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Doxycline in serum and bronchial secretions

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Hartnett, B. J. S. and Marlin, G. E. (1976). Thorax, 31, 144–148. Doxycline in serum and bronchial secretions. The concentration of doxycline hydrochloride was measured in serum and bronchial secretions in five patients with chronic bronchitis receiving doxycline orally in normal therapeutic dosage for seven days (200 mg day 1, 100 mg days 2 to 7). After the loading dose of 200 mg, serum concentrations ranged between 5-40 and 3-45 µg/ml (mean 4-33 µg/ml) at 3 hours, declining to between 2-28 and 1-21 µg/ml (mean 1-71 µg/ml) at 23 hours. The mean serum levels for days 2 to 7 were 2-15, 1-79, and 1-38 at 3, 8, and 23 hours respectively. There was considerable individual variability and a wide range of concentrations of doxycline in the sputum (0-07 to 2-10 µg/ml, mean 0-34 µg/ml). During the course of treatment there was a progressive increase in sputum levels and sputum/serum concentration ratios. There was no correlation between sputum concentration and degree of purulence. The clinical efficacy of doxycline does not appear to be related to sputum concentration, although the progressive increase in sputum doxycline levels may be relevant in preventing recurrence of acute infection when the drug is administered as long-term prophylactic therapy.

Tetracyclines are commonly used as bacteriostatic chemotherapy for the treatment of acute exacerbations of chronic bronchitis. The most common infecting organisms are Haemophilus influenzae and pneumococcus, which are both sensitive in vitro to tetracyclines (May, 1953 and 1964; May and May, 1963). The site of infection is the bronchial mucosa where an effective concentration of antibiotic must be reached to eliminate the organism (Hers and Mulder, 1953). A gradient of concentration of antibiotic presumably exists from blood through bronchial mucosa to bronchial secretions. The efficacy of treatment may also depend on the concentration of antibiotic in bronchial secretions. This concentration should be sufficient to suppress bacterial growth, otherwise there will remain a constant reservoir of organisms, and reinfection is likely, a major problem in the management of chronic bronchitis (May and Delves, 1965). An assessment of the response to chemotherapy may be provided if the concentration of antibiotic obtained in bronchial secretions is known.

Doxycycline (alpha-6-desoxytetracycline) hydrochloride is a tetracycline derivative which retains the antibiotic properties of this group but has other beneficial characteristics. The serum half-life in adult man is between 15 and 22 hours, so that a single daily dose is sufficient for therapy (Fabre et al., 1966; Rosenblatt et al., 1966; Jacobs and Robinson, 1969). There is only minimal depression of absorption when doxycline is administered with food (Rosenblatt et al., 1966), and the drug may be used safely in patients with renal insufficiency (Little and Bailey, 1970).

The purpose of this work was (a) to measure the concentration of doxycline in bronchial secretions in patients receiving doxycline orally in normal therapeutic dosage over a seven-day period, and (b) to determine the relationship between serum and sputum concentrations of doxycline.

METHODS

PATIENTS Five hospital patients with chronic bronchitis defined according to the terminology of the Ciba Guest Symposium Report (1959) were selected for this study. All patients were able to produce ample quantities of sputum throughout the day and none had received treatment with antibiotics or mucolytic agents during the previous three days. Four patients had acute exacerbations of chronic bronchitis and were producing mucus purulent sputum at the time of entry to the study.
Doxycycline in serum and bronchial secretions

There was no evidence of pneumonic consolidation on the chest radiograph of any patient. The informed consent of each patient was obtained after the procedure of the trial had been fully explained.

**DESIGN** Each patient received a seven-day course of doxycycline, starting with a loading dose of 200 mg at 0900 hours on the first day and subsequently 100 mg at the same time on the following six days. During the trial, supportive treatment with an aerosol bronchodilator (orciprenaline) and physiotherapy was continued at regular intervals. The patients received a light breakfast two hours before drug ingestion. Samples of venous blood and sputum were collected at intervals of 3, 8, and 23 hours after each dose of doxycycline. Sputum was accumulated in plastic containers during the 30-minute period before appointed collection times and was classified as mucoid, mucopurulent or purulent according to the quantity of pus present on naked-eye examination.

**ASSAY OF DOXYCYCLINE** The doxycycline concentration in serum and sputum samples was measured biologically by a large plate agar diffusion method, using a punch hole technique with *Bacillus cereus* (var. *mycoides* 11778) as test organism.

**Serum Assay** Standard solutions of doxycycline (0-1, 0-2, 0-4, 0-6, 1-0, and 2-0 μg/ml) were prepared using 1:2 Bovine Albumin 3.5% solution and phosphate buffer pH 4-5 as diluent.

