

Introduction and objectives New methods of high throughput sequencing provide unparalleled access to the human genome and transcriptome. We hypothesised that next generation DNA sequencing technologies would allow us to identify an elusive novel disease gene for a pulmonary vascular disease inherited as an autosomal dominant trait: The *HHT3* interval on chromosome 5 is predicted by linkage studies to contain a mutation causing pulmonary arteriovenous malformations and hereditary haemorrhagic telangiectasia.^{1,2}

Methods Published expressed sequence tag (EST) databases and tiling array data were used to supplement sequencing analysis of the *HHT3* interval. Sheared, Agilent SureSelect adaptor-ligated genomic DNAs from six related patients and four controls were hybridised to single stranded biotinylated RNA baits. Samples were pooled for multiplexed sequencing on an Illumina HiSeq2000. Sequence data were processed with RTA version 1.7.45, CASAVA Eland pair algorithm, and CASAVA 1.7 demultiplexing algorithms. Validations of sequence variants were performed using conventional PCR and Sanger sequencing.

Results Conventional exon-based sequencing strategies did not identify the *HHT3* causative gene mutation. For individual candidate genes, up to 108 alternatively spliced transcripts per gene were predicted from EST databases. For intergenic regions, tiling array data indicated that up to 44 different transcribed fragments were present in the nucleus and/or cytoplasm of different cell types. For each NextGen sequencing DNA sample, ~8 million reads per sample uniquely mapped to the *HHT3* interval which represents ~1/5,000 of the genome. Using a 2:1 threshold, an average of ~4,000 differences to NCBI36/hg18 were identified in each sample. 113 differences to NCBI36/hg18 were present in all six *HHT3*-affected individuals and absent in all four controls. 60% of novel shared variants were validated by wet lab PCR. Following exclusion in 100 normal chromosomes, and computational predictions of potential function, multiple candidate sequence variants remain.

Conclusions Genomic sequencing capturing intronic sequences yields challenging numbers of sequence variants for wet lab validations, even when multiple replicate chromosomal strategies are employed.

REFERENCES

- 1 Cole et al. *J Med Genet* 2005;42(7):577–582
- 2 Govani et al. *J Angiogen Res* 2010;11(2):15

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ROBO1/4-SLIT2 EXPRESSION IN PULMONARY VASCULAR CELLS: IMPLICATIONS FOR PAH?

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Introduction and objectives Pulmonary artery hypertension (PAH) is associated with inappropriate vascular remodelling and inflammation. Recent studies have shown that vascular cells express the transmembrane roundabout (Robo) proteins, Robo1 and Robo4, and that interaction with a secreted glycoprotein ligand, Slit2, controls cell migratory and inflammatory response. We hypothesise that Robo1, Robo4 and Slit2 are expressed on pulmonary artery (PA) endothelial cells (EC) and smooth muscle cells (SMC). We also hypothesise that Slit2 will modulate PAEC and PSMC migration and inflammatory mediator release.

Methods Real-time-PCR determination of Robo1, Robo4 and Slit2 expression and the house-keeping gene, GAPDH, in PAEC,

PASMC and for comparison, human umbilical vein endothelial cells (HUVEC); following incubation with TNF α (10ng/ml) or Slit2N (10nM) for 2h. Enzyme-linked immunosorbent assay measurements of granulocyte-macrophage-colony stimulating factor (GM-CSF) in supernatants of HUVEC, PAEC or PASMC pre-treated (1h) with Slit2N, followed by TNF α (17h). Migration assays (PAEC or PASMC) towards serum-containing medium (0.05 and 0.02%, respectively), for 4h with/ without Slit2N.

Results Basal mRNA expression of Robo1, Robo4 and Slit2 was detected in PAEC, PASMC and HUVEC (n = 3–4). Slit2N (2h) significantly (p < 0.05, n = 3) decreased Robo4 and Slit2 mRNA expression, but not Robo1, by 35% in PAEC; and had no effect on HUVEC or PASMC. TNF α had no significant effects on Robo1, Robo4 or Slit2, regardless of cell type. Despite a small (23%), but significant (p < 0.05) reduction of GM-CSF release from TNF α -activated HUVEC (n = 7), no similar effects were seen in PAEC or PASMC (n = 3). Moreover, whilst PAEC or PASMC migration to serum-containing medium increased (2.7- and 5.3-fold, respectively), co-incubation with Slit2 had no significant effect.

Conclusion The novel discovery of Robo1, Robo4 and Slit2 mRNA in PAEC and PASMC; and that Slit2 down-regulated Robo4 and Slit2 in PAEC, but not PASMC/HUVEC, might suggest negative feedback on the Robo4-Slit2 axis unique to PAEC. That neither PAEC nor PASMC responded to Slit2 in functional assays could reflect limitations in experimental assays. However, down-regulation of Robo4-Slit2 in PAEC might also explain lack of effect on GM-CSF release, when compared with HUVEC. Further studies to better delineate the role of the Robo-Slit2 pathway in PAH are required.

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THE ROLE OF DIFFERENTIAL TNFR SIGNALLING IN MAINTENANCE OF ALVEOLAR EPITHELIAL HOMEOSTASIS

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Alveolar epithelial activation and disruption of the alveolar epithelial barrier promote recruitment of neutrophils into the alveolar space and cause alveolar oedema respectively thereby playing key roles in the pathogenesis of Acute Respiratory Distress Syndrome (ARDS). Tumour necrosis factor alpha (TNF) is an early mediator of inflammation in ARDS. TNF signals through two cell surface receptors, TNFR1 and TNFR2 initiating distinct signalling pathways and cellular responses.

Using a novel, highly selective TNFR1 domain antibody (dAbTM), a dummy dAb and the dual TNFR antagonist AdalimumabTM, we investigated the role of differential TNFR signalling on human pulmonary alveolar epithelial cell (human alveolar type 2 cells and A549 cell line) activation, permeability and repair.

Human alveolar epithelial type 2 cells (haT2) expressed both TNFR, whilst A549 cells only expressed TNFR1. TNFR1 signalling mediated release of the neutrophil chemokines IL-8 and GMCSF as well as IL-6 in haT2 (pin vitro scratch model of epithelial (haT2) wound repair.

TNFR1 signalling induced pro-inflammatory cytokine expression from alveolar epithelial cells and mediated increased epithelial permeability. TNFR1 induced permeability did not appear to