A new method for tracheal reconstruction

H. ALETRAS, V. O. BJÖRK, G. HULTQUIST, AND L. RODRIGUEZ

From the Departments of Thoracic Surgery and Pathology, University Hospital, Uppsala, Sweden

Various methods and materials have been tried in dealing with tubular or 'sleeve' resection of the trachea. In relatively short tracheal defects, 2–4 cm. in clinical length, an end-to-end anastomosis gives the most satisfactory result (Kay, 1952; Crafoord, Björk, and Hilty, 1954; Björk, 1955; Paulson and Shaw, 1955; Mathey, Denis, and Binet, 1960). In some cases a 'montage' operation without prosthetic material can be used (Barclay, McSwan, and Welsh, 1957) even in longer resections of up to 5–6 cm.

To treat more extensive tubular defects the use of a prosthesis is necessary. Tracheal homografts, 'fresh' or 'pre-treated,' and aortic homografts (Jackson, O'Brien, Tuttle, and Meyer, 1950; Davies, Edmiston, and McCorkle, 1952; Rob and Bromley, 1953; Björk and Rodriguez, 1958) have been proved experimentally to be unsuccessful, as the graft is rejected by the tissues of the host.

Two forms of prosthesis of alloplastic material have been used both experimentally and clinically, alone or in combination with free or pedicled autogenous grafts. These are meshes (Rob and Bromley, 1953; Ferguson, Wild and Wangensteen, 1950; Swift, Grindlay and Clagett, 1952; Morfit, Neerken, Prevedel, Liddle, and Kircher, 1954; Keshishian, Blades, and Beattie, 1956; Dor and Carcassonne, 1958; Greenberg, 1960; Harrington, Beall, Morris, and Usher, 1962; Beall, Harrington, Greenberg, Morris, and Usher, 1962; Greenberg and Willms, 1962; Rob and Bateman, 1949; Ravitch, 1951; Brewer and Bai, 1955; Cummins and Waterman, 1957) and non-porous metallic or plastic tubes (Björk and Rodriguez, 1958; Keshishian et al., 1956; Dor and Carcassonne, 1958; Daniel, 1948; Clagett, Grindlay, and Moersch, 1948; Craig, Holmes, and Shabart, 1953; Rush and Clifton, 1956; Jesseph, Stevenson, Harkins and Merendino, 1956; Pressman and Simon, 1958; Moncrief and Salvatore, 1958; Taber and Tomatis, 1958; Bell, 1960; Michelson, Solomon, Maun, and Ramirez, 1961; Aronstam, Nims, and Winn, 1961; Kramish, Rutherford, Morfit and Lunt, 1961; Longmire, 1948; Cotton and Penido, 1952; Clagett, 1956; Burnett, Stanley, and Stringer, 1962).

The usual causes of the almost uniformly unsuccessful results of tracheal reconstruction using a prosthesis have been (1) stricture from granulation through the mesh or at the site of anastomosis; (2) pressure necrosis from the prosthesis; and (3) stricture of the trachea after dislodgment or removal of the prosthesis.

A promising method was reported by Ekeström (1956; 1958); a teflon frame is embedded for five to 12 weeks subcutaneously to form an autogenous tissue tube which is used for the prosthesis.

In order to avoid the two months' delay before a tracheal reconstruction can take place and to prevent stricture from a suture granuloma at the site of the anastomosis a new method has been developed. The aim of this paper is to present this method and the experimental results obtained.

**METHOD**

**THE INNER TUBE** To prevent stricture from granulations at the site of the anastomosis to the prosthesis it is necessary to extend the prosthesis 5 to 10 mm. on the inside of the trachea at each anastomosis. The suture to the divided tracheal end was therefore secured to a special ridge on the outside of the inner tube. Silastic reinforced by nylon and covered with dacron has been used in order to diminish the incidence of pressure necrosis (Fig. 1 A). Inner tubes of teflon, known to be well tolerated, have also been used for anastomosis to the trachea (Fig. 1 B to D).