**Sputum Assay** Sputum from six other bronchitic patients not receiving treatment with antibiotics was homogenized by centrifugation at 10 000 rev/min for 30 minutes for the preparation of 0-1, 0-2, 0-4, 0-8, 2-0, and 3-0 μg/ml doxycycline standards. In preliminary studies there was no significant difference in results obtained by preparing the sputum by conventional centrifugation, ultracentrifugation or proteolytic homogenization.

**Preparation of Assay Plates** A fresh overnight culture of *B. cereus* was prepared and washed from an agar slope with sterile distilled water to give a suspension having a 50% transmission at 650 μm on a visible spectrophotometer. The suspension was shaken for one hour to produce homogeneity and used as a 1% inoculum. Assay plates were prepared with a layer of 250 ml seed agar BBL 10937, with pH 7.0 containing 2-5 ml of the prepared suspension of *B. cereus*. The assay method then followed the procedure described by Campbell (1970). The lower limit of detectability of doxycycline by this method in both serum and sputum was 0.01 μg/ml, and the variability of estimations was less than 5%.

**RESULTS**

The mean serum and sputum doxycycline concentrations for the five patients at 3, 8, and 23 hours after each dose of the drug are shown in Tables I and II respectively. Statistical analysis of these results was performed using the paired Student's *t* test.

Peak serum doxycycline concentrations were recorded 3 hours after dosage on each day of the trial, and although there was a subsequent decline, appreciable concentrations were detected 23 hours after ingestion. After the loading dose of 200 mg,

### Table I

**Mean ± Standard Error of the Mean Serum Doxycycline Concentrations for the Five Patients at 3, 8, and 23 Hours After 200 mg on Day 1 and 100 mg on Days 2 to 7 of Treatment**

<table>
<thead>
<tr>
<th>Hours</th>
<th>Serum Concentrations (Mean ± SEM) (μg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Day 1</td>
</tr>
<tr>
<td>3</td>
<td>9.33 ± 0.35</td>
</tr>
<tr>
<td>8</td>
<td>2.92 ± 0.35</td>
</tr>
<tr>
<td>23</td>
<td>1.71 ± 0.19</td>
</tr>
</tbody>
</table>

### Table II

**Mean ± Standard Error of the Mean Sputum Doxycycline Concentrations for the Five Patients at 3, 8, and 23 Hours After 200 mg on Day 1 and 100 mg on Days 2 to 7 of Treatment**

<table>
<thead>
<tr>
<th>Hours</th>
<th>Sputum Concentrations (Mean ± SEM) (μg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Day 1</td>
</tr>
<tr>
<td>3</td>
<td>0.23 ± 0.12</td>
</tr>
<tr>
<td>8</td>
<td>0.19 ± 0.02</td>
</tr>
<tr>
<td>23</td>
<td>0.13 ± 0.02</td>
</tr>
</tbody>
</table>
serum concentrations ranged between 5.40 and 3.45 µg/ml (mean 4.33 µg/ml) at 3 hours, declining to between 2.28 and 1.21 µg/ml (mean 1.71 µg/ml) at 23 hours. The mean serum levels for days 2 to 7 were 2.15, 1.79, and 1.38 µg/ml at 3, 8, and 23 hours respectively. The peak serum levels on days 1 and 2 were significantly higher than on day 7 (p<0.01), confirming a loading dose effect and suggesting that plateau levels were achieved after two days of treatment.

Doxycycline was detectable in the sputum throughout the study, including the first specimens 3 hours after starting treatment. Sputum concentrations were lower than those achieved at corresponding times in the serum, ranging from 2.3% to 77.5% (mean 19.9%) of the serum value. Peak levels were observed usually 3 hours after dosage, and in most instances declined at 23 hours. There appeared to be a gradual accumulation of doxycycline in the sputum as the treatment progressed (see Table II and Figure). The sputum concentrations on days 1, 2, and 3 were significantly less than on days 3 to 7, day 7, and days 4 and 7 respectively (p<0.05). The sputum/serum concentration ratio also increased progressively throughout the trial (see Table III and Figure).

**FIGURE** Mean sputum concentrations and sputum/serum concentration ratios of doxycycline for the five patients for the one-day period after each dosage (200 mg on day 1, 100 mg on days 2 to 7) plotted against day of treatment.

<table>
<thead>
<tr>
<th>Hours</th>
<th>Sputum/ Serum Concentration Ratio (Mean ± SEM)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Day 1</td>
</tr>
<tr>
<td>3</td>
<td>0.053 ± 0.003</td>
</tr>
<tr>
<td>8</td>
<td>0.066 ± 0.010</td>
</tr>
<tr>
<td>23</td>
<td>0.075 ± 0.003</td>
</tr>
</tbody>
</table>

These ratios on days 1, 2, and 3 were significantly less than on days 2 to 7 (p<0.01), days 4 to 7 (p<0.05), and days 4 and 7 (p<0.05) respectively. Pus was eliminated from the bronchial secretions within 48 hours in all four patients who had mucopurulent sputum at the start of the trial.

