several intraperitoneal injections. Jaques (1949) found that histamine-free protamine preparations could still elicit hypotension. Presently, depletion of endogenous histamine was produced in group F dogs by premedication with compound 48/80. The hypotension and bradycardia was again elicited following protamine injection. This reinforces the findings of Jaques (1949) and Kern and Langner (1939) that histamine has no significant etiological role in these effects of protamine.

Myocardial contractility depression following protamine administration was demonstrated by Goldman, Joison, and Austen (1969) but disputed by Gourin et al. (1971a), who even advocated an inotropic effect of the drug on dog's myocardium. By estimating the maximal velocity of left ventricular contractile element shortening, the contractile state of an intact heart can be quantified. The techniques of Mason et al. (1970) were used in the present study, and the maximal velocity was computed from the instantaneous correlation of the rate of pressure developed in the left ventricular cavity during the isovolumic phase of contraction. All animals in groups A–F showed a marked decrease in myocardial contractility after protamine injection.

The findings in group G provided further evidence for the direct effect of protamine on the vascular tree. Protamine administration into the arterial inflow of the extracorporeal circulation was followed by a significant drop of femoral arterial pressure.

Acid-base changes are not involved because no significant pH changes were recorded after heparin-protamine injection.

From the results we conclude that the cardiovascular responses to protamine sulphate administration are due to a direct effect of the drug on the myocardium and vasculature.

This experimental conclusion was clinically applied to our patients receiving protamine infusion at the termination of cardiopulmonary bypass. To prevent hypotension, the following measures were successful in the majority of cases: (1) Administration of inotropic agents, in particular calcium chloride or calcium gluconate; isoproterenol or epinephrine to patients without serious tachycardia or significant arrhythmias. Norepinephrine might be used since it reverses the peripheral vasodilating effects of protamine without producing tachycardia. (2) If central venous and left atrial pressures permit, a fluid load is administered to compensate for the expanded capacity of the vascular tree.

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