**THE OUTER TUBE** An outer tube was created around the rigid inner tube by pedicled pericardium wrapped around the tube and sutured to itself and to the outside of the trachea at each end. Rings were then applied over the pericardium. The aim was that the rings should become embedded in the medistinal tissue and act as cartilage, preventing stricture of the newly formed trachea in case the inner tube should become dislodged. The rings were of two kinds, teflon rings, and rings of stainless steel covered with ivalon sponge. The teflon rings were kept in place by sutures through drill holes.
A new method for tracheal reconstruction

The operation Fifty-seven dogs were used. Anaesthesia was induced with intravenous thiopentone and a short-acting muscle relaxant (sucinylcholine chloride). Nitrous oxide and oxygen were then administered through an intratracheal tube with the aid of a volume-regulated Engström respirator. Thoracotomy was performed under the fourth rib on the right.

FIG. 1. Diagram showing the different prostheses (2) sutured in place covered by pedicled pericardium (4) and rings (3). A: Inner tube of silastic (2) covered with dacron (5) which is sewn on to the trachea (1). The pedicled pericardium (4) is surrounded by rings (3). B: Inner tube of teflon (2) sutured in place. C: Inner tube of teflon (2) with a straight sewing ridge for fixation to the trachea (1). D: Inner tube of teflon (2) with a rounded sewing ridge for enclosure of the tracheal end (1).

FIG. 2. (Above) The silk sutures are passed through the tube of dacron to the trachea so that the inner tube of silastic will come on the inside of the trachea, preventing stricture at the anastomosis. (Below) The inner tube is covered by a flap of pedicled pericardium and then by teflon rings. The rings are fixed by sutures through drill holes.
side. Theazygous vein was divided, the trachea liberated, and six to nine rings resected. The trachea was first divided at the cranial end and the peripheral portion was used for ventilation through a rubber tube introduced from the open thorax. The prosthesis was then sutured in position into the cranial portion of the trachea with isolated silk sutures to an airtight connexion (Fig. 2, above).

As a second step, the trachea was divided above the bifurcation, and four sutures were placed at equal distances in the trachea and secured to the prosthesis, after which ventilation could be continued at an increased volume through the tube introduced through the larynx. Air leaked around the central anastomosis until a sufficient number of sutures had been inserted.

As a third step, a pedicled flap of pericardium, with the pedicle situated cranially, thus preserving the blood supply, was created. This pericardial flap was brought behind the superior vena cava and around the prosthesis and sutured to itself and to both ends of the trachea. Then three to four rings of teflon or ivalon reinforced by stainless steel wire were placed outside the pericardium and fixed 0·5 cm. apart by sutures (Fig. 2, below). The chest was closed with one tube for drainage, and tracheostomy was performed in all cases.

RESULTS

CONTROL GROUP A group of six dogs were used as controls. Only the inner tube was introduced and left to see how long it would stay in place. In five cases a tube of silastic covered with a nylon sewing ring was used. In four dogs the prosthesis was displaced, with stricture formation from the surrounding granulation tissue after 36, 55, 97, and 183 days (Fig. 3). In one dog the prosthesis was removed by bronchoscopy after 50 days, which caused collapse of the trachea and death.

In the remaining dog (11) a teflon tube (Fig. 1D) was used and functioned well for four years after which the dog died during transport (Fig. 4).

Conclusion A silastic tube has a tendency to become dislodged, thus causing collapse of the trachea. A teflon tube was used successfully for tracheal replacement for four years in one dog.

GROUP A The prosthesis used was an inner tube of silastic and rings of ivalon reinforced by stainless steel. Twenty-four dogs were operated upon; there were seven long-term survivors and 17 dogs died within seven days. The immediate mortality was ascribed to suture insufficiency (3), pneumonia (3), error in anaesthesia (2), acidosis (6), haemorrhage from tracheostomy (1), embolism with hemiplegia (1), and haemothorax (1).

FIG. 3. (Above) Collapse of the unsupported new trachea after the inner tube of silastic had become dislodged 183 days after operation. (Below) Cartilage proliferation on the outer side of the tracheal ring close to the resectional margin. Weigert's haematoxylin and van Giesen, × 35.
A new method for tracheal reconstruction

FIG. 4. Gross appearance of the trachea (dog 11) four years after resection of nine rings. The teflon prosthesis was found to be in place. The sewing ridge helped fixation of the prosthesis but caused some pressure necrosis.