**DISCUSSION**

There was considerable individual variability and a wide range of concentrations of doxycycline in bronchial secretions, a finding observed with other antibiotics in chronic bronchitis (Hafez, Stewart, and Burnet, 1965; May and Delves, 1964 and 1965; Campbell, 1970; Stewart et al., 1970; May and Ingold, 1972; MacCulloch, Richardson, and Allwood, 1974; Stewart et al., 1974). Careful precautions were taken to avoid dilution of sputum by salivary contamination, although this is unlikely to alter sputum levels appreciably (May and Delves, 1965). The range of sputum concentrations of doxycycline was generally low (0.07 to 2.10 µg/ml, mean 0.34 µg/ml) and similar to that observed by MacCulloch et al. (1974) after a comparable dosage regimen (0.05 to 3.40 µg/ml, mean 0.86 µg/ml). This poor excretion into the sputum is particularly noticeable during the first 48 hours despite the administration of a loading dose. Campbell (1970) also observed that sputum tetracycline levels were lower during the first 24 hours of therapy and often only peaked after 48 hours. Administration of a higher daily dose of doxycycline might result in higher sputum concentrations.

The progressive increase in sputum doxycycline concentration during the course of treatment occurred despite lack of accumulation in the serum. This behaviour has not previously been observed with other antibiotics and might be related to the drug’s long half-life. However, penetration of doxycycline into bronchial secretions does not appear to be a simple diffusion process, because of lack of correlation between serum and sputum levels. An active transport mechanism for excretion into sputum would favour accumulation of the drug in this reservoir.
This would also be evident if the drug was bound to constituents of the sputum, eg, protein. It is known that doxycycline is highly bound to serum proteins (Rosenblatt et al., 1966). Doxycycline has high lipid solubility necessary for penetrating cell membranes readily and may also bind strongly to bronchial epithelial cell constituents. If the drug is released only slowly from these cells, its initial availability for penetrating into the sputum will be reduced.

There was no correlation between sputum concentrations and degree of purulence, in contrast to the behaviour of ampicillin whose transport is facilitated by the presence of pus (May and Delves, 1965; Stewart et al., 1970). Nevertheless, tissue levels of doxycycline during the first 48 hours were apparently adequate as pus was eliminated from the sputum in the four patients presenting with mucopurulent secretions. Liss and Norman (1975) found doxycycline levels of between 2-87 and 4-95 μg/g tissue in the lungs of patients undergoing cardiopulmonary surgery after 200 mg 24 hours, and after 100 mg 5–7 hours, before surgery. They observed by fluorescent microscopy the highest concentration of doxycycline in the bronchiolar epithelium. Gartmann (1975) determined doxycycline concentrations in human surgical lung specimens after 200 mg, 27 hours and 100 mg approximately 3 hours before surgery and found levels of between 1-0 and 8-5 μg/g in lung tissue, between 1-89 and 3-49 μg/g in bronchial wall, and usually below 1-0 μg/ml in bronchial secretions. The mean inhibitory concentration (MIC) of doxycycline against Haemophilus influenzae is 1-0 μg/ml and against the pneumococcus 0-19 μg/ml (Williamson, 1968), such levels being reached in lung and bronchial wall but not consistently in bronchial secretions for H. influenzae. In this present study, only in two patients during the latter half of treatment were sputum levels greater than the MIC for H. influenzae, but they reached that for the pneumococcus in all patients at least on one collection time each day.

Doxycycline has proved to be an effective antibiotic in the treatment of acute exacerbations of chronic bronchitis with efficacy similar to ampicillin (Aitchison, Grant, and Gould, 1968; Bennion-Pedley, 1969) and oxytetracycline (British Thoracic and Tuberculosis Association, 1973). The results of this study indicate that the clinical efficacy of doxycycline does not appear to be directly related to sputum concentration but more likely to levels achieved in lung and bronchial wall. The progressive increase in sputum concentration observed during treatment may be of relevance to the suppression of bacterial growth and the recurrence of acute infection when doxycycline is administered as long-term prophylactic therapy.

We wish to thank Dr. B. W. Gunner, Medical Director, Pfizer Pty Ltd, for providing helpful advice and supplies of doxycycline. We are also indebted to Mr. R. Keating, B. Pharm., Pfizer Pty Ltd, who performed the doxycycline assays, and to Dr. G. Graham for statistical advice.

REFERENCES


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