FIG. 5. Dog 22. A cannula is passed through the perforation to the oesophagus at the site of the cranial ivalon ring. None of the ivalon rings was healed in place 220 days after operation.

FIG. 6. (a) Bronchography of dog 26 one year after operation showed a good result with widely patent lumen and the rings in satisfactory position after removal of the prosthesis. (b) After removal of the inner tube by bronchoscopy a fistula to the oesophagus developed, causing death. (c) After one year the teflon rings were still not completely held in place but they were in direct communication with the lumen once the inner tube had been removed.
Of the seven survivors, three dogs had late perforations of the oesophagus. In dogs 22 and 24 the perforation was caused by the ivalon ring reinforced by stainless steel wire, after 220 days (Fig. 5) and 194 days respectively. In dog 26 the oesophagus was perforated when the inner tube was removed by bronchoscopy 360 days after operation (Fig. 6).

Three other dogs had the inner tube removed by bronchoscopy, and in all three the ivalon rings were thereafter found to be loose in the lumen of the trachea. Dog 21 was sacrificed 408 days after operation (Fig. 7) and on the same day as the bronchoscopic removal of the inner tube and two rings. The third ring caused tracheal obstruction with dyspnoea. Dogs 23 and 27 died 15 days after removal of the inner tube. The ivalon rings were found in an abscess in the trachea (Fig. 8). The remaining dog (25) died from pneumonia 123 days after operation. The inner tube was in the correct position.

Conclusion An inner tube of silastic has a tendency to become displaced. Ivalon rings reinforced by stainless steel are not suitable. In two cases the rings perforated the oesophagus. When the inner tube was removed the rings were not healed in place but were usually found to be loose in the trachea and obstructing the airway. They did not prevent stricture formation.

GROUP B The prosthesis used was an inner tube of teflon and ivalon rings. One dog (28) with an inner tube of teflon (Fig. 1 D) had a bronchoscopy performed after 282 days. The teflon tube could not be removed in spite of several attempts being made. The dog died 20 days later. Necropsy showed that the lungs were in good condition and the ivalon rings were partly healed in place (Fig. 9).

FIG. 7. Metaplastic squamous epithelium in the margin of the reconstructed part.

FIG. 8. Dog 27. (a) Bronchography 313 days after operation showed a wide open lumen and the ivalon rings in good position after removal of the prosthesis. (b) This dog died 15 days after removal of the inner tube. All the inner rings were lying loose in the trachea causing airway obstruction.
FIG. 9. Dog 28. (a) Bronchography shows a wide open tracheal prosthesis but the ivalon rings are too close together in the centre. (b) Necropsy specimen. The ivalon rings are only partly held in place after 282 days. (c) A newly formed bridge of cartilage between two tracheal rings close to the resectional margin. Weigert's haematoxylin and van Gieson, ×33.

FIG. 10. Necropsy specimen from dog 12 sacrificed after 121 days. The teflon rings are only partly embedded in tissue.
Conclusion A teflon tube with big extensions for suturing may firmly heal in place and it may be impossible to remove it owing to the extensions. The ivalon rings were not completely covered or invaded by fibrous tissue.

GROUP C The prosthesis used was an inner tube of silastic with teflon rings. Of eight dogs, three died, two from pneumonia, four and five days after operation, and one from the operation.

Five dogs survived. Of those, one (12) died 121 days after operation as a result of another operation performed by another team. The silastic tube was found to be in place but the teflon rings were not completely embedded in tissue (Fig. 10). The second dog (32) died from distemper after 109 days; the tube was in place. Three dogs (13, 15, and 16) are still alive and well after six months.

GROUP D The prosthesis used was an inner tube of teflon with rings of teflon. Eighteen dogs were operated upon. Thirteen dogs died in the early post-operative period due to suture insufficiency (5), bronchopneumonia (3), distemper (3), and from an unknown cause (2).

Of the five survivors, one (14) died from distemper after 76 days (Fig. 11). Another (20) died after 226 days; one ring was well encapsulated. One dog (19) died 622 days after operation, and after that the inner prosthesis was removed. Two dogs (17 and 47) died after 505 days and 1408 days (3 years 11 months) respectively. Necropsy showed that the teflon rings were well embedded (Figs. 12 and 13).

Conclusion Attempts to remove the teflon tubes with sewing extensions have not been successful. On the other hand, it has been proved that fine rings of teflon can be embedded in the pedicled pericardium and mediastinal tissue to form a rigid tube with a smooth inner surface.

PATHOLOGY The degree of the changes in the reconstructed part of the trachea and adjacent tissues can be seen in Table I. Only a few added comments are necessary.

The inflammation in the reconstructed part of the trachea often had the appearance of granulation tissue. In some cases a capillary-rich granulation tissue could be seen protruding fungous-like into the tracheal lumen.

The connective tissue was in many cases hyalinized and sclerotic with the appearance of scar tissue.

Foreign body reaction could be seen around the suture or prosthesis material. Only a few foreign body giant cells have usually been observed.

Cartilage proliferation of a rather high degree was seen in several cases close to the anastomosis. Sometimes there was a proliferation of the outer side of the cartilage rings. In some cases there was a proliferation between two rings connecting them with each other, and in others small cartilage fragments were seen in the connective tissue in the vicinity of the cartilage rings which appeared to be newly formed. The newly formed cartilage rings were often fibrous.

Epithelial regeneration was observed in only a few cases within areas in the middle of the reconstructed part. More often, endothelial-like cells were seen on the surface of the sclerotic connective tissue. In such cases inflammation in the connective tissue was only slight, if present at all. The origin of these cells is obscure. They probably originate from fibroblasts in the granulation tissue or from fibroblasts or fibrocytes of the grafted tissue. There is no real proof that these cells are regenerating epithelial cells.

DISCUSSION

From the results summarized in Table II it is evident that teflon was tolerated as an inner tube better than silastic, and it functioned well for four years in two dogs. Furthermore, tubes of silastic did not prevent pressure necrosis with oesophageal perforations. A teflon tube has thinner walls and
FIG. 12. Dog 47. (a) The teflon inner tube was found to be in place after 505 days, and all the teflon rings were well embedded. (b) After removal of the inner tube a smooth surface was found on the newly created trachea, well supported by the embedded teflon rings. (c) Endothelial-like cells in the surface of the reconstructed part. Haematoxylin and eosin, ×900. (d) Connective tissue surrounding a gap below a teflon ring with two small rests of teflon and foreign body granuloma. Weigert's haematoxylin and van Gieson, ×560.
FIG. 13. Dog 17. (a) The inner teflon tube was left in place for 3 years 11 months. (b) The teflon rings were well embedded, and a rigid, newly formed trachea with a smooth inner surface was found. In this dog there was no sewing ridge in the teflon tube, and there was a smooth wall between the newly formed and the old trachea. (c) A teflon ring with a connective tissue capsule in the old tracheal wall. Weigert's haematoxylin and van Gieson, × 35. (d) The margin of a teflon ring with slight inflammatory reaction in the surrounding connective tissue. Weigert's haematoxylin and van Gieson, × 360. (e) Regenerating epithelial cells in the surface of the reconstructed part. Haematoxylin and eosin, × 900.

a greater lumen is therefore obtained. Mucus did not stick to either material. Ivalon was found to be an unsuitable material for the rings as none became embedded and healed in place. The teflon rings, however, were embedded in the newly formed trachea and assumed an appearance and function similar to those of the tracheal cartilage. The fact that teflon tubes have functioned well and also that they have been well tolerated for up to four years is encouraging. The technique of suturing with the tubes inside the end of the trachea prevented suture granuloma with stricture at the anastomosis. In human beings, a pedicled flap of pleura could be used instead of a pedicled

### TABLE I

<table>
<thead>
<tr>
<th>Group</th>
<th>Dog No.</th>
<th>Inflammation with or without Tissue</th>
<th>Sclerotic Connective Tissue</th>
<th>Foreign Body Giant Cells</th>
<th>Cartilage Proliferation</th>
<th>Prosthesis or Tracheal Cartilage in Cartilage</th>
<th>Prosthesis Material or Such Material</th>
<th>Inflammation of Connective Tissue</th>
<th>Foreign Body Giant Cells</th>
<th>Ulceration</th>
<th>Tracheal Mucous Membrane</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Central Part</td>
</tr>
<tr>
<td>Control</td>
<td>4</td>
<td>++</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td></td>
<td>11</td>
<td>+(+)+</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>(+)</td>
<td>0</td>
<td>(+)</td>
<td>0</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>A</td>
<td>21</td>
<td>++++++</td>
<td>+</td>
<td>0</td>
<td>(+)</td>
<td>+++</td>
<td>++</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td></td>
<td>22</td>
<td>++++++</td>
<td>+</td>
<td>0</td>
<td>(+)</td>
<td>+++</td>
<td>++</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td></td>
<td>23</td>
<td>++++++</td>
<td>+</td>
<td>0</td>
<td>(+)</td>
<td>+++</td>
<td>++</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td></td>
<td>24</td>
<td>++++++</td>
<td>+</td>
<td>0</td>
<td>(+)</td>
<td>+++</td>
<td>++</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td></td>
<td>25</td>
<td>++++++</td>
<td>+</td>
<td>0</td>
<td>(+)</td>
<td>+++</td>
<td>++</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td></td>
<td>26</td>
<td>++++++</td>
<td>+</td>
<td>0</td>
<td>(+)</td>
<td>+++</td>
<td>++</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>B</td>
<td>28</td>
<td>++</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>0</td>
<td>(+)</td>
<td>+</td>
</tr>
<tr>
<td>C</td>
<td>12</td>
<td>++++</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td></td>
<td>32</td>
<td>++++</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>D</td>
<td>14</td>
<td>+++++</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td></td>
<td>17</td>
<td>+++++</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>0</td>
<td>+</td>
<td>+</td>
</tr>
</tbody>
</table>

*= in limited areas of higher degree; †= suture granuloma.

### TABLE II

<table>
<thead>
<tr>
<th>Group</th>
<th>No. of Dogs</th>
<th>Immediate Mortality</th>
<th>Survivors</th>
<th>Perforation</th>
<th>Longest Survival</th>
<th>Fixation</th>
<th>Function</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>5</td>
<td>0</td>
<td>5</td>
<td>0</td>
<td>183 days</td>
<td>Bad</td>
<td>Stenosis</td>
</tr>
<tr>
<td>A</td>
<td>24</td>
<td>1</td>
<td>17</td>
<td>7</td>
<td>408 days</td>
<td>Bad</td>
<td>Excellent</td>
</tr>
<tr>
<td>B</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>282 days</td>
<td>Good</td>
<td>Stenosis when inner tube removed</td>
</tr>
<tr>
<td>C</td>
<td>8</td>
<td>3</td>
<td>5</td>
<td>0</td>
<td>6 months</td>
<td>Good</td>
<td>Good</td>
</tr>
<tr>
<td>D</td>
<td>18</td>
<td>13</td>
<td>5</td>
<td>0</td>
<td>4 years</td>
<td>Good</td>
<td>Good</td>
</tr>
</tbody>
</table>

2g**
Björk, Crafoord, 368

Burnett, Beall, A. J.

Aronstam, Brewer, removed.

of pericardium. The shape of the teflon prosthesis used permitted excellent fixation. However, the teflon extension injured the surrounding tissue and rings when the inner tube was removed. The function of the outer tube as a trachea with supporting teflon rings could not be tested. The new principle of creating the trachea as an outer tube of thin supporting teflon rings placed at short intervals over a pedicled flap of pericardium has proved to be possible. The rings are embedded in tissue during the time the inner tube functions as a trachea. The results seem promising in this very difficult field of tracheal reconstruction.

SUMMARY

A new principle for tracheal reconstruction is presented. Thin teflon rings are embedded in the mediastinal tissue over a pedicled flap of pericardium placed over an inner tube of teflon for immediate function. The observations from dog experiments carried on over a period of four years are reported.

REFERENCES


A New Method for Tracheal Reconstruction

H. Aletras, V. O. Björk, G. Hultquist and L. Rodriguez

Thorax 1964 19: 358-368
doi: 10.1136/thx.19.4.358

Updated information and services can be found at:
http://thorax.bmj.com/content/19/4/358.citation

These include:

Email alerting service
Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

Notes

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/